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Research Article

Keywords: restaurant waste, fruit waste, Black soldier fly (BSF), Geometric morphometrics, Discriminant Function Analyses (DFA)

Posted Date: July 19th, 2021

DOI: <https://doi.org/10.21203/rs.3.rs-710103/v1>

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Geometric Morphometrics Captures Variations in Wing Development of Black Soldier Flies

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Abstract

The black soldier fly is considered an important insect due to its ability to convert organic waste into high quality organic fertilizer. Studies have shown that food quality at the larval stage contributes greatly to their development. In this study, we applied geometric morphometric measurements to assess the variations in the development of the black soldier fly fed on different food substrates as captured in the wing. Eggs of black soldier fly were collected from the field and cultured on different food substrate such as; restaurant waste, fruit waste, wheat bran and layer meal until maturity. The right fore wings of 140 individuals of black soldier fly were used for the experiment. The results showed a significant difference between the shape of landmarks within each food substrate group. Landmarks 3, 7 and 9, corresponding to end of first radial vein R₁, intersession between medio- cubital vein and cubito- anal vein (m-cu and CuA), and intersession between cubito Anal vein and Anal vein (CuA + CuP) contributed most to the variation between the different food substrates. Partial least square showed a strong association between food substrate and wing development. Therefore, the variations in wing development could be due to the impact of nutritional compositions quality of the different food substrates.

1.0. Introduction

The Black soldier fly (BSF), *Hermetia illucens*, L. (Diptera: Stratiomyidae) is one of the important insects in entomophagy and perhaps the best insect feed products in the EU [1-5]. Its larvae is proposed as a promising technology for treating organic wastes or livestock manure [6-8]. Black soldier fly larvae are voracious feeders of decomposing organic materials and has the capacity to reduce these wastes by 42 – 70 %, [8-10]. This convert it into high quality organic fertilizer, which restores soil fertility. The larvae also reduce potential pollutants such as organic chemicals in manure and cause significant reduction of *E. coli* and *Salmonella enterica* in chicken and cattle manure [11,12]. The larval biomass contain around 42% protein and 35% fat [13], based on dry matter, including favourable profiles of essential amino acids [13], which are higher

40 compared to every other plant-and animal-based food ^[14]. These properties make BSFL a
41 suitable feed supplement candidate for fish and various livestock feed production ^[10, 15].

42 Most of the studies on black soldier fly, (*Hermetia illucens*), have focused on its high rate
43 of conversion of biomass to protein and lipid, and its potential role in waste management ^[16].
44 Globally, little is known about geometric morphometric assessment on the variation of wing
45 development of black soldier flies fed on different food substrates. The development of some
46 features such as the wing of the black soldier fly have been shown to vary ^[17], depending on the
47 quantity and quality of the diet supplied to them ^[18]. Geometric analysis of the size of the wings
48 is a widely used method for studying the effect larval diet quality on the growth of Diptera ^[19].

49 Morphometric analysis is a technique which measures shape by Cartesian landmark and
50 semi landmark coordinates, which has the potential of distinguishing morphological shape
51 variables. This technique was applied to study the variability in size of laboratory strains of flies
52 and variations among natural inhabitants ^[20]. Wing size variation assessment (based on centroid
53 size analysis) provide valid and reliable statistical information ^[21]. Geometric morphometrics
54 (GM) also indicates phenotypic variability and population structure with less effort ^[22], despite,
55 its quality for wing assessment, studies on *H. illucens* species are still limited ^[23-24]. In GM, both
56 wing size and shape are assessed autonomously for heritability and environmental sensitivity ^[25].
57 Variances from size and shape show consistent information about genetic variation of natural
58 populations showing high and stable heritability ^[26-29].

59 Morphometric characters may indicate reliable information concerning the biology of
60 insects, due to the influence of genetic and environmental conditions. Studies on insect wing
61 geometry could provide significant information for species identification and description of
62 population diversity. Geometric morphometric analyses utilize the wings due to its ability to
63 provide well-defined landmarks for morphometrics and the metric properties for precise
64 quantitative information. Landmarks are the intersections that biological structures are sampled.
65 These points provide particular geometric description of the modifications in shape of a structure.
66 A number of entomologists have carried out commendable works on insect wing morphometrics
67 as well as in mosquitoes. The current research is therefore based on the hypothesis that different
68 larvae substrates can contribute to the variability in adult black soldier's wing vein development.
69 This study was conducted to investigate the variation in the wing development of black soldier
70 flies fed on different substrates through landmark-based geometric morphometric analysis. The
71 findings of this study can provide insight into the population structure, ecology, and taxonomic
72 identity of the black soldier fly. Considering the importance of black soldier fly wings in insect
73 behaviour and physiology, the differences could also provide useful information on the quality of
74 diet, distribution, and variability in the wing development based on the food substrate they were
75 fed on.

76 77 **2.0. Results**

78
79 Results obtained from all the analyses showed significant differences ($P < 0.0001$)
80 between wing characters of black soldiers fly larval development fed on different food
81 substrates. Further, three wing veins showed significant variation in black soldier fly populations
82 that were fed on different food substrates. These veins were the following; end of first radial vein
83 R_1 , intersession between medio- cubital vein and cubito- anal vein (m-cu and CuA), and
84 intersession between cubito Anal vein and Anal vein (CuA + CuP). Procrustes ANOVA showed
85 a significant difference between the shapes of the landmarks ($P = 0.0001$) and the centroid size

86 ($P < 0.0001$) of the landmarks within groups of black soldier flies fed on different food
 87 substrates (Table 1).

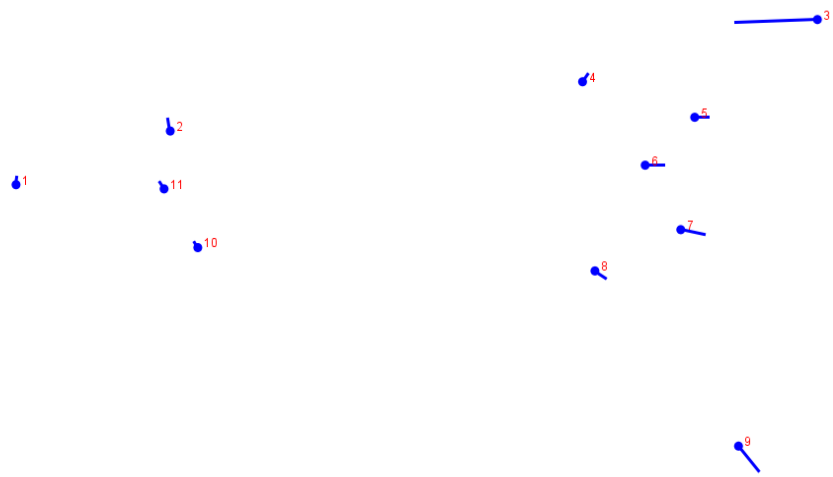
88
 89

90 **Table 1: The table below shows ANOVA values for the centroid and shape of landmarks.**

Centroid Size					
Effect	SS	MS	Df	F	<i>P</i>-(param.)
Individual	461497.326867	153832.442289	3	8.33	< 0.0001
Residual	2420517.234600	18477.230798	131		
Shape					
Individual	0.01855603	0.0003436302	54	2.57	0.0001
Residual	0.31537027	0.0001337448	2358		

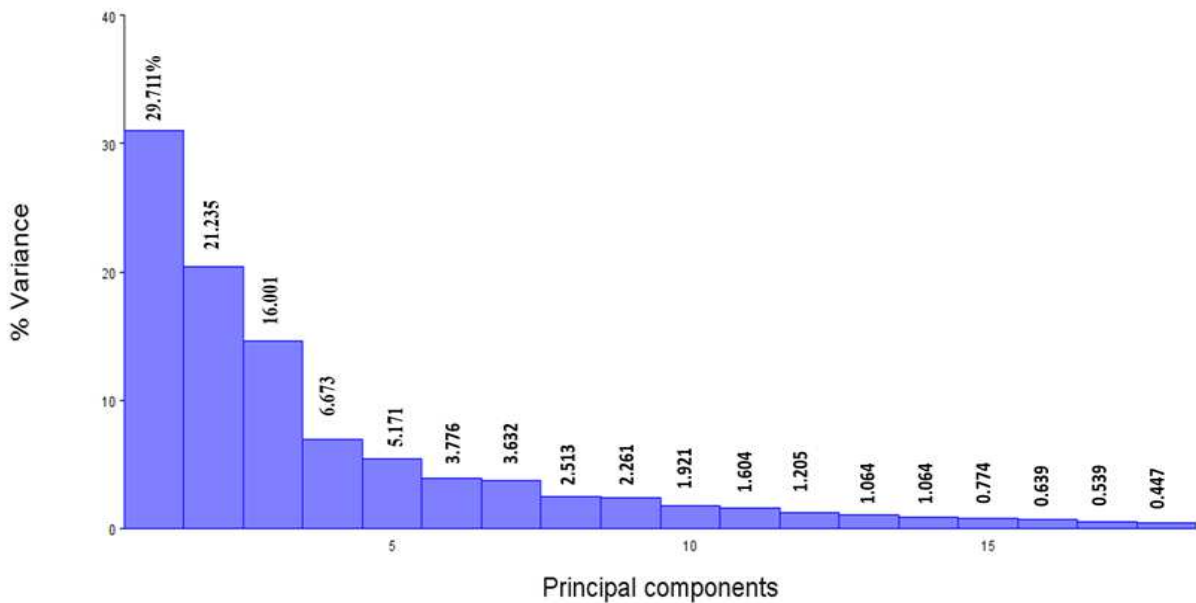
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92 During the principal component analyses, it was found that landmark 3 contributed most
 93 to the variability within the population followed by landmarks 7 and 9. (Figure 1). These
 94 landmarks correspond to the following wing features; end of first radial vein R_1 , intersession
 95 between medio- cubital vein and cubito- anal vein (m-cu and CuA), and intersession between
 96 cubito Anal vein and Anal vien (CuA + CuP). A sum of the Eigen score values of the first three
 97 components (PC1, PC2, PC3) contributed 66.94% of the total variability with each contributing
 98 29.711%, 21.235 and 16.001% respectively (Figure 2).



99

100 **Figure 1: This shows the shape changes or deviations from the centroid for each landmark.**



101

102 **Figure 2: This shows the eigen values for each component.**

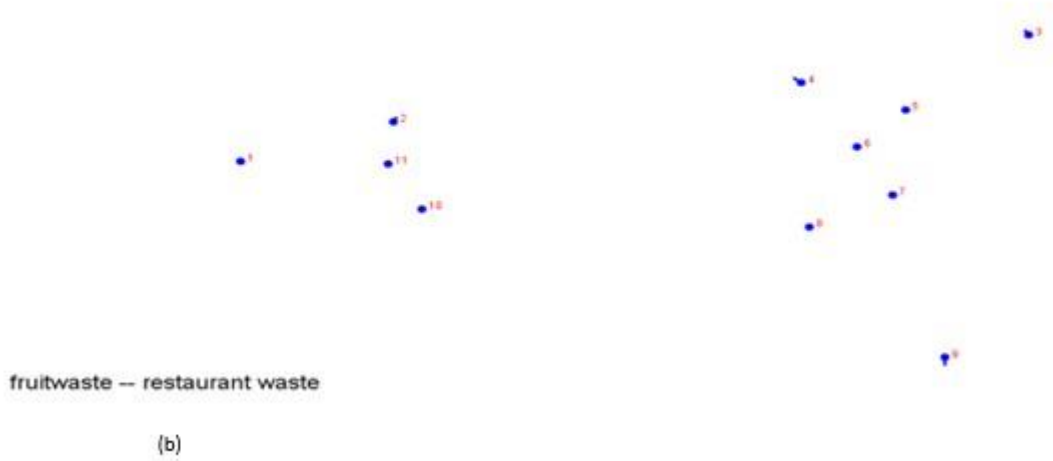
103 Discriminant Function Analyses (DFA) showed significant comparative differences
 104 within the populations of black soldier fly fed on different food substrates. Wing veins
 105 representing landmarks 3, 4 and 9 showed greater deviation from the centroid in all analyses
 106 except layer meal and restaurant waste where landmark 7 also showed marked variation from the
 107 centroid. In the black soldier fly populations that were fed on fruit waste and restaurant waste,
 108 significant differences ($P = 0.0253$) were observed with 79.41% of the population were found to
 109 be distinct with 55% accuracy in cross validation tests. On the other hand, DFA score for
 110 populations fed on restaurant waste were 82% discriminated with 62% accuracy in cross
 111 validation (Figures 3a & 4a). Further to this, a nonsignificant difference ($P = 0.1499$) was
 112 observed between populations fed on fruit waste and layer meal where DFA scores of 82% and
 113 76% respectively with cross validation of 62% and 61% respectively. (Figure 3b & 4b).
 114 Discriminant function analysis between samples fed on fruit waste and wheat bran showed non -
 115 significant differences ($P = 0.0624$), with DFA scores of 76% and 82% respectively while cross
 116 validation was observed at 56% and 59% respectively (Figure 3c & 4c). Results obtained from
 117 DFA for samples fed on layer meal-restaurant waste, layer meal-wheat bran, restaurant waste-
 118 wheat bran showed significant differences within groups with respective P values of 0.0001,
 119 0.0022 and 0.0017. The DFA scores for layer meal-restaurant waste were 90% and 94% with
 120 cross validation accuracy of 67 % and 65 % respectively, while scores between layer meal-wheat
 121 bran 85% in each group with cross validation of 67% and 76% respectively. Further, results
 122 obtained for DFA score discriminated restaurant fed sample by 85%, similar for samples fed by
 123 wheat bran with cross validation of 71% and 62% respectively (Figures 3d-f, 4d-f).



124

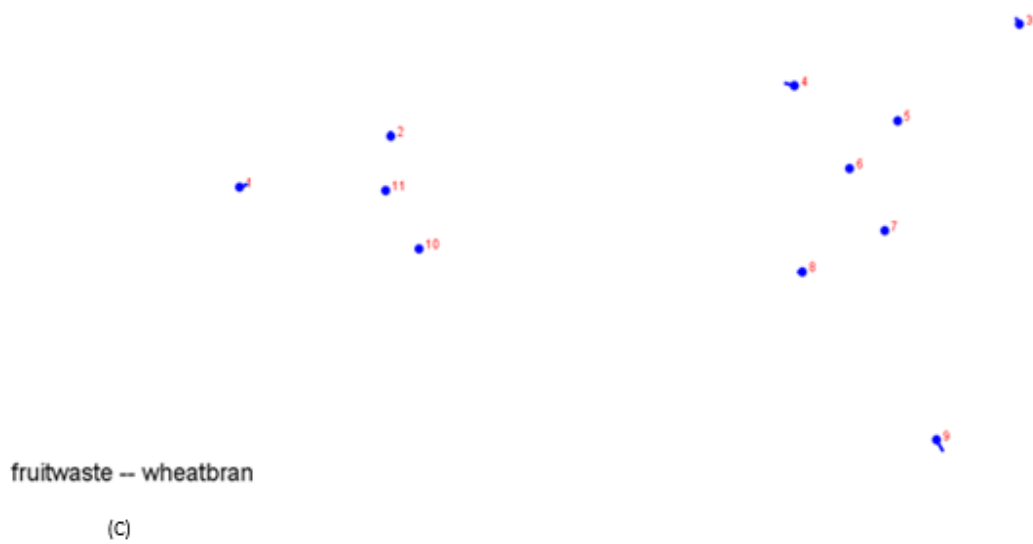
125 **Figure 3a: Graphical representation of shape difference for discriminant function analyses**
 126 **between samples fed on fruit waste and layer meal**

127



128

129 **Figure 3b: Graphical representation of shape difference for discriminant function analyses**
 130 **between samples fed on fruit waste and restaurant waste**

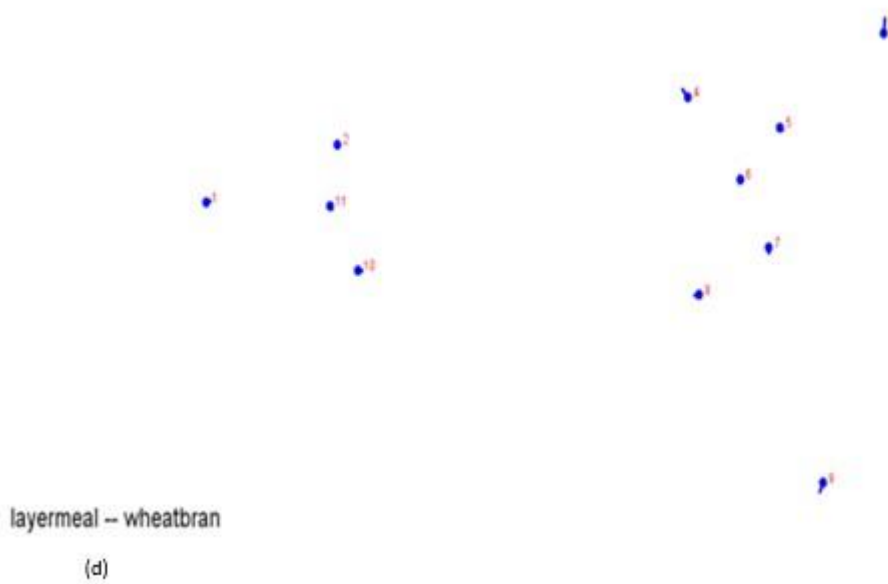


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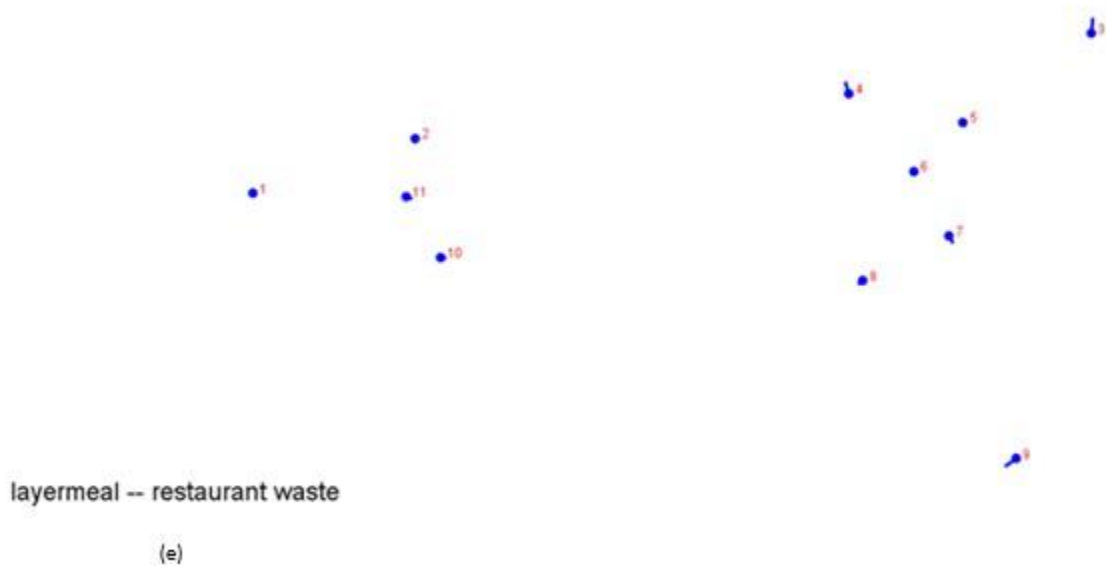
133 **Figure 3c: Graphical representation of shape difference for discriminant function analyses**
 134 **between samples fed on fruit waste and wheat bran**

135



136

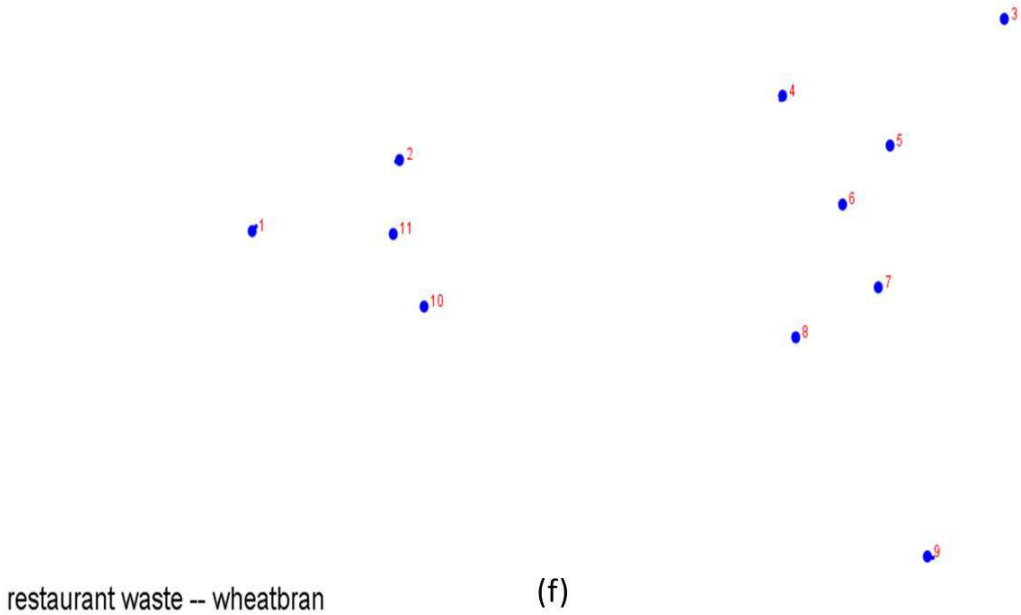
137 **Figure 3d: Graphical representation of shape difference for discriminant function analyses**
138 **between samples fed on layer meal and wheat bran**
139



140

141 **Figure 3e: Graphical representation of shape difference for discriminant function analyses**
142 **between samples fed on layer meal and restaurant waste**
143

144

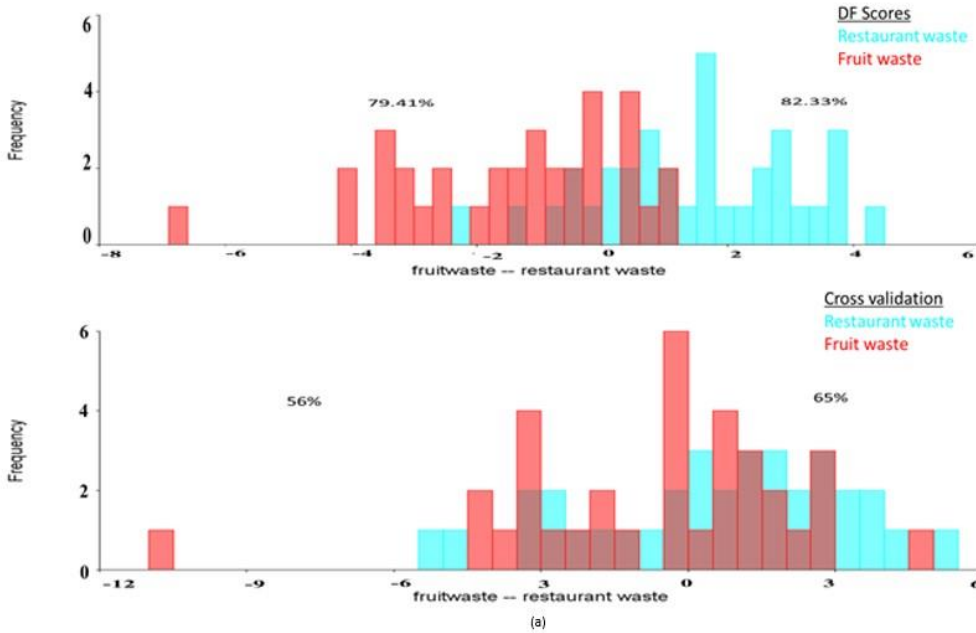


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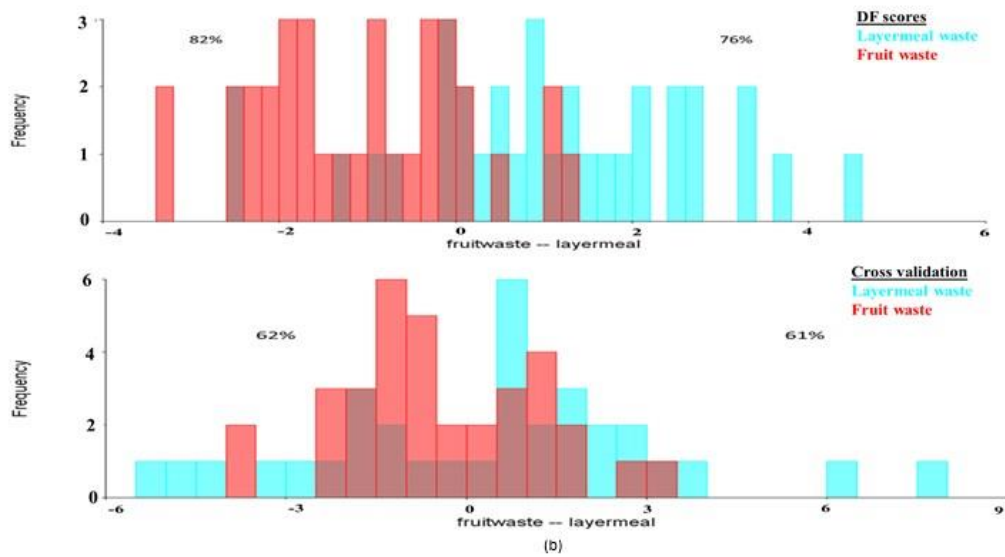
147 **Figure 3f: Graphical representation of shape difference for discriminant function analyses**
 148 **between samples fed on restaurant waste and wheat bran**

149



150

151 **4a: Graphical representation of discriminant function scores (a) and cross validation scores**
 152 **(b) for fruit waste – restaurant waste**



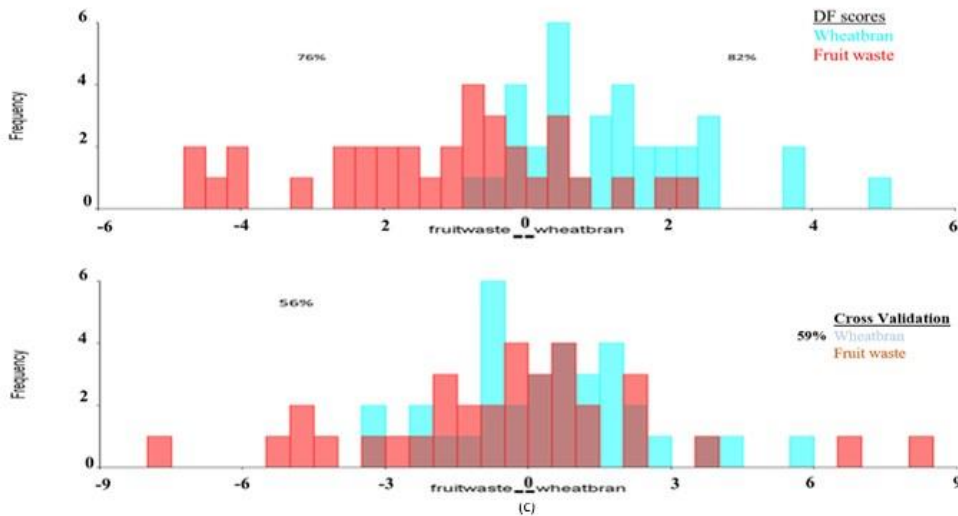
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154 **Figure 4b: Graphical representation of discriminant function scores (a) and cross**
 155 **validation scores (b) for fruit waste –layer meal**

156

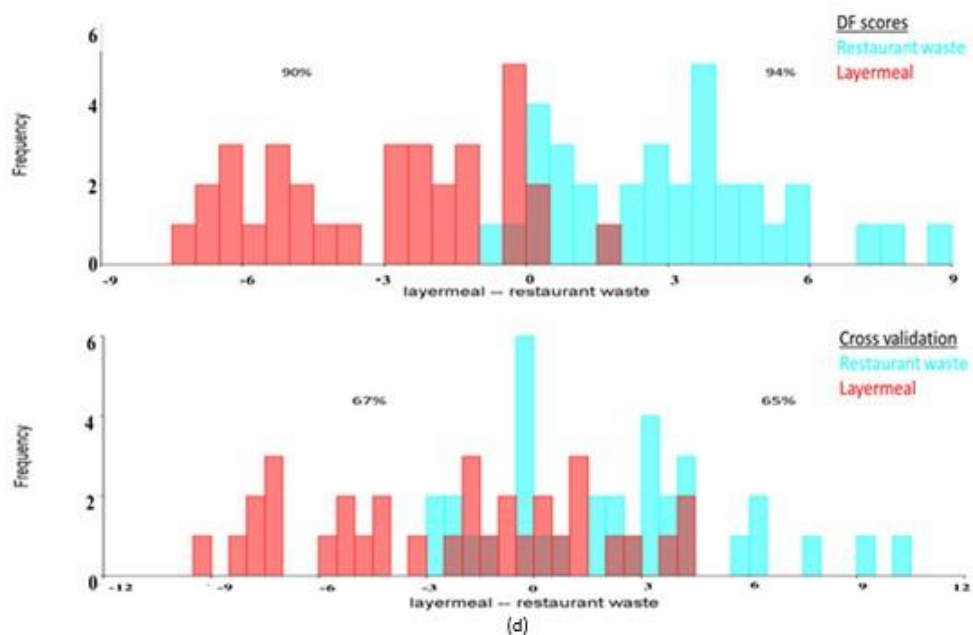
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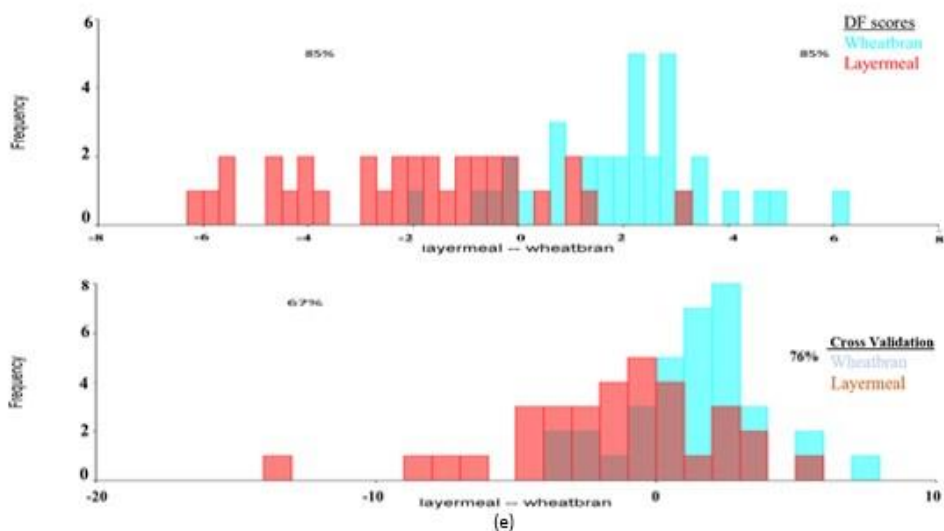


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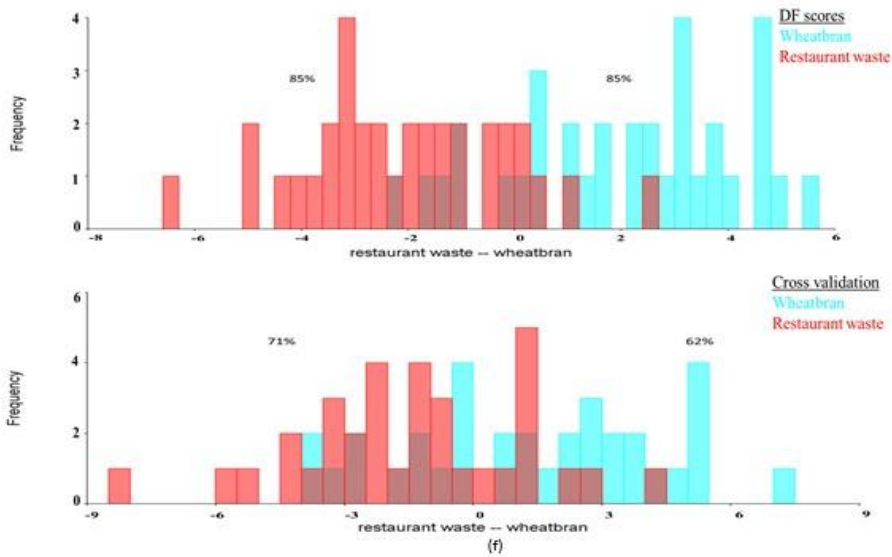
160 **Figure 4c: Graphical representation of discriminant function scores (a) and cross**
 161 **validation scores (b) for fruit waste – wheat bran**



162
 163 **Figure 4d: Graphical representation of discriminant function scores (a) and cross**
 164 **validation scores (b) for layer meal – restaurant waste**

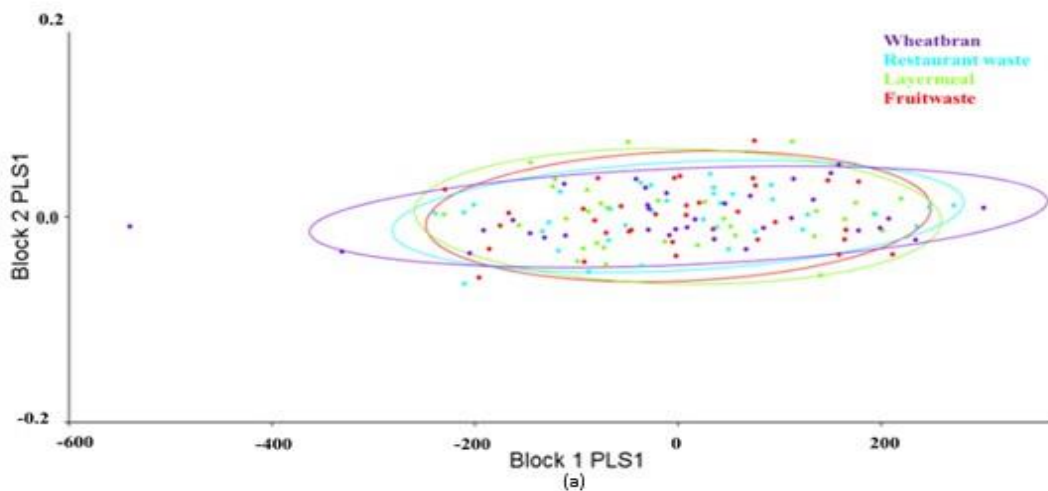


165
 166 **Figure 4e: Graphical representation of discriminant function scores (a) and cross**
 167 **validation scores (b) layer meal -wheat bran**

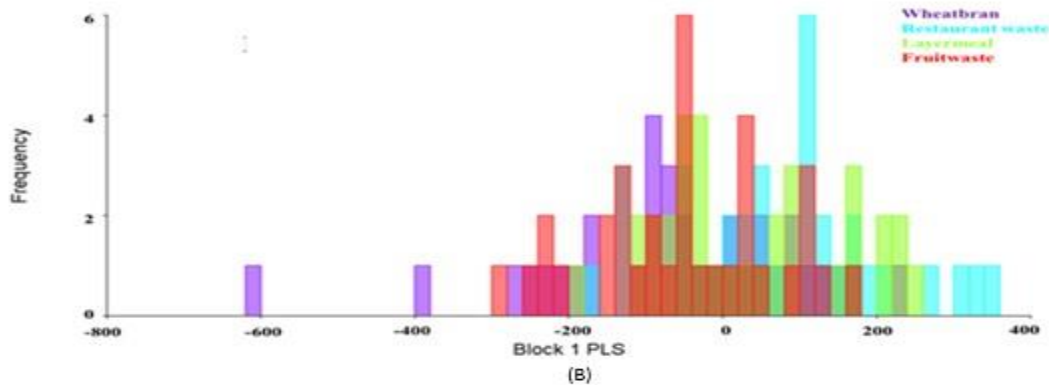


168
 169 **Figure 4f: Graphical representation of discriminant function scores (a) and cross validation**
 170 **scores (b) restaurant waste – wheat bran**

171
 172 Graphical representation of partial least squares showed a strong association between
 173 black soldier fly feed substrate and wing feature development with overall strength of association
 174 between feed substrate having RV coefficient of 0.0224. Further, populations fed on wheat bran
 175 showed the highest variability within wing characters. In figures 5a, the ellipse for wheat bran
 176 population further spread from the centroid on across the latitudinal axis as compared to
 177 populations fed on other food substrates. Samples fed on restaurant waste were also observed to
 178 show variation around the centroid, with fruit waste and layer meal exhibiting almost the same
 179 extent of variability with their respective groups (Figure 5a, 5b).
 180



181
 11



182
 183
 184 **Figure 5 a & b: Graphical representations of partial least squares showing association**
 185 **among food substrates and wing character variability.**

186
 187 **3.0. Discussions**

188 The development of insect features including wing features is attributed to their food
 189 resources and genetic traits ^[30]. In black soldier flies, wing developments vary, depending on
 190 the quantity and quality of the food supplied to them at the larval stage ^[17-18]. In the present study
 191 the results showed three main wing features that were significantly affected when BSF
 192 populations fed on the different food substrate. Generally, in insect wing vein development, the
 193 presence of additional or incomplete veins between fore wing cells occur around the marginal
 194 and cubital wing cells. In this present study, three main wing veins and vein intersections were
 195 found to be variable between cubital and medial veins. These variations in veins could therefore
 196 be due to (high sensitivity in response to the quality of nutritional compositions of the different
 197 food substrates. The present study also showed significant variations between the type of food
 198 substrate and the wing vein characteristics of BSF. Previous reports indicate that organic wastes
 199 release several volatiles such as; alcohol, aldehydes, and organic acids during decomposition that
 200 attract insects in response to variable developmental needs ^[31-33].

201 This suggest that these significant variations within the wing vain features could possibly
 202 be due to the presence of some volatile compositions in the substrates. However, it appears that
 203 the contribution of each food substrate to the variability is greatly dependent on the amount of
 204 volatile compositions present. Larvae that are cultured for waste treatment and feed production,
 205 are reared on wastes such as; vegetable, fruit, and other market and restaurant waste, with the
 206 aim of reducing cost of colony maintenance ^[33]. In this present study discriminant assessments
 207 showed BSF samples fed on layer meal-restaurant waste, layer meal-wheat bran, and restaurant
 208 waste-wheat bran showed significant differences within groups. However, no significant
 209 differences were observed within groups that fed on fruit waste- layer meal, fruit waste-
 210 restaurant waste and fruit waste -wheat bran. This possibly implies that in wing vein
 211 development of BSF, earlier sets of food substrate may induce significant growth differences,

212 compared with the latter sets of food substrates. This result will be useful in choosing best feed
213 substrate for any breeding experiment.

214 Poor nutrition may stimulate the production of alatae by the aphid *Myzus persicae*,
215 however, “low energy” diets do not support physiological conditions necessary for wing
216 development [34]. Suboptimal nutrition is often thought to stimulate wing formation in aphids by
217 influencing the “milieu intérieur” in a way that stimulates the process of wing determination.
218 Though the nutritional compositions of each feed substrate were not tested, populations fed on
219 wheat bran showed the highest variability or dispersion with restaurant waste fed populations in
220 close trend. This indicates that, comparatively, wheat bran contains high energy compositions,
221 which stimulated wing vein development by influencing the physiology of the of the black
222 soldier fly. The cross-talk of bombyxin and 20-hydroxyecdysone are reported to stimulate cell
223 division and growth of wing imaginal disks [35]. Moreover, the level of bombyxin in insects’
224 hemolymph is modulated by the brain in response to variation in nutrition, as well as the
225 mechanism that coordinates the growth of internal organs with overall somatic growth. The
226 contribution of high-level energy substrate to variability in wing vein development has been
227 reported in *Drosophila suzukii* on carbohydrate-rich media [36]. It is reported that adult flies
228 showed a strong preference for carbohydrate-rich foods in response to oviposition and in
229 transpecific interactions, however, larval survival and eclosion rate were strongly dependent on
230 protein rich food substrates.

231 **4.0. Conclusions**

232 The results of this study show that three wing veins seem to show significant variation in
233 black soldier fly populations that were fed on different food substrates. These veins include the
234 following; end of first radial vein R₁, intersession between medio- cubital vein and cubito- anal
235 vein (m-cu and CuA), and intersession between cubito Anal vein and Anal vein (CuA + CuP).
236 Moreover, there was a strong relationship between the different food substrates and BSF wing
237 vein developments. Black soldier fly samples that were fed on layer meal-restaurant waste, layer
238 meal-wheat bran, restaurant waste-wheat bran showed significant differences within groups.
239 However, no significant differences were observed within groups that fed on fruit waste- layer
240 meal, fruit waste -restaurant waste and fruit waste -wheat bran.
241 Populations fed on wheat bran showed the highest variability or dispersion with restaurant waste
242 fed populations in close trend, while fruit waste and layer meals showed low variations in
243 populations around the centroid. Larvae from the same generation were used for these
244 experiments, hence the results were not affected by genetic variation altering compatibility with
245 diet. Our discovery could provide theoretical basis and a new perspective for studies on black
246 soldier fly wing developments.

247 **5.0. Materials and Methods**

248 **5.1. Rearing and sample collection**

249 Larvae were obtained from a colony of black soldier fly, established from egg clutches
250 trapped from the wild and breed in the laboratory over seven years at the Ghana Atomic Energy
251 Commission, Accra, Ghana. Four groups of 100 individuals were selected and each group was
252 fed on different feed substrates namely; restaurant waste, fruit waste, wheat bran and layer meal.
253 The experiments were conducted in triplicate in plastic containers (Smart store classic 2, 21 x17
254 x11 cm) with netted lids, kept under ambient temperature of 28°C. In each box, 100 larvae (>0.2
255
256

257 cm in size, 5day old) were placed, giving a larval density of 0.6 larvae cm². Feed given was
258 computed based on a formula by Ewusie et al., [33] as follows:

259

260

$$261 \text{ Feed amount (g)} = \frac{\text{Number of larvae} \times 0.1 \text{ g} \times 25 \times 70\%}{MC} \quad (1)$$

262

263

264 Where MC = moisture content of diet

265 0.1 = Amount of feed per larvae per day [33]

266 25 = Estimated larval feeding duration (days) [33, 37].

267 70% = Adjusted moisture content of diets.

268 When the larvae reached the prepupal stage, they were removed with forceps into plastic
269 pupation containers with sterilized sand as the pupation material till eclosion under ambient
270 temperature and light. As soon as eclosion was seen in a container, the entire setup was placed in
271 a medium-sized wooden adult cage and any dead adult was picked and transferred into bottles
272 containing 70% alcohol solution for temporal storage till all eclosed adults died. The specimens
273 were sent to the museum of the Department of Conservation Biology and Entomology,
274 University of Cape Coast, Ghana, where extraction of wings, mounting of wings, capturing
275 images of wings and morphometric assessments were done.

276

277 **5.2. Geometric Morphometric Assessment**

278 **5.2.1. Slide Preparation**

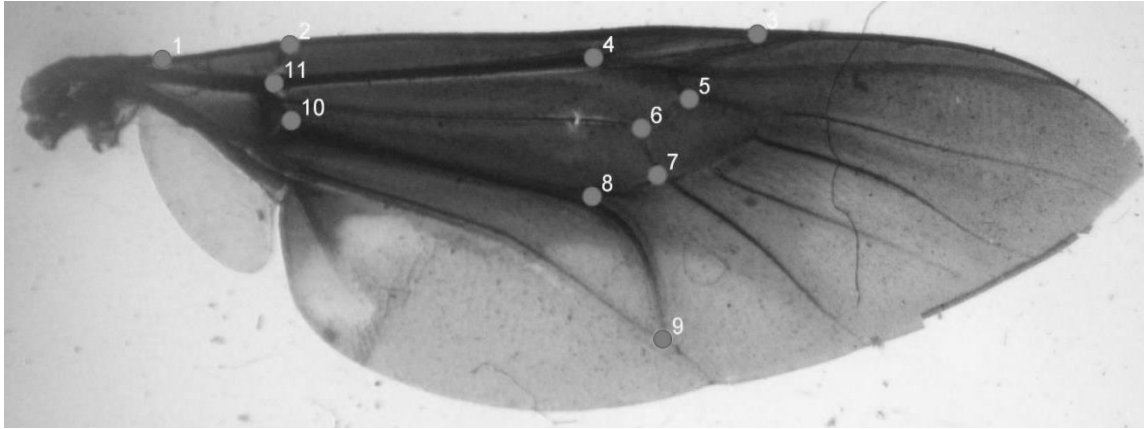
279 Specimens of approximately 55 adult black soldier flies were sampled from each of the
280 food substrate groups. The right forewing of each adult (total 140 wings) was removed, placed
281 between a microscope slides and secured in position with a microscope cover slip that had a drop
282 of transparent nail polish carefully applied to the ends. Wings were relaxed in 30% alcohol
283 before mounting unto slides. Each wing was photographed with a digital camera attached to a
284 microscope. Wing images were captured and an image file was created in JPEG format. One
285 TPS file was created from the image files using TPSUtil software (version 1.76; 38).

286

287 **5.2.2. Wing Venation Characters**

288 Eleven homologous landmarks were plotted at the junctions of the wing venation of the
289 adult black soldier flies using tpsDig2 software version 2.31 [39]. One TPS file grouped all the
290 eleven landmarks derived from each of the 140 wings of the black soldier flies fed on different
291 food substrates (Figure 5).

292



293

294 **Figure 6: Forewing of black soldier fly with eleven homologous landmarks.**

295 **5.3. Statistical Analyses**

296 The Procrustes coordinates of the 11 were first aligned using MorphoJ software (version
 297 1.03) and existing shape changes within the different fly populations bred on different substrates
 298 were evaluated. Procrustes ANOVA was done to assess the relative amounts of variation among
 299 individuals, of asymmetry and measurement possible error. Further, Principal component
 300 analysis assessment provided the most significant features contributing to variability within the
 301 group. Discriminant function analysis was used to distinguish between groups of adults fed on
 302 different food substrates and this revealed the probability of correct and incorrect classification
 303 of samples for each food substrate in pairs [25, 40]. In this work, partial least squares (PLS) were
 304 used to examine patterns of Covariation between black soldier flies fed on different food
 305 substrates. In geometric morphometric, one or more of these sets contain shape data [25].

306

307

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309

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425

426 **Author Contributions:** Conceptualization, R.C. and E.A.E, Data curation, R. C., E.A.E. and O.
427 G., Formal analysis, R. C. E. A. E., O. G. and I. D. Methodology, O. G., R.C., E.A.E I. B., S. Y.,
428 F. A., A. N.. Supervision, R. C. and E.A.E. Writing original draft, O. G., R.C., E.A.E I. B., S. Y.,
429 F. A., A. N. Writing review & editing, R. C., E. A. E. and PQ

430 **Funding:** This article was fully funded by Biotechnology and Nuclear Agriculture Research
431 Institute, Ghana Atomic Energy Commission, Accra, Ghana.

432 **Conflicts of Interest:** The authors declare no conflict of interest. The funders had no role in the
433 design of the study; in the collection, analyses, or interpretation of data; in the writing of the
434 manuscript; or in the decision to publish the results.

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