

Elovanoids downregulate canonical SARS-CoV-2 cell-entry mediators and enhance protective signaling in human alveolar cells

Jorgelina Calandria

Louisiana State University Health New Orleans

Surjyadipta Bhattacharjee

Louisiana State University Health New Orleans

Marie-Audrey I. Kautzmann

Louisiana State University Health New Orleans

Aram Asatryan

Louisiana State University Health New Orleans

William C. Gordon

Louisiana State University Health New Orleans

Khanh V. Do

Louisiana State University Health New Orleans

Bokkyoo Jun

Louisiana State University Health New Orleans

Pranab K. Mukherjee

Louisiana State University Health New Orleans

Nicos A. Petasis

University of Southern California

Nicolas G. Bazan (✉ nbazan@lsuhsc.edu)

Louisiana State University Health New Orleans <https://orcid.org/0000-0002-9243-5444>

Research Article

Keywords: Lipid mediators, SARS-CoV-2, ACE2 receptor, Spike protein, alveolar cells, receptor-binding domain, lung

Posted Date: September 11th, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-75121/v1>

License:  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Version of Record: A version of this preprint was published at Scientific Reports on June 10th, 2021. See the published version at <https://doi.org/10.1038/s41598-021-91794-z>.

Abstract

The pro-homeostatic lipid mediators elovonoids (ELVs) attenuate cell binding and entrance of the SARS-CoV-2 receptor-binding domain (RBD) in human primary alveoli cells in culture. We uncovered that very-long-chain polyunsaturated fatty acid precursors (VLC-PUFA,_{n-3}) activate ELV biosynthesis in lung cells. Both ELVs and their precursors reduce the binding to RBD. ELVs downregulate angiotensin-converting enzyme 2 (ACE2) and enhance the expression of a set of protective proteins hindering cell surface virus binding and upregulating defensive proteins against lung damage. These findings open avenues for potential preventive and disease-modifiable therapeutic approaches for COVID-19.

Introduction

The high transmissibility of Severe Acute Respiratory Syndrome–coronavirus 2 (SARS-CoV–2) is due, at least in part, to infectivity in lung type II alveolar cells¹ SARS-CoV–2 triggers a wide range of disease phenotypes with severe acute respiratory distress syndrome (ARDS), including interstitial pneumonia² and viral sepsis³. Here, we tested if the pro-homeostatic lipid mediators, the elovonoids (ELVs)^{4–7}, would block the entrance of the spike (S) protein receptor-binding domain (RBD) that would prompt a protective response against SARS-CoV–2 infection.

Lung alveoli viral attachment through the S protein RBD to ACE2 is followed by proteolytic activation for fusion and viral cell entry^{8–10}. We use human alveolar primary cell cultures (Fig. 1a, and Extended Data Fig. 1). Most of the cells are type II (oil red, specific marker, Fig. 1a, right panel, and Extended Data Fig. 1, bottom panels) and positive to Foxj1, HT2–280 antigen, and β -tubulin IV (Fig. 1a, right panel; Fig. 1g,h and Extended Data Fig. 1). Pneumocytes type II are also mobile, showing lamellae or filopodia positive to HT2–280, a specific type II (Fig. 1b). We exposed these cells to RBD (from S protein)-Alexa 594 for 24 hours. In parallel, we used Nucleocapsid protein N as a specificity control of RBD internalization. In 3D reconstructions of Z-stack images (Imaris software, Bitplane, UK), the lipophilic staining (cell mask) shows a dense membrane above the nuclear zone that is localized close to oil red (Fig. 1a,c *i-viii*).. A below view (Fig. 1c *vi*) shows that the RBD protein signal (red) passes through the membrane (white) to the intracellular space surrounding the nucleus (blue), and can also be seen in the above view (Fig. 1c *vii,viii*).. Herein, we demonstrate for the first time that RBD was shown to be internalized in SARS-CoV–2 since previous work have shown the same for SARS-CoV–1¹¹. When IL1 β or TNF α was added, the internalized RBD signal was increased (Fig. 1d). In addition, Nucleocapsid (N) protein, a structural viral protein not involved in ACE2 and SARS-CoV–2 interaction¹², is at the same level as the control with no protein added that accounted for autofluorescence. RBD was internalized at higher rates than N. In digitalized images, plotted vs. Z-axis in a Z-stack shows the differential position of the N protein versus the RBD with respect to the membrane level (Fig. 1c *i-v,ix-xi* and Fig. 1d). This observation documents that the N protein remains on the membrane and the extracellular side while the RBD spans intracellularly, passing through to the cytoplasm and demonstrating that RBD internalization is specific and dependent on IL1 β and TNF α (Fig. 1d,e).

RBD-Alexa 594 internalization was decreased +/- IL1 β + TNF α when ELV-N32 and ELV-N34 were added (Fig. 1f, upper panel). In addition, acetylenic ELV-N32 or ELV-N34 (Extended Data Fig. 2) showed a steep decrease in RBD protein internalization +/- IL1 β (Fig. 1f, lower panel). Moreover, the addition of the ELVs precursors 32:6 or 34:6 reduces RBD located below the membrane, suggesting that the pneumocytes convert these precursors into ELVs and thus prevent RBD internalization (Fig. 1f, upper panel).

The reduction in RBD internalization is partially due to a decrease in ACE2 since acetylenic ELV-N32 or ELV-N34 decreases ACE2 mRNA and methyl-ester elovonoids also reduce the protein content in pneumocytes type II (Fig. 1g, plot, and Fig. 1i). In addition, ELV-N32 decreased TMPRSS2 expression (Fig. 1h) in the presence of IL1 β (Fig. 1h, plot).

Since ELVs stimulate protective proteins expression in cells confronted with uncompensated oxidative stress⁵⁻⁷, we next explored if these lipids under conditions that downregulate canonical SARS-CoV-2 cell-entry mediators in pneumocytes will also activate protective proteins synthesis. We found that ELV heightens the expression of Sirtuin 1 (Fig. 1j), RNF146 (Extended Data Fig. 3a,b), PHB, Bcl-XI, and Bcl2 (Extended Data Fig. 3c-e). These proteins are involved in pro-homeostatic cellular functions. Sirtuin 1 (Silent information regulator factor 2-related enzyme 1) is a NAD(+)-dependent deacetylase of histone and non-histone proteins and transcription factors, and its regulatory functions target inflammation, aging, mitochondrial biogenesis, and cellular senescence¹³. RNF146 is an E3 ubiquitin-protein ligase that degrades polyubiquitinated proteins, thus protecting cells from Parthanatos cell death¹⁴. PHB (prohibitin type I) functions comprise scaffolding mitochondrial protein, adaptor in membrane signaling, transcriptional co-regulator, and neuroprotection⁶. Bcl-XL and Bcl2 downregulate apoptosis and inflammasome formation¹⁵. Our data suggest that, in addition to halting the entrance of the RBD, ELVs in the lung curb cell-damaging/apoptotic events and thus sustains homeostasis by counteracting inflammation over-activation by the formation of protective proteins.

An evolving question prompted by our data is whether alveolar cells in culture can synthesize ELVs. Thus, we incubated human alveolar cells with the precursors VLC-PUFAs (32:6 or 34:6) and then analyzed the products by LC-MS/MS. Interestingly, we found that ELVs are in fact, formed. ELV-N32 was synthesized where the precursor 32:6 was added and not in cells exposed to 34:6. Inversely, ELV-N34 was found in the cultures where 34:6 was added and not in cells exposed to 32:6 (Fig. 2a,c). These results demonstrate that alveolar cells are endowed with pathways for the biosynthesis of ELV-N32 and ELV-N34 (Fig. 2b). We show MS fragmentation for stable derivatives of intermediaries (Fig. 2a-c) as well as of ELVs themselves (Fig. 2a). Moreover, we uncovered that ELVs were actively released from cells to the incubation media, indicating that they act both as autocrine and paracrine mediators.

Our findings contribute to broadening our understanding of the duality of ACE2 in lung function and diseases. In health, ACE2 fosters lung homeostasis by generating Ang-(1-7) and enhancing host defense that would counteract ACE2 virus-induced downregulation of proinflammatory signaling. Herein, we show that ELVs uncover another participant when RBD of the S protein binds to ACE2 and enters alveolar cells in culture. The ELVs are likely part of a fast and coordinated pro-homeostatic inflammatory

downregulatory response. To be tested in the future is the prediction that delayed ELV-mediated protective responses would lead to severe lung and systemic inflammation. So direct virus triggered cell damage is critical, but also the activation of the induction of protective proteins. Is diet engaged in building precursors of ELVs in the lung? Diet has been shown to affect ACE2 expression¹⁶ and the supply to build ELV precursors^{7,17}. This may contribute to explaining why some patients develop hyper-inflammatory/immune responses and severe disease, but others experience mild or even asymptomatic COVID-19. Questions that remain to be addressed include whether the expression of the protective proteins identified here in the alveoli are activated all at once? Are they coordinated with adaptive immune responses to limit virus spread? Are enzymes for ELV synthesis under tight transcriptional control so that the mediators are expressed at appropriate times and/or levels? To our knowledge, ELVs are the first protective mediators to be identified in the human alveoli confronted with the RBD of the S protein.

Additional research will be needed to elucidate the molecular mechanisms of ACE2 downregulation. Also, the use of the entire S protein instead of the RBD, as in our present study, will provide the connection between cell attachment and cell entrance, as affected by ELVs and VLC-PUFAs, since proteases expression is correlated with ACE2 downregulation. Moreover, the use of the intact virus would offer a direct demonstration of the significance of ELVs. Since the SARS-CoV-2 affects nasal mucosa, GI, the eye, and the nervous system exploring the protective potential of ELVs in other cell types would further expand the scope of our observations beyond the lung. Our results provide a foundation for future research and offer specific mediators for interventions to modify disease risk, progression, and protection of the lung from COVID-19 or other pathologies.

Methods

Primary cultures of human alveolar cells and assessment of protein internalization.

We have used primary cell cultures of human alveoli, which consist of a mixture of ciliated cells, club cells, type I pneumocytes, and type II pneumocytes (PromoCell, HSAEpC). We have characterized the histology and immunocytochemistry in these primary cultures (Fig. 1a,b,g,h). We performed all the experiments in 48 wells with passage 4 cells seeded at 15000 cells per cm² density. The cells were incubated to confluency and maintained in the proprietary Medium provided by Promocell with the addition of Pen/Strep and exposed to 0.5 g of tagged protein per well for 24 hours in the presence or absence of 10 ng/ml IL1 β (PeProTech Inc., Rocky Hill, NJ Cat# 200-01B) and/or 10 ng/ml TNF α (Cell Sciences Inc., Newburyport MA. Cat# CRH520B). After this period, cells were incubated 10 min with 1/1000 cell mask (Thermo Scientific cat# C37608) medium and fixed using PFA 4%. After fixation, nuclei were stained with 10 g/ml Hoechst 33342 (Thermo Scientific cat#H3570). To characterize the cell types in culture after fixation, we performed immunocytochemistry using the following primary antibodies: HT1-53, a marker of pneumocytes type I, and HT2-280 (Terrace Biotech cat#HT1-53 and HT2-280);

Foxj1 (Santa Cruz Biotech, sc-53139) and β -Tubulin IV marker of pneumocytes type II (Abcam cat# ab179509). ACE2 (Santa Cruz Biotech, sc-390851) and TMPRSS2 (Abcam cat #Ab109131) were used for Immunostain the two mentioned proteins in cell culture.

ACE2 Protein abundance using Jess technology.

The western assay was performed using a Jess Protein Simple system (San Jose, CA, USA) following the manufacturer's protocol. Briefly, samples were lysed with RIPA buffer containing a protease inhibitor cocktail (Sigma, Cat. P8340). Soluble protein concentration was determined by BCA assay (Thermo Fisher Scientific, Cat. 23225) and 0.4 μ g used/reaction. Samples were heated at 95 °C/5 min, and 3 μ L of each sample were loaded. The 12–230 kDa cartridge (Protein Simple–#SM-W004) was used. Primary antibodies were diluted in antibody diluent 2 buffer (Protein Simple, #042–203), and the working solution of secondary antibodies was provided by the company (Protein Simple, #042–206). For data analysis, the area of spectra that matched the molecular weight of the target protein was used (Fig. 1j). We used the anti-ACE2 antibody from Abcam (cat# ab108252) in a concentration of 1 g/ml. The standardization was performed using total protein stain and using an anti-GAPDH antibody from Santa Cruz Biotech (cat# Sc-25778).

Quantification of RNF-146.

Western blot was performed from human primary alveolar cells, as described in Calandria *et al.*, 2015 (18). Briefly, cell lysates were produced using RIPA buffer supplemented with protease inhibitor cocktail (Sigma, cat# P8340.St Louis MO). Total protein (30 mg) was mixed with Laemmli buffer containing DTT and loaded in Novex 4–12% precast gels and ran in X-Cell running system at 120V for 1.5 hours. The transference was performed using the Trans Blot Turbo dry transferring system (Bio-Rad, Hercules CA) on low fluorescent background PVDF membranes (GE Healthcare, Piscataway NJ). Membranes were incubated with the corresponding primary antibodies overnight. Primary antibodies used RNF-146 (UC Davis/NIH-Neuromab Lab Facility, cat #75–233) and GAPDH (Santa Cruz Biotech Cat# sc-47724). After this period, the membranes were incubated with fluorescent-tagged secondary antibodies (GE healthcare, cat# PA45011) for 1 hour and imaged. Data was acquired using ChemiDoc MP (Biorad). Densitometric analysis was performed using ImageLab 6.0.1. (Biorad).

Preparation of tagged RBD and N nucleocapsid proteins.

We have obtained the Recombinant SARS-CoV-2, S1 Subunit Protein (RBD), and Recombinant SARS-CoV-2 Nucleocapsid Protein from Raybiotech (Peachtree Corners GA, cat# 230–30162–1000 and 230–30164–500 correspondingly). The proteins were labeled using Alexa Fluor™ 546 Protein Labeling Kit from Thermo Scientific (cat# A10237) following manufacturer directions except for the RBD that was purified from the dye with Amicon-Ultra 10K cutoff filters (Merck, Millipore cat#UFC201024) instead of the

column provided by the kit has a restrictive MW of 50KD (the recombinant RBD protein was 25KDa). The recovery of the protein and labeling efficiency was measured using nanodrop and was about 80% recovery and 0.02 dye molecules per aminoacid. We added 0.5 g per well of protein.

Quantitation of cell surface binding and internalization of tag RBD of the viral S protein.

0.5 g of Alexa 594-conjugated RBD domain belonging to the SARS-CoV-2 virus Spike protein (Raybiotech. Cat. 230-30162-1000) was incubated with human alveolar cells for 24 hours. After this period, Cell Mask (Thermofisher Scientific, cat#C37608) was added to a final concentration of 1 in 1000 and left in the incubator for 10 minutes. The cells were then fixed with 4% Paraformaldehyde in PBS, washed three times, and Nucblue (Thermofisher Scientific cat# R37606) was added for nuclear staining. The images were taken using an Olympus FluoView 3000 laser confocal microscope as z-stacks with a fixed step size of 1.6 μ m. The zero was registered for each well specifying -30 and 30 μ m as the lower and upper limit in the Z-axes. Using the Z-drift compensation system, 9 blind positions were set up per well, and the image acquisition was performed automatically. The pictures were processed using Imaris 9.5.1 software (Bitplane) to render the 3D image and assess the position of the different surfaces (elements rendered with the surface function) along the Z-axes. The portion of the tagged protein that was internalized was calculated from the total sum of intensity that crossed the membrane inner limit to the intracellular space in the Z-axes. The proportion was calculated in percentage of internalization.

Real-Time PCR using Taqman probes and SYBR green assay.

cDNA was produced using 1 g of total RNA extracted by RNeasy (Qiagen, Hilden Germany, cat# 74104). The first strand of cDNA was produced using iScript™ Reverse Transcription Supermix for RT-qPCR (BioRad cat # 1708840). The quantification of Sirt 1, RNF-146, Bcl2, Bcl-xl was performed using SYBRgreen assay with primers designed in house (Table 1) using SsoAdvanced Universal SYBR Green Supermix (Biorad cat#1725270). The quantification of ACE2 and TMPRSS2 mRNA was performed using Taqman probes (Biorad cat# qHsaCEP0051563 & qHsaCIP0028919 respectively) labeled with FAM and standardized using PGK1 probe labeled with HEX (Biorad cat# qHsaCEP0050174).

Table 1

Gene name	Accession number	Primer	Seq 5' to 3'
Sirtuin 1	NM_012238.5	SIRT1F	TGCTGGCCTAATAGAGTGGCA
		SIRT1R	CTCAGCGCCATGGAAAATGT
Prohibitin 1	NM_001281496.1	PHB1F	AAGGCTCGTTTCTGGGCATCTC
		PHB1R	CTGGACCCTCTCACACGCA
Bcl2l1/Bcl-XL	Z23115.1	Bcl-XLF	AGGCGGATTTGAATCTCTTTCTCT
		Bcl-XLR	CCCGGTTGCTCTGAGACATT
RNF-146/Iduna	NM_001242844.1	RNF146F	ATTCCCGAGGATTTCTTGACA
		RNF146R	GTCATCGTACTGCCACCA
Bcl2	NM_000633.3	Bcl2F	GATGTGATGCCTCTGCGAAG
		Bcl2R	CATGCTGATGTCTCTGGAATCT

Declarations

Acknowledgements

The authors are grateful to an Institutional Grant from the School of Medicine, LSUHSC (to N.G.B.).

Author contributions

N.G.B. and J.C. designed the experiments and wrote the manuscript. J.C. conducted the Imaris analysis and blind coding of samples, S.B. and J.C. prepared the lipids and incubated cells. S.B. maintained human alveolar cell cultures. A.A. performed JESS-Western Blots. J.C. and M-A.I.K. prepared Alexa 594-conjugated RBD fragment of SARS-CoV-2 Spike protein. M-A.I.K. took images by confocal microscopy. K.V.D. perform RNA extraction, dd-PCR. N.G.B. and J.C. broke the code and analyzed the results of the experiments. B.J. did LC-MS/MS. P.K.M. performed Western Blot analysis. N.A.P. provided synthetic ELV-N32 and ELV-N34. W.C.G. perform the immunohistology.

Competing Interest Declaration

The authors declare no competing interests.

References

1. Tian S, Xiong Y, Liu H, Niu L, Guo J, Liao M, *et al.* Pathological study of the 2019 novel coronavirus disease (COVID-19) through postmortem core biopsies. *Mod Pathol* 2020;**33**:1007–14. <https://doi.org/10.1038/s41379-020-0536-x>.
2. Raghu G, Wilson KC. COVID-19 interstitial pneumonia: monitoring the clinical course in survivors. *The Lancet Respiratory Medicine* 2020;**0**: [https://doi.org/10.1016/S2213-2600\(20\)30349-0](https://doi.org/10.1016/S2213-2600(20)30349-0).
3. Li H, Liu L, Zhang D, Xu J, Dai H, Tang N, *et al.* SARS-CoV-2 and viral sepsis: observations and hypotheses. *Lancet* 2020;**395**:1517–20. [https://doi.org/10.1016/S0140-6736\(20\)30920-X](https://doi.org/10.1016/S0140-6736(20)30920-X).
4. Bhattacharjee S, Jun B, Belayev L, Heap J, Kautzmann M-A, Obenaus A, *et al.* Elovans are a novel class of homeostatic lipid mediators that protect neural cell integrity upon injury. *Sci Adv* 2017;**3**:e1700735. <https://doi.org/10.1126/sciadv.1700735>.
5. Bazan NG. Docosanoids and elovans from omega-3 fatty acids are pro-homeostatic modulators of inflammatory responses, cell damage and neuroprotection. *Mol Aspects Med* 2018;**64**:18–33. <https://doi.org/10.1016/j.mam.2018.09.003>.
6. Jun B, Mukherjee PK, Asatryan A, Kautzmann M-A, Heap J, Gordon WC, *et al.* Elovans are novel cell-specific lipid mediators necessary for neuroprotective signaling for photoreceptor cell integrity. *Sci Rep* 2017;**7**:5279. <https://doi.org/10.1038/s41598-017-05433-7>.
7. Do KV, Kautzmann M-A, Jun B, Gordon WC, Nshimiyimana R, Yang R, *et al.* Elovans counteract oligomeric β -amyloid-induced gene expression and protect photoreceptors. *Proc Natl Acad Sci USA* 2019:201912959. <https://doi.org/10.1073/pnas.1912959116>.
8. Tang T, Bidon M, Jaimes JA, Whittaker GR, Daniel S. Coronavirus membrane fusion mechanism offers a potential target for antiviral development. *Antiviral Res* 2020;**178**:104792. <https://doi.org/10.1016/j.antiviral.2020.104792>.
9. Andersen KG, Rambaut A, Lipkin WI, Holmes EC, Garry RF. The proximal origin of SARS-CoV-2. *Nat Med* 2020;**26**:450–2. <https://doi.org/10.1038/s41591-020-0820-9>.
10. Hoffmann M, Kleine-Weber H, Schroeder S, Krüger N, Herrler T, Erichsen S, *et al.* SARS-CoV-2 Cell Entry Depends on ACE2 and TMPRSS2 and Is Blocked by a Clinically Proven Protease Inhibitor. *Cell* 2020;**181**:271-280.e8. <https://doi.org/10.1016/j.cell.2020.02.052>.
11. Wang S, Guo F, Liu K, Wang H, Rao S, Yang P, *et al.* Endocytosis of the receptor-binding domain of SARS-CoV spike protein together with virus receptor ACE2. *Virus Res* 2008;**136**:8–15. <https://doi.org/10.1016/j.virusres.2008.03.004>.
12. Zeng W, Liu G, Ma H, Zhao D, Yang Y, Liu M, *et al.* Biochemical characterization of SARS-CoV-2 nucleocapsid protein. *Biochem Biophys Res Commun* 2020;**527**:618–23. <https://doi.org/10.1016/j.bbrc.2020.04.136>.
13. Barnes PJ, Baker J, Donnelly LE. Cellular Senescence as a Mechanism and Target in Chronic Lung Diseases. *Am J Respir Crit Care Med* 2019;**200**:556–64. <https://doi.org/10.1164/rccm.201810-1975TR>.

14. Belayev L, Mukherjee PK, Balaszczuk V, Calandria JM, Obenaus A, Khoutorova L, *et al.* Neuroprotectin D1 upregulates Iduna expression and provides protection in cellular uncompensated oxidative stress and in experimental ischemic stroke. *Cell Death Differ* 2017;**24**:1091–9. <https://doi.org/10.1038/cdd.2017.55>.
15. Bruey J-M, Bruey-Sedano N, Luciano F, Zhai D, Balpai R, Xu C, *et al.* Bcl-2 and Bcl-XL regulate proinflammatory caspase-1 activation by interaction with NALP1. *Cell* 2007;**129**:45–56. <https://doi.org/10.1016/j.cell.2007.01.045>.
16. Mukerjee S, Zhu Y, Zsombok A, Mauvais-Jarvis F, Zhao J, Lazartigues E. Perinatal exposure to Western diet programs autonomic dysfunction in the male offspring. *Cell Mol Neurobiol* 2018;**38**:233–42. <https://doi.org/10.1007/s10571-017-0502-4>.
17. Bazan NG, Molina MF, Gordon WC. Docosahexaenoic acid signalolipidomics in nutrition: significance in aging, neuroinflammation, macular degeneration, Alzheimer's, and other neurodegenerative diseases. *Annu Rev Nutr* 2011;**31**:321–51. <https://doi.org/10.1146/annurev.nutr.012809.104635>.
18. Calandria JM, Asatryan A, Balaszczuk V, Knott EJ, Jun BK, Mukherjee PK, Belayev L, Bazan NG. NPD1-mediated stereoselective regulation of BIRC3 expression through cREL is decisive for neural cell survival. *Cell Death Differ* 2015;**22**:1363–1377. <https://doi.org/10.1038/cdd.2014.233>.

Figures

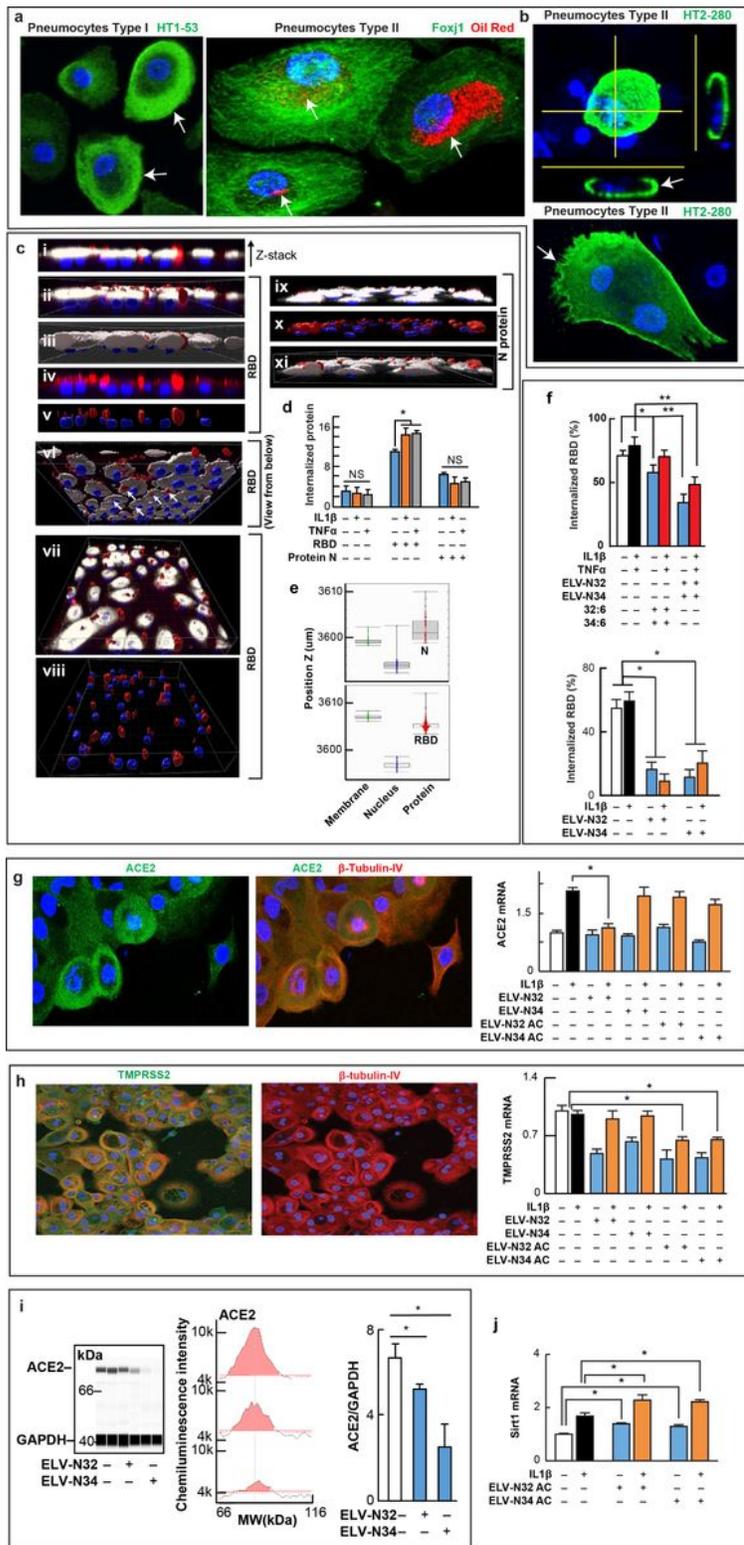


Figure 1

Specific internalization of Spike's protein RBD is reduced by Elovandoids and their precursors in human primary alveolar cells. a, Characterization of Pneumocytes type I (left panel) and type II (right panel) in the alveolar culture. Pneumocytes type I are reactive to HT1-53 (green), and pneumocytes type II are positive for oil red (red), a marker for the type II lipid/lamellar bodies and for ciliated cell marker Foxj1 (green) which is required for cilia formation and is an early marker of epithelial cell differentiation, recovery, and

function. b, Pneumocytes type II stained positive to HT2-280 labeled (green) type II human lung cells. c-e, Differential entrance of S protein vs. N in human alveolar cells in culture. Z-stack shows signal distribution (upper panel), and in mobile cells undergoing cell division (lower panel). c, XZ planes of a Z-stack showing white: membrane, red: RBD tagged with Alexa Fluor 594 and blue nuclei. i through v depict S protein view from XZ plane digitalized of the IMARIS image; vi to viii view from below, above and digitalized showing the internalized protein (white arrows); ix to xi XZ plane depicting the labeled N protein that remains in the surface or within the plasma membrane. Viral Nucleocapsid N protein was tagged with Alexa Fluor-594. XZ plane of a Z-stack image of alveoli culture after 24 h exposure to N protein. d, Quantification of internalized signal when no protein, N and S in the presence or absence of IL1 β or TNF α . e, position of the protein signal in the Z-axes. The plots show digitalized elements position in the Z-axis from 2 representative images of N protein (top panel), and for RBD (bottom panel). The membrane thickness was taken as reference in each picture to determine % of proteins above (free protein), within (bound protein), and below (internalized protein) where green dots are membrane signal, blue dots are nuclei, and red dots are protein. The Y-axis denotes the μ m thickness of the image or Z. The boxes showed the median (middle), and the upper and lower lines depict the Quartile 1 and 3. The whiskers showed the maximum and minimum position adopted in the Z-axes. f, RBD protein internalization decreased by ELV-N32, 34, and their precursors, VLC-PUFAs 32:6 and 34:6. The addition of ELV-N32 and ELV-N34, 2 μ M of the acetylene form together (top panel) or 1 μ M separated methyl ester form (bottom panel), and their precursors 32:6 and 34:6 (2 μ M) showed a decrease in the Alexa-Fluor 594-RBD internalization. g-i, Expression of ACE2 and TMPRSS2 in alveolar cells. Immunocytochemistry of alveolar cells shows signal of ACE2 (g, left panels) and TMPRSS2 (h, left panels) in pneumocytes type II expressing β -tubulin-IV and the mRNA expression for both genes by Taqman qPCR (g-h). i, ACE2 protein quantification (by Jess-capillary Wester Blots-Protein Simple: left panel depicts ACE2 bands at 90 KDa for ACE2 and 40KDa for GAPDH; in the middle panel is shown the area below the pic intensity used to quantified the ACE2 band and the plot shows the quantitation of N=2 samples of alveolar cells +/- ELV-N32 and N34. j, The expression of Sirt1 mRNA, an ELV-N32 and N34 target. Bars depict the mean of N=24 pictures plus/minus standard error. P< 0.05. Membrane (white), S or N Protein (red), and nuclei (blue). Boxes color code all the histograms in the figure.

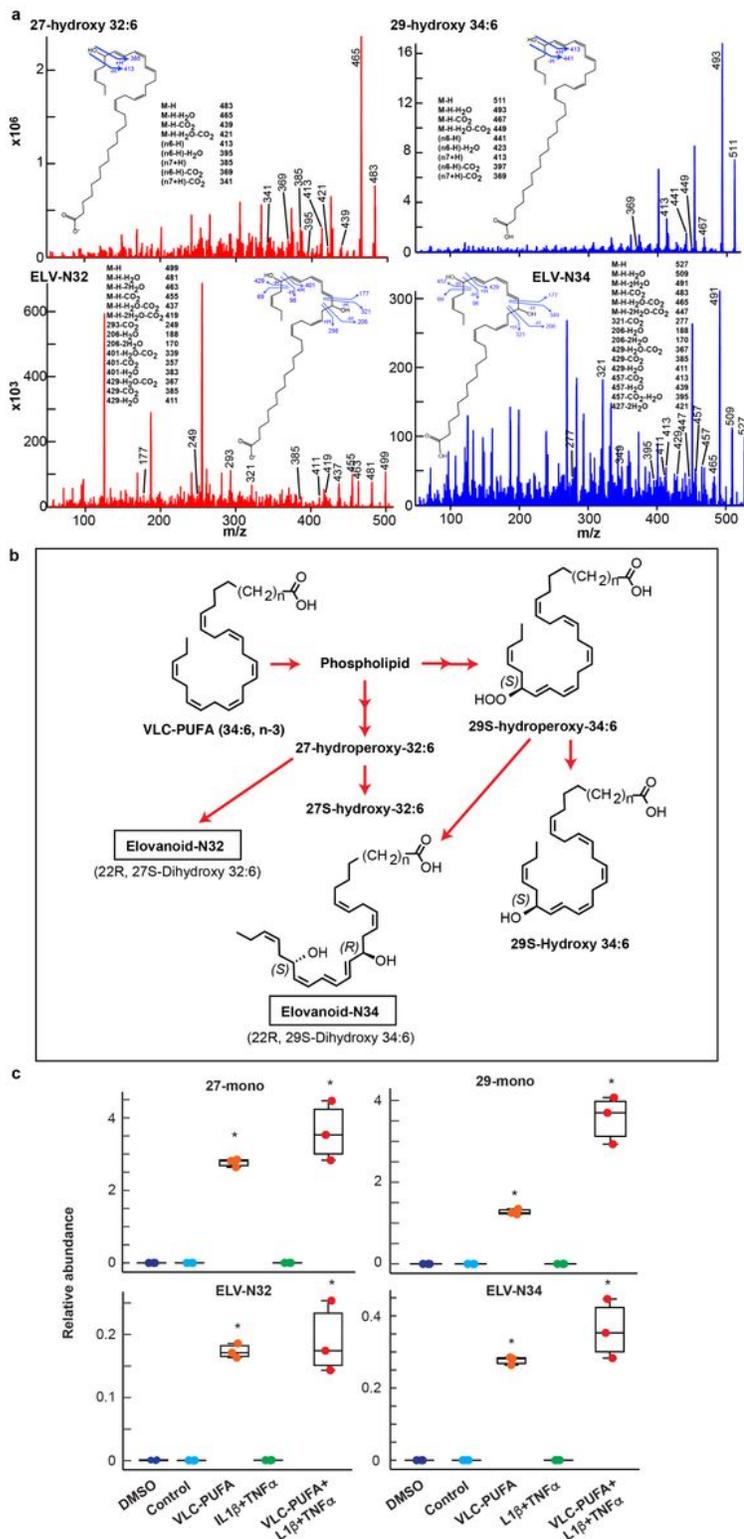


Figure 2

Fate of VLC-PUFAs (C32:6 and C34:6, 2 μ M each) added to human primary alveolar cells cultures towards ELV synthesis. a, 27-monohydroxy-32:6, 29-monohydroxy-34:6, and ELVs are formed. Full fragmentation of 27-monohydroxy-32:6 and 29-monohydroxy-34:6 show matches to their theoretical peaks (top panels). The inserts depict the structures of 27-monohydroxy-32:6 and 29-monohydroxy-34:6 along with the product ions as they are cleaved at a given bond. The complete structures of ELV-N32 and ELV-N34 were

confirmed to be: ELV-N32: (14Z,17Z,20R,21E,23E,25Z,27S,29Z)-20,27-dihydroxydo-triaconta-14,17,21,23,25,29-hexaenoic acid; ELV-N34: (16Z,19Z,22R,23E,25E,27Z,29S,31Z)-22,29-dihydroxytetra-triaconta-16,19,23,25,27, 31-hexaenoic acid (bottom panels). b, Synthesis Pathways for ELV-N32 and ELV-N34 from VLC-PUFAs 34:6, n-3. c, Relative abundance of the elovanoids N32 and N34 (bottom panels) and their intermediates 27 mono-hydroxy and 29 mon-hydroxy (top panels) when the alveolar cells are exposed to the precursors VLC-PUFAs: 32:6 and 34:6 in the presence or absence of 10ng/ml IL1 β and 10 ng/ml TNF α for 24 hours. The plot shows the box upper limit 3rd Quartile, bottom side 1st quartile, middle line: the median and the whiskers denote the maximum and minimum observations. *p<0.05 in t-test comparisons with the respective control.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [ExtendedData.pdf](#)