

Single Fibre Swelling Behavior for Natural and Man-Made Cellulose Fibres Under Industrial Steeping Conditions

Xiang You (✉ xiang.you@scitech.fi)

Scitech-Service Oy <https://orcid.org/0000-0002-1456-1357>

Feng Chen

ETH zurich

Yibo Ma

Aalto University

Annariikka Roselli

Metsa Spring Oy

Eric Enqvist

SciTech-Service Oy

Heikki Hassi

SciTech-Service Oy

Research Article

Keywords: Swelling, cotton, dissolving wood pulp, regenerated cellulose, cell wall

Posted Date: July 16th, 2021

DOI: <https://doi.org/10.21203/rs.3.rs-689489/v1>

License: © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

1 **Single fibre swelling behavior for natural and**
2 **man-made cellulose fibres under industrial**
3 **steeping conditions**

4 Xiang You^{1,*}, Feng Chen^{2,3}, Yibo Ma², Annariikka Roselli^{1,4}, Eric Enqvist¹, Heikki Hassi^{1,*}

5 ¹ *Scitech-Service Oy, Helsinki, FI-00130, Finland*

6 ² *Department of Bioproducts and Biosystems, School of Chemical Engineering, Aalto*
7 *University, 00076 Aalto, Finland*

8 ³ *Wood Materials Science group, Institute for Building Materials, ETH Zurich, 8093 Zurich,*
9 *Switzerland*

10 ⁴ *Metsä Spring Oy, Espoo, FI-02150, Finland*

11 * E-mail: xiang.you@scitech.fi, heikki.hassi@scitech.fi

12

13 Abstract

14 Swelling behavior of cotton, dissolving wood pulp (DWP), viscose staple fibre (VsF), and Tencel staple
15 fibre (TsF) in varying sodium hydroxide (NaOH) were investigated by means of optical microscopy and
16 were characterized by molecular mass distribution, X-ray diffractometer, and dynamic vapor
17 sorption. Effect of temperature (20-45 °C) and duration (0-120 min) was studied. The results reveal
18 that the swelling ratio of fibre in alkali solution depends on fibre accessibility and NaOH
19 concentration. Among all the materials, VsF exhibited the highest swelling ratio and lowest swelling
20 ratio has been observed for cotton fibre. The results suggest that the swelling is limited by the
21 presence of plant cell wall structures in cotton and DWP, rather from fringed-fibrillar, semi-crystalline
22 sub-structures, which result from the inherent tendency of cellulose molecules to form such
23 structures during the biosynthesis of plant cell walls as well as during the formation of regenerated
24 cellulosic textile fibre in wet-spinning.

25

26

27 *Keywords: Swelling, cotton, dissolving wood pulp, regenerated cellulose, cell wall*

28

29 **Declarations**

30 **Funding**

31 The financial support from Jenny and Antti Wihuri foundation (Grant No. 131219).

32 **Conflicts of interest/Competing interests**

33 The authors declare that they have no conflict of interest.

34 **Availability of data and material**

35 The authors declare that the data and material are available.

36 **Code availability**

37 Not applicable

38 **Authors' contributions**

39 Dr. Xiang You was responsible for the experimental design with Dr. Feng Chen and
40 Dr. Annariikka Roselli, performed the experimental work, analyzed the corresponding
41 results and wrote the manuscript as principal author, with copyediting by Dr. Feng
42 Chen, Dr. Yibo Ma, and Dr. Heikki Hassi, under the supervision of Dr. Eric Enqvist
43 and Dr. Heikki Hassi.

44 **Additional declarations for articles in life science journals that report the results
45 of studies involving humans and/or animals**

46 Not applicable

47 **Ethics approval**

48 Not applicable

49 **Consent to participate**

50 The authors declare that they agree to participate.

51 **Consent for publication**

52 The authors declare that they agree to publication.

53

54 Introduction

55 Cellulose-based plant fibres are highly interesting sources of polymers for producing
56 bio-based products. Native or derivatized cellulose has been used as a renewable
57 resource in a wide variety of industries (Isogai and Atalla 1998; Cuissinat and Navard
58 2006, 2008). Cotton and technical celluloses (chemical pulps) are the main cellulose
59 sources for industrial applications, for example, making paper, textile fibres and films
60 (Klemm et al. 2005; Olsson and Westman 2013). However, as a non-thermoplastic
61 polymer, the properties of cellulose prevent it from being processed for certain
62 applications (Kihlman et al. 2012, 2013; Zhang et al. 2013).

63 Due to the high degree of crystallization and strong inter- and intramolecular hydrogen
64 bonds, cellulose is relatively stable and is insoluble in water and many conventional
65 solvents. Moreover, the hydrophobic interactions between the cellulose molecular
66 sheets in the crystallite also restrict the dissolution of cellulose (O'Sullivan 1997;
67 Nishiyama et al. 2002, 2003; Foress and Fremer 2003). Currently, only few solvents
68 can directly dissolve cellulose (Gross and Chu 2010). The direct cellulose solvents can
69 be classified into the system: aqueous media and non-aqueous media. Aqueous media
70 consists of aqueous inorganic complexes (Cuam, Cuen), aqueous alkaline (NaOH),
71 acids and molten inorganic salt hydrates ($ZnCl_2/H_2O$, LiSCN). Non aqueous media
72 includes ionic liquids (ILs), organic solvent in presence of inorganic salts
73 ($LiCl/DMAc$), DMSO/TBAF and amine oxide monohydrate (NMMO) (Heinze and
74 Koschella 2005; Liebert 2010; Sen and Martin 2013; Olsson and Westman 2013). Of
75 all direct solvents, only NMMO monohydrate is used as direct cellulose solvent on an
76 industrial scale for spinning textile fibres (Lyocell process) (Chen et al. 2020).

77 The dissolution mechanisms of cellulose fibres in various direct solvents have been
78 studied in detail. When native cotton fibres and wood fibres are placed in a certain
79 swelling or dissolution agent, they show nonhomogeneous swelling. The solvent tends
80 to first penetrate the Primary (P) cell wall through pores and dissolve the cellulose in
81 the Secondary (S) cell wall. Since the cellulose chains are still trapped inside the fibre
82 structure and the solvent penetration continues, due to osmotic pressure, the increasing
83 solvent volume causes the expansion of the P cell wall (ballooning) (Cuissinat and
84 Navard 2006; Le Moigne et al. 2010; Olsson et al. 2014; Mäkelä et al. 2018). According
85 to the study of Cuissinat and Navard (2006), the behavior of DWP or cotton fibres in
86 various solvent systems (for example, NaOH-water or NMMO with 25 to 35% water)
87 can be characterized by five particular modes: Mode 1) Accelerated dissolution by
88 disintegration into cellulose fragments. Mode 2) Swelling in a large area by ballooning
89 followed by dissolution. Mode 3) Large swelling by ballooning, but without dissolution.
90 Mode 4) Homogeneous swelling without dissolution. Mode 5) No swelling and no
91 dissolution. These modes reflect the quality of the solvents from excellent, moderate,
92 weak, and non-solvent. In less powerful solvents ballooning may be followed by
93 bursting of S1 and, then dissolution. (Cuissinat and Navard 2006; Zhang et al. 2013;
94 Mäkelä et al. 2018; Chen et al. 2020). In the case of DWP fibres, the swelling of S2
95 layer of the S wall begins from the lumen (for example, filled with NaOH solution),
96 ballooning occurs and generates tension to fibrils of P wall and S1 layer, which are less
97 prone to swelling due to their more random fibril orientation. This tension further
98 hinders the swelling of P and S1. P wall bursts and form collars that remain undissolved,

99 which locally prevent swelling of S wall. Finally, also S1 layer bursts and gets
100 fragmented and dissolved in the form of aggregates. The same mechanisms apply to
101 cotton fibres. However, cotton is considered more difficult to swell than wood pulp,
102 which could be explained by the many sub-layers in its cell wall structure, with
103 different direction of orientation (Klemm et al. 2005).

104 Various factors must be taken into account to understand fibre swelling and dissolution
105 behavior: their chemical composition, properties of primary and secondary layers in
106 natural fibres or properties of a skin in some man-made fibres, porosity, and cellulose
107 crystallinity and molecular weight (Abu-Rous et al. 2006). For instance, viscose fibre
108 has folded perimeter and a skin-core structure, due to immediate solidification of
109 outermost fibre surface, under which there is a properly oriented skin layer. The core
110 is poorly oriented, due to, i.e. the non-Newtonian flow of viscose dope through
111 spinneret orifices. On the other hand, Lyocell fibre has a more uniform fibre structure
112 because of slower solvent exchange between the fibre and the coagulation media.

113 To the best of our knowledge, there is no report comparing the swelling behavior of
114 natural, chemical pulp, and man-made fibres, under the same conditions. In man-made
115 cellulosic fibres (MMCFs) manufacturing processes, it's critical to avoid (usually
116 microscopic) poorly swollen, unfragmented local domains of cellulose in the cell walls
117 of the pulp. Therefore, the dynamics of the swelling and/or dissolution processes is of
118 special importance.

119 This paper studies the swelling ratio and morphological changes of the structures of the
120 single fibre and free-floating cellulosic fibres in different NaOH solution
121 concentrations and temperatures. The behavior of cotton, DWP, VsF, and TsF with
122 varying NaOH solutions conditions is observed by using optical microscopy. This work
123 may contribute to the understanding of the cellulose swelling behaviors and the further
124 work on the development of the viscose process. The aim is to identify sources of such
125 defects (poorly swollen or undissolved, mostly microscopic residues) in spinning dope,
126 which reduce the filterability or increase spinneret clogging rate in industrial MMCF
127 manufacture.

128 **Experimental methods**

129 **Materials**

130 Raw cotton fibre without seed was obtained from Zhengzhou Share Machinery Co.,
131 Ltd, China. A commercially available DWP was obtained from Domsjö, Sweden. Two
132 other man-made fibres VsF (1.3 DTEX, 39 MM, bright, raw-white) and TsF (1.3
133 DTEX, 38 MM, bright, raw-white) were donated by Lenzing, Austria. The
134 characteristics of the fibres are presented in the Table 1. The solution was prepared by
135 dissolve solid NaOH pallet in the milli-Q water to 5.5, 12, and 18% NaOH
136 concentrations. All concentrations ratios are presented as % by weight (wt). All
137 materials were cut into shorter length (fibre length ≤ 5 mm) prior to the test.

138 **Method**

139 *Molecular mass distribution (MMD) of cellulose fibres*

140 MMD of the raw materials was determined by gel permeation chromatography. About
141 500 mg of the fibre sample was accurately weighted into a sample bottle. The samples
142 were activated by a water–acetone-N,N-dimethylacetamide (DMAc) sequence. The
143 activated samples (50 mg each) were dissolved in 90 g L⁻¹ lithium chloride containing
144 DMAc at room temperature and under gentle stirring. The samples were then diluted
145 to 9 g L⁻¹ LiCl/DMAc, filtered with 0.2 µm syringe filters, and analysed in a Dionex
146 Ultimate 3000 system with a guard column, four analytical columns (PLgel Mixed-A,
147 7.5 x 300 mm) and refractive index (RI) detection (Shodex RI-101). Flow rate and
148 temperature were 0.75 mL min⁻¹ and 25 °C, respectively. Narrow pullulan standards
149 (343–2,500 kDa, PSS GmbH) were used to calibrate the system. Number-average and
150 weight-average molar mass, M_n and M_w , were calculated.

151 *Determination of the crystallinity index by X-ray diffractometer (XRD)*

152 XRD patterns of all macroscale cellulose fibres were collected in the transmission
153 mode setting of an X-ray instrument (SmartLab, Rigaku) that was operated at 45 mA
154 and 200 kV. All fibre samples were cut into debris and mounted on a multi-position
155 sample holder. The Scans were performed in a 2θ range between 5° and 60° by $\theta/2\theta$
156 mode.

157 The background profile was estimated under the same conditions, and it was subtracted
158 from the obtained data. The crystallinity index (*CRI*) of fibres was estimated with the
159 Segal method, using the intensity of crystal plane (I_{200}) at $2\theta = 22.7^\circ$ and of amorphous
160 parts ($I_{amorphous}$) at $2\theta = 18^\circ$:

161
$$CRI (\%) = 100 \times \left(\frac{I_{200} - I_{amorphous}}{I_{200}} \right) \quad (1)$$

162 *Dynamic vapor sorption (DVS)*

163 A DVS intrinsic apparatus (Surface Measurement system, London, UK)) with a
164 measuring accuracy of 0.1 µg was used for water vapor sorption analysis. Around 10
165 mg of pre-dried grinded fibres were placed in the sample pan and preconditioned at a
166 relative humidity (RH) of 0% at 25 °C until the equilibrium was reached using nitrogen
167 (flow rate 100 cm³ s⁻¹). The adsorption cycle was performed in 0% RH steps to a
168 maximum of 90% RH, and vice versa for the desorption cycle. In both cases, the RH
169 was kept constant until a defined equilibrium condition was reached, for example,
170 when sample mass change was below 0.001% per minute over 10 min. Then RH was
171 increased to 90% and sample weight evolution and time to reach equilibrium (t_s) were
172 recorded. For desorption the RH was decreased to 0%, and time to reach equilibrium
173 (t_{des}) were recorded. Moisture sorption M_s (from M_0 to M_{90} at 90% RH) were calculated
174 as follows:

175
$$M_s = \frac{M_{90} - M_0}{M_0} \times 100\% \quad (2)$$

176 *Optical microscopy observations of fibre swelling*

177 One single fibre was placed between two glass plates with the in-between distance
178 around 140 μm controlled by spacers. The distance between two glass slides is much
179 larger than the fibre's thickness. The ends of the fibre were fixed by tape and 1-2 drop
180 of solvents added. Fibre diameter was recorded by a Zeiss Axio optical microscope in
181 transmission mode equipped with a CCD camera (Metallux 3, Leitz) and Linkam TMS
182 91 hot stage to control temperature. The evolution of the fibre diameter in time t was
183 calculated as D_t/D_0 , where D_0 and D_t are initial diameter of the fibre at $t=0$ and
184 diameter of fibre at time t , respectively.

185 **Results and discussion**

186 **Physico-chemical properties of cellulose fibres**

187 Cotton is a natural cellulose fibre (cellulose content $\sim 99\%$) with a complex morphology;
188 its elementary fibre consists of more secondary (S) cell wall layers than wood fibre.
189 The sub-layers of S wall are highly ordered and have a compact structure with the
190 microfibril lying parallel to one another (Cuissinat and Navard 2006). DWP has special
191 properties including high α -cellulose content ($>90\%$ of dry matter) and relatively low
192 molecular weight comparing to the cotton fibre (Mozdyniewicz et al. 2013). DWP is a
193 raw material for man-made cellulosic fibre production. VsF is made via xanthation of
194 alkali cellulose made of DWP, followed dissolution and regeneration of the fibres by
195 wet spinning in acidic coagulation bath. Due to the nature of the spinning, VsF consists
196 of a dense skin and spongy core (Rous et al. 2006). On the other hand, TsF is made via
197 direct dissolution in NMMO and spinning using Lyocell process. TsF has more
198 homogeneous morphology, with a semipermeable skin, porous middle zone and dense
199 fibre core (Rous et al. 2006, 2007).

200 To better understand the swelling behavior of the selected materials, a few physical
201 properties of the material need to be characterized. The number-average (M_n) and
202 weight-average (M_w) molecular weights of cellulose in each fiber are presented in Table
203 1, and the corresponding molar mass distribution are demonstrated in Figure 1a. The
204 MMD of the raw cotton show a narrow distribution, and this is in accordance with what
205 others have found for native cotton fibres (Timpa and Ramey 1989; Kleman-Leyer et
206 al. 1992; Palme et al. 2014). Unlike cotton, DWP sample revealed subtle shoulder and
207 high molecular weight peak indicating the presence of both low molar mass
208 hemicellulose and relative long-chain cellulose. A distinct low molecular weight peaks
209 for VsF and TsF are similar for all man-made fibres. It has to be noticed that cotton and
210 VsF needed more time (2 weeks) to completely dissolve in LiCl/DMAc solution prior
211 to the chromatographic analysis. The reason for cotton is due to high molecular weight
212 and possible reason for VsF is due to low accessibility caused by hornification.

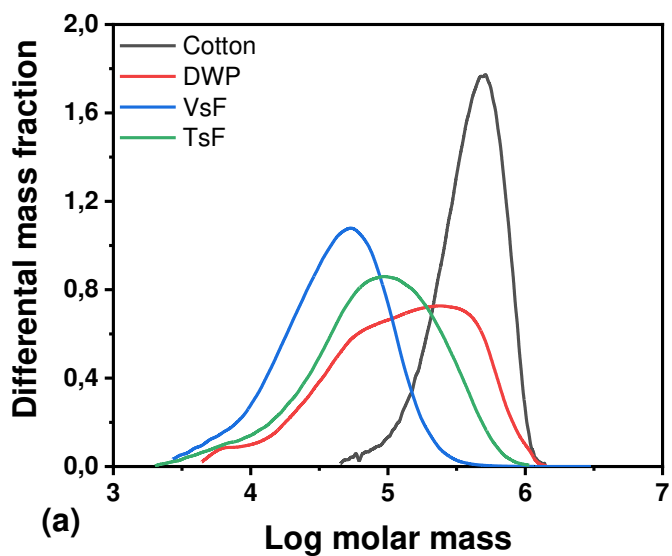
213 The crystallinity of the material also governs their swelling behavior and it was
214 analysed by XRD. The XRD patterns for the fibres are shown in Figure 1b. Cotton
215 cellulose spectrum accordingly represents Cell-I, containing both crystalline and
216 amorphous regions. Its Cell-I with crystallinity index of approximately 49.8%, which
217 is typical for plant cellulose (Placet et al. 2012; Moryganov et al. 2018; Chen et al.
218 2020). DWP has slightly lower crystallinity comparing to cotton due to intensive

219 pulping procedure and present of the small amount of amorphous hemicellulose. The
 220 MMCFs demonstrate the Cell-II allomorph, as expected. The *CRI* is around 32.2% and
 221 39% for VsF and TsF, respectively (Table 1). These results are in agreement with the
 222 previously reported crystallinity of MMCFs (Kreze and Malej 2003).

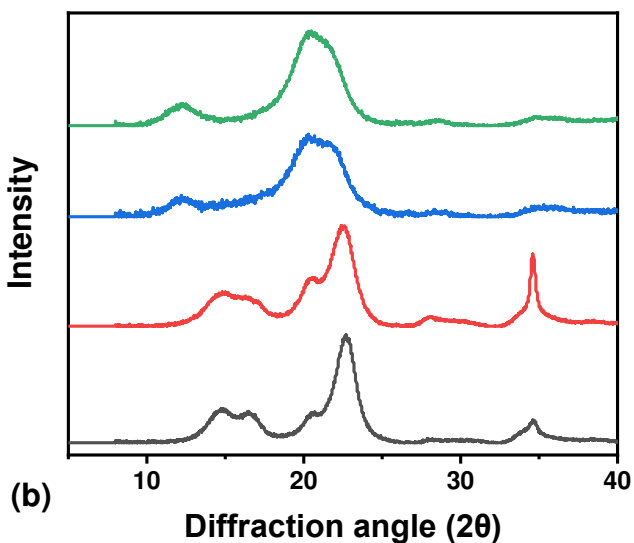
223 To compare the accessibility of the fibres, the dynamic water vapor sorption and
 224 desorption were measured. Moisture sorption (M_s), time to reach equilibrium t_s from 0
 225 to 90% RH, and time to reach equilibrium t_{des} from 90 to 0% RH for each fibre are
 226 shown in Table 1. Cotton absorbs the least amount of vapor probably because of the
 227 “resistant” primary wall (Okubayashi et al. 2004; Xie et al. 2011) and higher cellulose
 228 crystallinity (*CRI* 49.8%). In the study of Abu-Rous (2006), cotton showed very small
 229 pores in the bulk of the fibre, but the drying cracks and flat pores between the sheets of
 230 the secondary wall appear as larger pores. Similar to the cotton fibre, the DWP also
 231 demonstrate relatively low vapour sorption due to the high crystallinity (*CRI* 45.8%).
 232 VsF absorbs the highest amount of moisture (M_s) due to its lowest crystallinity (*CRI*
 233 32.2%) and wide pore size distribution from nanometre to micrometre scale
 234 (Okubayashi et al. 2005; Abu-Rous et al. 2006). Time to reach equilibrium, $t_s = 1260$
 235 min, is the longest for VsF among the selected fibres probably could be explained by
 236 the hornification phenomena (Siroka et al. 2008) as viscose fibres may be overdried,
 237 and also because the absorption of a larger amount of vapor takes more time which
 238 related to its large surface area. TsF as one type of Lyocell fibre contains only
 239 nanopores in the bulk of the fibre with a slight gradient in pore density, and a very
 240 porous skin layer. As a result, TsF shows slightly lower moisture sorption than VsF. In
 241 the literature studies, cotton appears to absorb only little water; Tencel shows uniform
 242 water absorption over the whole fibre cross section and uneven water distribution was
 243 found in Viscose fibre (Abu-Rous et al. 2006).

244 **Table 1.** Characteristics of cellulose fibres.

Sample	M_n (kDa)	M_w (kDa)	<i>CRI</i> (%)	M_s (%)	t_s (min)	t_{des} (min)
Raw cotton	324	444	49.8	12.71	1113.1	1170.2
DWP	59	208	45.8	13.84	1260.1	1220.2
VsF	26	57	32.2	21.76	1674.8	2243.0
TsF	40	127	39	18.85	1497.4	1884.7



245



246

247 **Figure 1.** (a) Molecular mass distributions (MMD) and (b) X-ray diffraction (XRD) profiles of cotton,
248 DWP, VsF and TsF

249 **Fibre swelling behavior in NaOH solution**

250 *Cotton*

251 As shown in Figure 2, dried cotton fibre collapses into a flattened bean-like cross
252 section. It also twists along its length so that it looks like a twisted and flattened ribbon
253 (Figure 2, 0 min). These twists are called convolutions: there are about 60 convolutions
254 per centimetre (Dochia et al. 2012). In this study, the diameter of convolutions was
255 measured to indicate the evolution of cotton fibre swelling ratio. The treatment
256 temperature of 36 °C and 45 °C were chosen because they are close to the process
257 conditions in the viscose process. Moreover, for viscose process, higher concentration

258 is necessary to ensure that the conversion to the alkoxide derivative proceeds to an
259 acceptable extent. In industrial viscose process, typical NaOH concentration is around
260 17-19 wt%. (Cuissinat and Navard 2008; Wilkes 2011).

261 The images of cotton fibre evolution in NaOH solution at 45 °C are shown in Figure 2.
262 Fibres show minor swelling, intermediate swelling, and swelling, in a 5.5, 12, and 18
263 wt% NaOH-water solvent in the selected conditions 45°, respectively. The swelling
264 behavior of cellulose fibres in the relatively low NaOH solution indicates the weak
265 solvating power. This may be reflected by the non-swelling/dissolution in amorphous
266 state and crystalline region. When the cotton fibres are steeped in 12, and 18 wt%
267 NaOH solution, cotton fibres loss of crystalline structure observed by the intermediate
268 swelling and swelling phenomenon within 10 minutes. For all three NaOH solution
269 concentrations, fibre swelling ratio kept constant after 10 minutes (Figure 3). The
270 morphological changes of the cotton fibres at 45 °C are slightly higher comparing to
271 that at 36 °C, swelling ratios 1.5:1.4, 2:1.7, 2.3:2 at 5.5, 12, and 18wt % concentrations,
272 respectively. In industrial alkaline steeping conditions is generally performed at 30 –
273 55 °C.

274

275

276

277

278

279

280

281

282

283

284

285

286

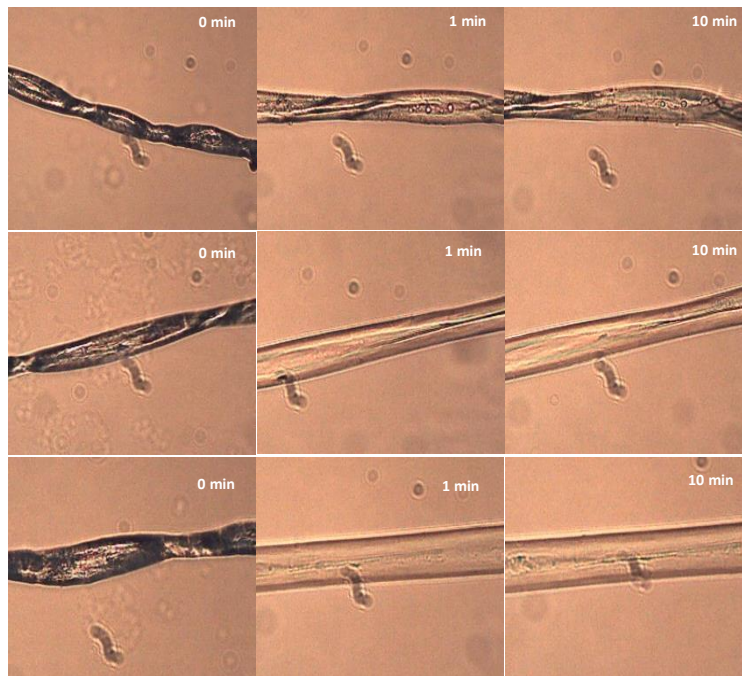
Cotton

5.5 wt%

12 wt%

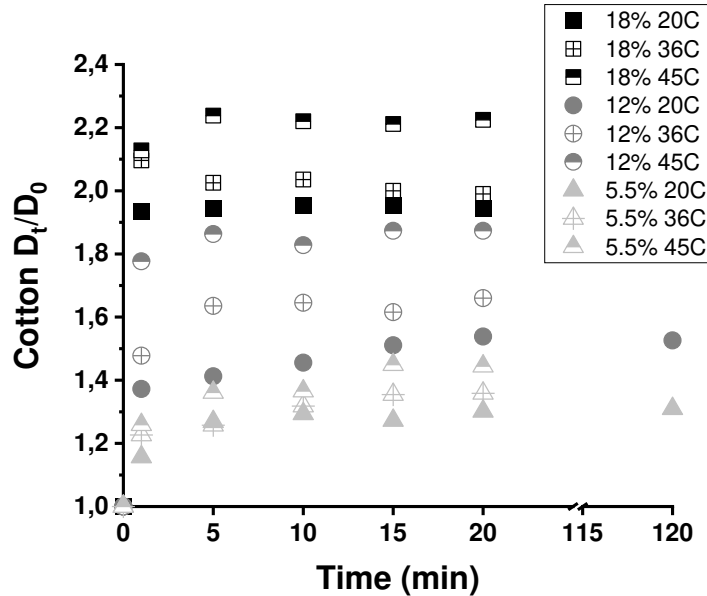
18 wt%

40 μm



287

Figure 2. Optical microscopy images of cotton at 45 °C in 5.5, 12, 18 wt% NaOH solution.



288

289 **Figure 3.** Evolution of cotton normalized diameter as a function of time.

290

291 *Dissolving wood pulp*

292 In the industrial scale viscose process, bleached dissolving wood pulp is typically the
 293 raw material and treated with steeping lye which contains a NaOH concentration of
 294 about 18 wt%. In this step and during the subsequent pre-aging, cellulose is converted
 295 into alkali cellulose for xanthation and its degree of polymerisation is adjusted to a
 296 level required for further processing (Mozdyniewicz et al. 2013). However, no suitable
 297 literature data are available for the swelling behavior of the DWP and the temperature
 298 and NaOH concentration ranges occurring during steeping of DWP. Therefore, this set
 299 of experiments describing the basis of DWP steeping, as a first step of viscose fibre
 300 manufacture.

301 Figure 4 presents the morphological changes of DWP fibres in 5.5, 12 and 18 wt%
 302 NaOH solutions at 36 °C. from the images, we have observed a pronounce increase in
 303 the fibre diameter and the inner layer of the fibre has been disintegrated at 12 wt%
 304 solution. DWP fibres steeped in both 5.5 and 18 wt% solution showed similar swelling
 305 behavior. At 20 °C and 36 °C (Figure 5) fibre swells more in 12 wt% NaOH solution
 306 in such relative low reaction temperatures, which had been confirmed in previous
 307 studies. This effect reflected that the solvating power of NaOH-water solution is
 308 governed by both NaOH concentration and treatment temperature. An increase in the
 309 temperature associated with too high or too low NaOH concentration decrease
 310 solvating power of the NaOH solvent. At higher temperature (45 °C), the NaOH-water
 311 solution dissolves more of the short-chain material. Some depolymerization also takes
 312 place during steeping, and the extent of this will increase with increasing temperature
 313 that eventually lead to partial dissolution of the pulp (Mozdyniewicz et al. 2013). This
 314 could be explained by the low DP of DWP which can be nearly fully dissolved, with
 315 few insoluble particles present.

316

317

318 DWP

319 5.5 wt%

320

321

322 12 wt%

323

324

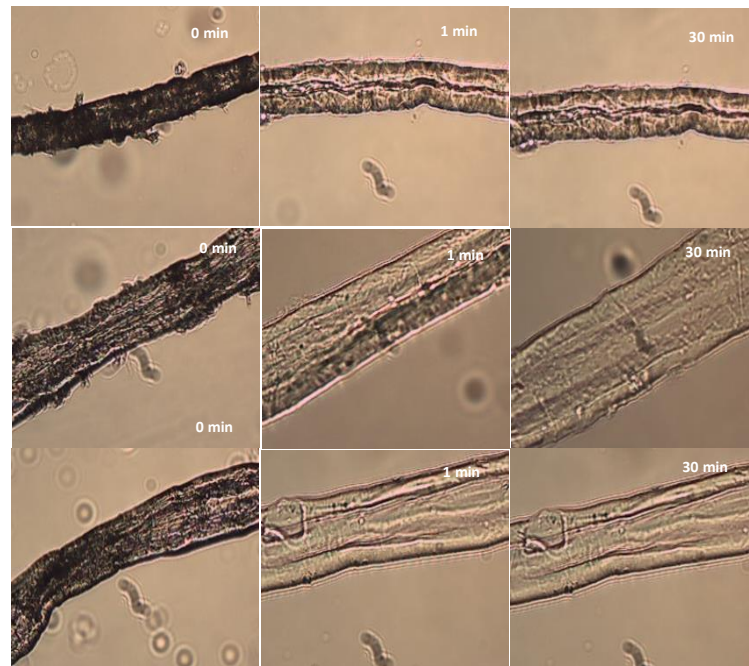
325

326 18 wt%

327

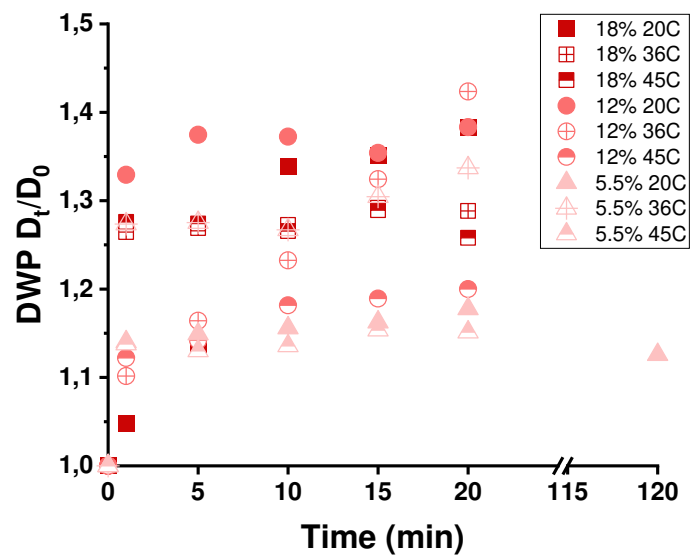
40 μ m

328



329

Figure 4. Optical microscopy images of DWP at 36 °C in 5.5, 12, 18 wt% NaOH solution.



330

331

Figure 5. Evolution of DWP normalized diameter as a function of time.

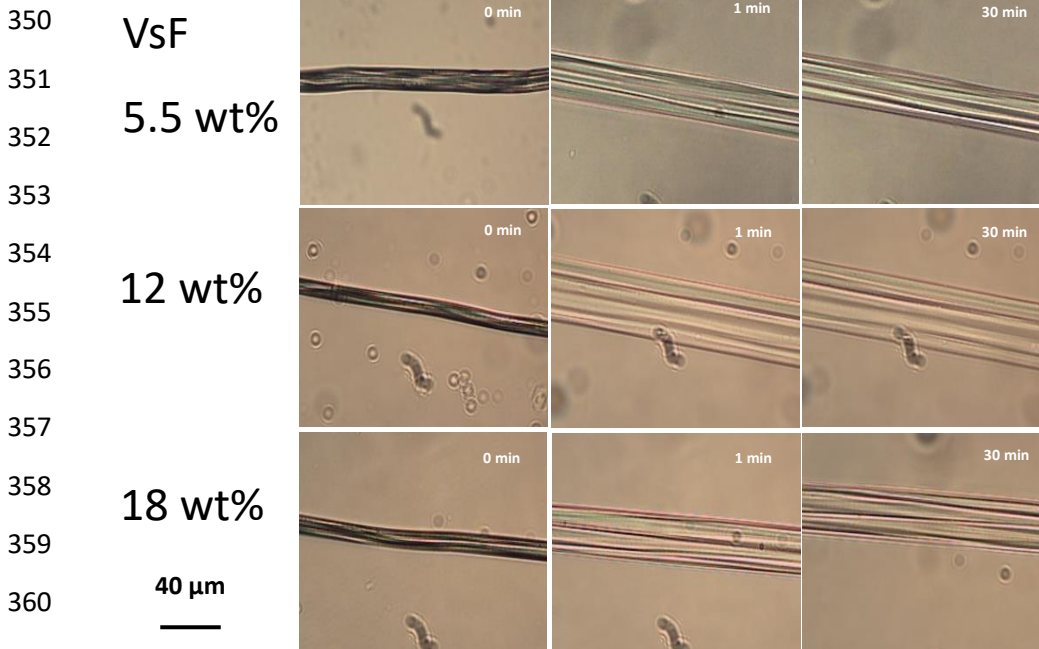
332

333 *Viscose staple fibre and Tencel staple fibre*

334 Alkali treatment transforms the structure of native cellulose I of DWP and cotton into
335 cellulose II. This transformation is generally known as mercerization. For regenerated
336 fibres composed of cellulose II, the alkali treatment only increases the amorphous
337 phase of the fibres (Rojo et al. 2013). The fibres swelling behavior upon the addition
338 of the solvents at different temperatures are presented in Figure 6 and 7.

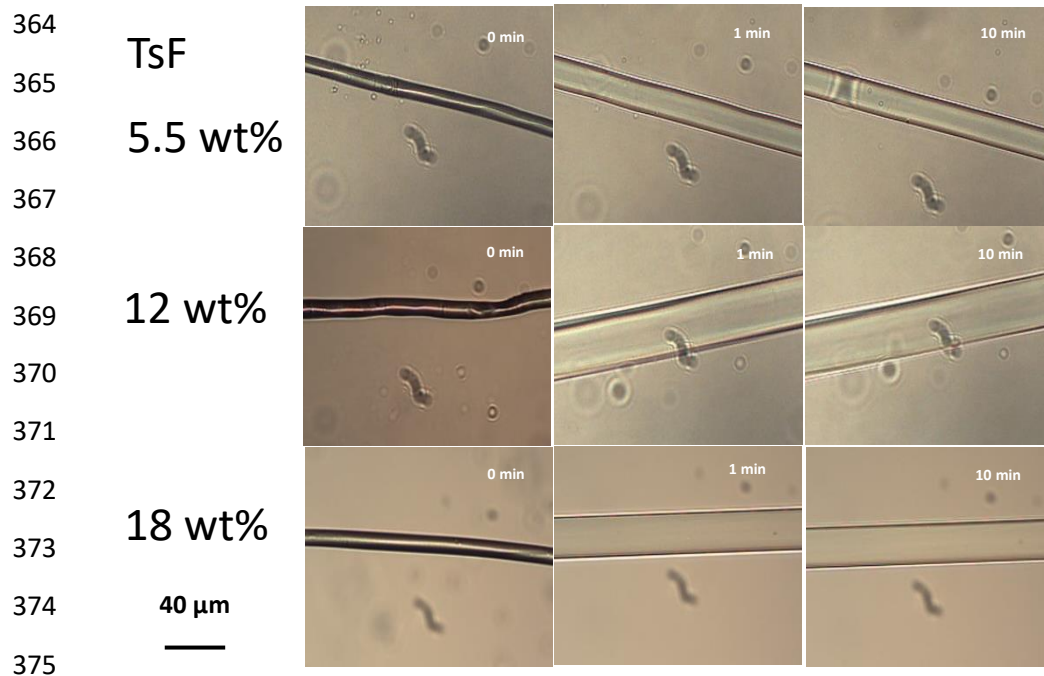
339 VsF and TsF fibre swell (Figure 6 and 7) more in 12 wt% NaOH solution, which is
340 similar to DWP swelling those fibres swelling increase as the concentration is reduced.
341 According to literature, pulp swelling to a maximum at around 11 wt% NaOH
342 concentration (Wilkes, 2011). VsF and TsF swelling radially in a homogeneous way.
343 These conditions increase the amorphous regions of the fibres and swell the microfibrils
344 and fibres. Fibre swelling ratio of viscose fibers were different from that of Tencel
345 fibers, which was attributed to its inhomogeneous (skin-core) fiber structure compared
346 to homogeneous fiber structure of Tencel fibers. The normalized diameter D_t/D_0 of
347 fibres placed in NaOH is plotted in Figure 8 a,b as a function of time. VsF was swelling
348 severe at 45 °C. In contrast, TsF was swelling severe at 20 °C.

349

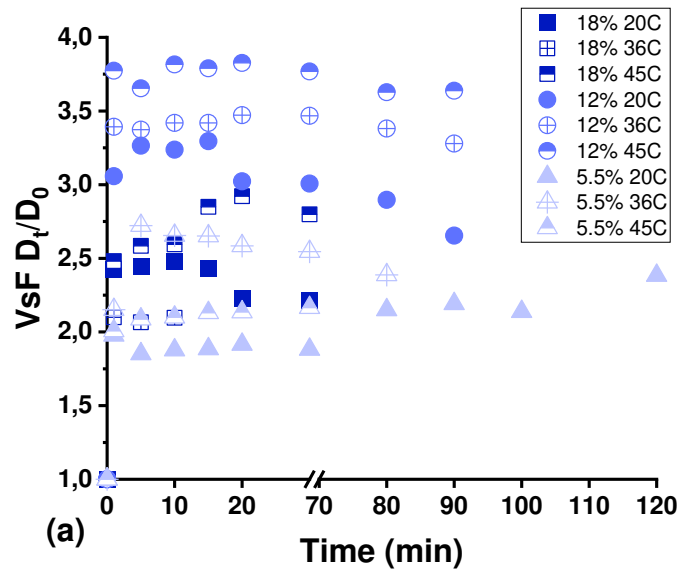


362 **Figure 6.** Optical microscopy images of VsF at 45 °C in 5.5, 12, 18 wt% NaOH solution.

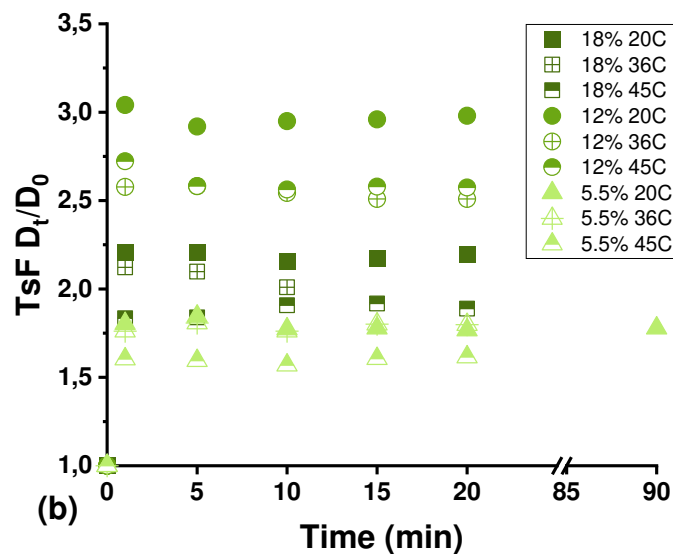
363



376 **Figure 7.** Optical microscopy images of TsF at 20 °C in 5.5, 12, 18 wt% NaOH solution.



377



378

379 **Figure 8.** Evolution of (a) VsF, and (b) TsF normalized diameter as a function of time.

380

381 **The overall conversion in the viscose process**

382 The microfibrillar structure of a man-made cellulosic fibre resembles strongly the
 383 original fibrillar structure of DWP cell wall layers. To make a spinning/processable
 384 dope, we know that cellulosic spinning dopes are not molecularly dispersed systems.
 385 They comprise ordered structures like fringed micelles, which tend to further aggregate
 386 into larger structures. Therefore, the critical factor is not to dissolve better, what is
 387 already properly dissolved, but to avoid poorly swollen, poorly dissolved domains of
 388 cellulose (Schuster et al. 2003; Le Moigne 2008).

389 As the purpose of this study was to gain better understanding of the sources of poorly
390 swollen and poorly dissolved residues in viscose spinning dope by comparing the
391 swelling of cotton, DWP, VSF and TSF, the experimental conditions used in this study
392 don't represent the real swelling behaviour during the steeping process with large scale.
393 During the steeping stage of the viscose process, the raw material concentration is
394 higher than suppresses the penetration of the liquor into the raw material due to the
395 reduced solid to liquor ratio. On the other hand, the formation of the raw material also
396 influences the swelling behaviour. In case of DWP, pulp sheet may be used instead of
397 powders for steeping. The extreme compact structure and low surface area of the pulp
398 sheet seriously lower the swelling rate of the cellulose. As for the raw material with
399 long fibre format, the surface area of the material has also to be reduced by means of
400 grinding in order to improve the penetration of the liquor into the materials. Grinding
401 is also very important to avoid the accumulation of the fibres on the impeller which
402 may lead to non-homogenous lye.

403 **Conclusions**

404 These results give good basis for more detailed identification of the sources and
405 formation mechanisms of incompletely dissolved cellulosic particulates and gels in
406 alkali-water based spinning dopes due to bad swelling. Such defects in spinning dopes
407 are often the main factor limiting the production efficiency of viscose spinning lines
408 and fibre quality. The results suggest that the swelling is limited by the presence of
409 plant cell wall structures, which result from the inherent tendency of cellulose
410 molecules to form such structures during the biosynthesis of plant cell. However, also
411 other factors like hornification of fibres during drying may limit swelling.

412 **Acknowledgments**

413 The financial support from Jenny and Antti Wihuri foundation (Grant No. 131219) is gratefully
414 acknowledged. Authors wish to thank Maija Nenonen and Sari Larmu for simulating industrial viscose
415 process.

416 **References**

- 417 Abu-Rous M, Ingolic E, Schuster KC (2006) Visualisation of the fibrillar and pore morphology of
418 cellulosic fibres applying transmission electron microscopy. *Cellulose* 13:411-419.
419 <https://doi.org/10.1007/s10570-006-9052-5>
- 420 Budtova T, Egal M, Gavillon R, Gericke M, Heinze T, Liebert T, Roy C, Schlufter K, Navard P (2010)
421 Comparison of Solution-State Properties of Cellulose Dissolved in NaOH/Water and in Ionic
422 Liquid (EMIMAc). *ACS Symp Ser* 1033:179-196. <https://doi.org/10.1021/bk-2010-1033.ch010>
- 423 Budtova T, Navard P (2016) Cellulose in NaOH-water based solvents: a review. *Cellulose* 23:5-55.
424 <https://doi.org/10.1007/s10570-015-0779-8>
- 425 Chen F, Sawada D, Hummel M, Sixta H, Budtova T (2020) Swelling and dissolution kinetics of natural
426 and man-made cellulose fibres in solvent power tuned ionic liquid. *Cellulose* 27:7399-7415.
427 <https://doi.org/10.1007/s10570-020-03312-5>
- 428 Cuissinat C, Navard P (2006) Swelling and Dissolution of Cellulose Part II: Free Floating Cotton and
429 Wood Fibres in NaOH-Water-Additives systems. *Macromol Symp* 244:19-30.

- 430 <https://doi.org/10.1002/masy.200651202>
- 431 Cuissinat C, Navard P (2008) Swelling and dissolution of cellulose, Part III: Plant fibres in aqueous
432 system. *Cellulose* 15:67-74. <https://doi.org/10.1007/s10570-007-9158-4>
- 433 Dochia M, Roskwitalski Z (2012) Cotton fibres. In: Kozłowski RM (ed) *Handbook of Natural Fibres*, Vol.
434 1. Woodhead Publishing, Cambridge, UK, pp 11-23
- 435 Foress KG, Fremer KE (2003) The nature and reactions of lignin - a new paradigm. Oy Nord Print Ab,
436 Helsinki, pp 128
- 437 Gross AS, Chu JW (2010) On the molecular origins of biomass recalcitrance: The interaction network
438 and solvation structures of cellulose microfibrils. *J Phys Chem B* 114:13333-13341.
439 <https://doi.org/10.1021/jp106452m>
- 440 Heinze T, Koschella A (2005) Solvents applied in the field of cellulose chemistry- a mini review.
441 *Polimeros* 15:84-90. <https://doi.org/10.1590/S0104-14282005000200005>
- 442 Isogai A, Atalla R (1998) Dissolution of cellulose in aqueous NaOH solutions. *Cellulose* 5:309-319.
443 <https://doi.org/10.1023/A:1009272632367>
- 444 Kihlman M, Aldaeus F, Chedid F, Germgård U (2012) Effect of various pulp properties on the solubility
445 of cellulose in sodium hydroxide solutions. *Holzforschung* 66:601-606.
446 <https://doi.org/10.1515/hf-2011-0220>
- 447 Kihlman M, Medronho B, Romano A, Germgård U, Lindman B (2013) Cellulose dissolution in an Alkali
448 based solvent: Influence of additives and pretreatments. *J Braz Chem Soc* 24:295-303.
449 <http://dx.doi.org/10.5935/0103-5053.20130038>
- 450 Kleman-Leyer K, Agosin E, Conner AH, Kirk TK (1992) Changes in molecular size distribution of
451 cellulose during attack by white rot and brown rot fungi. *Appl Environ Microbiol* 58:1266-1270.
452 <https://doi.org/10.1128/aem.58.4.1266-1270.1992>
- 453 Klemm D, Heublein B, Fink HP, Bohn A (2005) Cellulose: fascinating biopolymer and sustainable raw
454 material. *Angew Chem Int Ed* 44:3358-3393. <https://doi.org/10.1002/anie.200460587>
- 455 Kreze T, Malej S (2003) Structural characteristics of new and conventional regenerated cellulosic
456 fibres. *Text Res J* 73:675-684. <https://doi.org/10.1177/004051750307300804>
- 457 Le Moigne N (2008) Swelling and dissolution mechanisms of cellulose fibres. Doctoral dissertation,
458 École Nationale Supérieure des Mines de Paris, pp 9-32
- 459 Le Moigne N, Jardeby K, Navard P (2010) Structural changes and alkaline solubility of wood cellulose
460 fibres after enzymatic peeling treatment. *Carbohydr Polym* 79:325-332.
461 <https://doi.org/10.1016/j.carbpol.2009.08.009>
- 462 Liebert T (2010) Cellulose solvents- remarkable history, bright future. *Cellulose Solvents: For Analysis,*
463 *Shaping and Chemical Modification.* Publisher: Am Chem Soc Washington DC, pp 3-54.
464 <https://doi.org/10.1021/bk-2010-1033.ch001>
- 465 Mäkelä V, Wahlström R, Holopainen-Mantila U, Kilpeläinen I, King A (2018) Clustered Single Cellulosic
466 Fibre Dissolution Kinetics and Mechanisms through Optical Microscopy under Limited
467 Dissolving Conditions. *Biomacromolecules* 19:1635-1645.
468 <https://doi.org/10.1021/acs.biomac.7b01797>
- 469 Medronho G, Lindman B (2014) Competing forces during cellulose dissolution: From solvents to
470 mechanisms. *Curr Opin Colloid Interface Sci* 19:32-40.
471 <https://doi.org/10.1016/j.cocis.2013.12.001>

- 472 Moryganov AP, Zavadskii AE, Stokozenko VG (2018) Special features of X-ray analysis of cellulose
473 crystallinity and content in flax fibres. *Fibre Chem* 49:382-387. <https://doi.org/10.1007/s10692->
474 018-9904-4
- 475 Mozdyniewicz DJ, Nieminen K, Sixta H (2013) Alkaline steeping of dissolving pulp. Part I: cellulose
476 degradation kinetics. *Cellulose* 20:1437-1451. <https://doi.org/10.1007/s10570-013-9926-2>
- 477 Nishiyama Y, Langan P, Chanzy H (2002) Crystal structure and hydrogen-bonding system in cellulose
478 I β from synchrotron X-ray and neutron fibre diffraction. *Am Chem Soc* 124:9074-9082.
479 <https://doi.org/10.1021/ja0257319>
- 480 Nishiyama Y, Sugiyama J, Chanzy H, Langan P (2003) Crystal Structure and Hydrogen Bonding System
481 in Cellulose I α from Synchrotron X-ray and Neutron Fibre Diffraction. *Am Chem So.* 125:14300-
482 14306. <https://doi.org/10.1021/ja0257319>
- 483 O'Sullivan AC (1997) Cellulose: the structure slowly unravels. *Cellulose* 4:173-207.
484 <https://doi.org/10.1023/A:1018431705579>
- 485 Olsson C, Idström A, Nordstierna L, Westman G (2014) Influence of water on swelling and dissolution
486 of cellulose in 1-ethyl-3-methylimidazolium acetate. *Carbohydr Polym* 99:438-446.
487 <https://doi.org/10.1016/j.carbpol.2013.08.042>
- 488 Olsson C, Westman G (2013) Direct dissolution of cellulose: background, means and applications.
489 *Cellulose - Fundamental Aspects*. Publisher: InTech., Rijeka, pp. 143-178.
490 <http://dx.doi.org/10.5772/52144>
- 491 Palme A, Idström A, Nordstierna L, Brelid H (2014) Chemical and ultrastructural changes in cotton
492 cellulose induced by laundering and textile use. *Cellulose* 21:4681-4691.
493 <https://doi.org/10.1007/s10570-014-0434-9>
- 494 Peng H, Dai G, Wang S, Xu H (2017) The evolution behavior and dissolution mechanism of cellulose in
495 aqueous solvent. *J Mol Liq* 241:959-966. <https://doi.org/10.1016/j.molliq.2017.06.103>
- 496 Placet V, Trivaudey F, Cisse O, Gucheret-Retel V, Boubakar ML (2012) Diameter dependence of the
497 apparent tensile modulus of hemp fibres: a morphological, structural or ultrastructural effect?
498 *Compos Part A Appl Sci Manuf* 43:275-87. <https://doi.org/10.1016/j.compositesa.2011.10.019>
- 499 Rojo E, Virginia Alonso M, Domínguez JC, Saz-Orozco BD, Oliet M, Rodriguez F (2013) Alkali treatment
500 of viscose cellulosic fibres from eucalyptus wood: Structural, morphological, and thermal
501 analysis. *J Appl Polym Sci* 130:2198-2204. <https://doi.org/10.1002/app.39399>
- 502 Rous MA, Ingolic E, Schuster KC (2006) Visualisation of the fibrillar and pore morphology of cellulosic
503 fibres applying transmission electron microscopy. *Cellulose* 13:411-419.
504 <https://doi.org/10.1007/s10570-006-9052-5>
- 505 Rous MA, Varga K, Bechtold T, Schuster KC (2007) A new method to visualize and characterize the
506 pore structure of TENCEL (Lyocell) and other man-made cellulosic fibres using a fluorescent dye
507 molecular probe. *J Appl Polym Sci* 106:2083-2091. <https://doi.org/10.1002/app.26722>
- 508 Schuster KC, Aldred P, Villa M, Baron M, Loidl R, Biganska O, Patlazhan S, Vavard P, Rűf H, Jericha E
509 (2003) Characterising the emerging Lyocell fibres structures by ultra small angle neutron
510 scattering (USANS). *Lenzinger Berichte* 82:107-117
- 511 Sen S, Martin J, Argyropoulos D (2013) Review of cellulose non-derivatizing solvent interactions with
512 emphasis on activity in inorganic molten salt hydrates. *ACS Sustain Chem Eng* 1:858-870.
513 <https://doi.org/10.1021/sc400085a>
- 514 Sixta H, Michud A, Haura L, Asaadi S, Ma Y, King A, Kilpeläinen I, Hummel M (2015) Ioncell-F: a high-
515 strength regenerated cellulose fibre. *Nord Pulp Pap Res J* 30:43-57.

- 516 <https://doi.org/10.3183/npprj-2015-30-01-p043-057>
- 517 Timpa JD, Ramey HH (1989) Molecular characterization of three cotton varieties. *Text Res J* 59:661-
518 664. <https://doi.org/10.1177/004051758905901105>
- 519 Wilkes A (2011) The viscose process. In: Woodings C (ed) *Regenerated cellulose fibres Vol. 1. The*
520 *Textile Institute, Woodhead Publishing Limited, Cambridge, pp. 37-61*
- 521 Zhang S, Wang W, Li F, Yu J (2013) Swelling and dissolution of cellulose in NaOH aqueous solvent
522 systems. *Cellul Chem Technol* 47:671-679
- 523