

# Chronic HBV Infection Associates with Improvement of Lipid Profile and Steatosis-Related Impairment in Nonalcoholic Fatty Liver Disease: A Lipidomic Analysis

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## Research

**Keywords:** Lipidomics, nonalcoholic fatty liver disease, chronic hepatitis B virus infection, lipids metabolism

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# Abstract

## Background

Chronic hepatitis B virus (HBV) infection exerts an impact on lipid metabolism, but its interaction with dysmetabolism-based nonalcoholic fatty liver disease (NAFLD) remains uncertain. The present study performs lipidomic and pathological comparison in NAFLD patients with or without chronic HBV infection so as to highlight its effects on lipid metabolism and metabolic liver disease.

## Methods

Biopsy-proven Chinese NAFLD patients with (NAFLD-HBV group, n=21) or without chronic HBV infection (NAFLD group, n=41) were enrolled in the case-control study. Their serum lipid profile was subjected to individual investigation by ultra-performance liquid chromatography–tandem mass spectrometry (UPLC-MS/MS). Steatosis, activity, and fibrosis (SAF) scoring revealed the NAFLD-specific pathological characteristics. The effects of chronic HBV infection on demographic and clinical manifestations, serum lipidomics, and pathological characteristics were statistically assessed.

## Results

Chronic HBV infection associated with global alteration of serum lipid profile in NAFLD patients. Upregulation of phosphatidylcholine (PCs), choline plasmalogen (PC-Os) and downregulation of free fatty acids (FFAs), lysophosphatidylcholine (LPCs) dominated the HBV-related lipidomic characteristics. Compared to those of NAFLD group, the levels of serum hepatotoxic lipids (FFAs: FFA16:0, FFA16: 1, FFA18:1, FFA18:2) were significantly lowered in the NAFLD-HBV group. These low-level FFAs demonstrated correlation to statistical improvements in aspartate aminotransferase activity (FFA16:0,  $r = 0.34$ ,  $P = 0.003$ ; FFA16:1,  $r = 0.37$ ,  $P < 0.001$ ; FFA18:1,  $r = 0.33$ ,  $P < 0.001$ ; FFA18:2,  $r = 0.42$ ,  $P < 0.001$ ), hepatocyte steatosis (FFA16: 1,  $r = 0.39$ ,  $P = 0.01$ ; FFA18:1,  $r = 0.39$ ,  $P = 0.015$ ; FFA18:2,  $r = 0.33$ ,  $P = 0.036$ ), and ballooning (FFA16:0,  $r = 0.31$ ,  $P = 0.037$ ; FFA16:1,  $r = 0.45$ ,  $P < 0.001$ ; FFA18:1,  $r = 0.37$ ,  $P = 0.005$ ; FFA18:2,  $r = 0.30$ ,  $P = 0.013$ ).

## Conclusion

Chronic HBV infection may present improvements in the serum lipidomics and steatosis-related pathological characteristics of NAFLD.

## Trial registration

Trial registration number: ChiCTR-DDT-13003983. Registered 13 May, 2013, <http://www.chictr.org.cn/enIndex.aspx?proj=5584>.

## Introduction

Chronic hepatitis B virus (HBV) reflects a worldwide chronic liver disease that affects over 250 million people, especially in the Chinese population [1, 2]. By the studies published to date, there is an inverse association of viral indices (*e.g.*, HBsAg, HBV-DNA) and incidence of hyperlipidemia (*e.g.*, hypertriglyceridemia, hypercholesterolemia), fatty liver and metabolic syndrome (MetS)[3, 4]. Thus, chronic HBV infection is proposed to exert a beneficial impact on lipid metabolism, probably on the basis of HBV-host interaction[5–8].

Recent decades have witnessed the rapid growing incidence of nonalcoholic fatty liver disease (NAFLD), a metabolic stress-induced chronic liver disease, in China by the prevalence of overweight/obesity and/or sedentary lifestyle[9, 10]. In result, concurrent NAFLD is now identified in 13.5% (12/ 91) and 14% (260/ 1915) chronic hepatitis B infection patients from Hong Kong (China) and Hang Zhou (Zhe Jiang, China), respectively[7, 11].

Serving as one of the components of MetS, NAFLD has been well described to demonstrate an intimate association with abnormalities in systemic lipid metabolism. The individuals with NAFLD exhibit a strong positive association with hyperlipidemia[12]. While patients with hypertriglyceridemia are independently predisposed to the risk of NAFLD[13]. Given the reciprocal causation of NAFLD and hyperlipidemia, they are supposed to be affected by the concurrent HBV infection. However, the role of chronic HBV infection in NAFLD and related lipometabolic disorders remains uncertain until now.

We, therefore, recruited the biopsy-proven Chinese NAFLD patients with or without chronic HBV infection. Ultra-performance liquid chromatography–tandem mass spectrometry (UPLC-MS/MS) was then employed to investigate their difference in serum lipidomics. Integrated analysis of demographic and clinical manifestations, lipid profiles, and hepatic pathological characteristics further revealed the effect of chronic HBV infection on lipid metabolism, hepatic steatosis and related impairments of NAFLD patients.

## Materials And Methods

### Patients

A total of 62 NAFLD patients with (NAFLD-HBV group, n = 21) or without chronic HBV infection (NAFLD group, n = 41) were enrolled from the inpatients of Xinhua Hospital, Shanghai during May 2012 to May 2014. Subjects with ongoing or recent alcohol abuse (alcohol intake > 20 g/day for male, > 10 g/day for female), anti-HCV IgG/IgM positive, autoimmune hepatitis, drug-induced liver injury, primary biliary cholangitis, Wilson's disease and other causes of liver steatosis were excluded. Each participant of the study was exposed to pathological evaluation by liver biopsy. Patients with hepatocyte steatosis (> 5%) were diagnosed to be NAFLD, and those with seropositivity for hepatitis B surface antigen (HBsAg) for at least six months were defined as chronic HBV infection (Guidelines for the Prevention, Care and Treatment of Persons with Chronic Hepatitis B Infection, 2015)[14]. This study was approved by the Research Ethics Committee of Xinhua Hospital, and informed consent was obtained from each patient.

## Clinical and laboratory assessment

Demographical characteristics including age, gender, height, weight, waist-to-hip ratio and body mass index (BMI) were obtained from medical record. Blood samples were collected from each patient after 12-h fasting. Alanine aminotransferase (ALT), aspartate aminotransferase (AST), gamma-glutamyl transferase ( $\gamma$ -GT) and alkaline phosphatase (ALP) test were performed by a multichannel automatic analyzer (Bayer ADVIA 1650, Moss, Norway). Fasting plasma glucose (FPG), triacylglycerol (TG) and total cholesterol (TC) were measured using Wako Bioproducts (Wako Pure Chemical Industries, Richmond, VA, USA). HBV infection assays were routinely performed to measure hepatitis B surface antigen (HBsAg), hepatitis B e antigen (HBeAg) and antibody against HBeAg (anti-HBe) by Enzyme-linked immunosorbent assay (Abbott Laboratories, North Chicago, IL, USA). The HBV DNA levels were detected by real-time polymerase chain reaction assay with a detection limit of 100 IU/mL.

## Hepatic histopathological examination

Each liver biopsy sample was reviewed by three pathologists who were blinded to the present study. The steatosis, activity, and fibrosis (SAF) scoring system was employed for the evaluation of NAFLD [15, 16]. In detail, the SAF score was assessed on the basis of Steatosis (S0, < 5%; S1, 5–33%; S2, 34–66%; S3, > 66%), Activity (sum of lobular inflammation: 0, no foci per 200  $\times$  field; 1, < 2 foci per 200  $\times$  field; 2, 2–4 foci per 200  $\times$  field; 3, > 4 foci per 200  $\times$  field), ballooning (0, none; 1, few balloon cells; 2, many cells/prominent ballooning), and Fibrosis (F0, none; F1, perisinusoidal or portal fibrosis; F2, perisinusoidal and periportal fibrosis without bridging; F3, bridging fibrosis; F4, cirrhosis). Chronic hepatitis B infection was defined by the typical periportal/portal hepatitis with piecemeal necrosis of hepatocytes.

## UPLC-MS/MS

The serum lipidomics were analyzed as described previously [17]. In brief, lipids were extracted from the collected serum samples, and analyzed by UPLC (Waters, Milford, USA) combined with a triple TOF 5600 mass spectrometer (AB SCIEX, USA) platform together with thirteen quality control (QC) samples. Lipids separation was performed using a UPLC ACQUITY C8 BEN column (2.1  $\times$  100 mm; 1.7  $\mu$ m; Waters, Milford, USA). The mobile phases consisted of (A) 60% acetonitrile in water, 10 mmol/L ammonium acetate, and (B) 90% isopropanol in acetonitrile, 10 mmol/L ammonium acetate. Gradient elution was carried out at a flow rate of 0.26 mL/min with the gradient conditions as follows: 0–1.5 min, 32% B; 1.5–14 min, 32–85% B; 15.5–15.6 min, 85–97% B; 15.6–18 min, 97% B; 18–20 min, 97–32% B. Mass spectrometry was performed in positive and negative electrospray ion modes. Data acquisitions were applied using Analyst TF 1.6 software (AB SCIEX, Framingham, MA). LipidView/PeakView and MultiQuant 2.0 (AB SCIEX, Framingham, MA) were used for lipid identification and quantification, respectively. After being normalized with corresponding internal standards, the detected lipids data in QCs were evaluated based on their relative standard deviation (RSD), and only those with RSD below 30% were subjected to further analysis.

## Statistical analysis

Statistical analyses were performed using SPSS Statistics software version 23.0 and R software version 4.0.2. The continuous variables were presented as mean  $\pm$  SD or median (interquartile range), and were subjected to comparison using unpaired Student's t test. Multivariate analysis including principal component analysis (PCA) was performed using R 4.0.2, and orthogonal partial least squares-discriminant analysis (OPLS-DA) was performed using SIMCA 14.1 (MKS Umetrics, Malmö, Sweden). The differential serum lipids with both multivariate and univariate significance (OPLS-DA VIP  $>$  1.0 and  $P <$  0.05) were filtered on the basis of variable importance in the projection (VIP), S-plot and  $P$  value (unpaired Student's t-test). Spearman's correlation was used to exploring the correlativity between serum lipid profile and hepatic pathological parameters. Statistical significance was defined as a two-side  $P$  value  $<$  0.05.

## Results

### Chronic HBV infection exhibited beneficial effects on BMI, hepatocyte steatosis and related impairments

In the present study, demographic, clinical and pathological indices were compared between NAFLD patients with (NAFLD-HBV group) or without chronic HBV infection (NAFLD group). Interestingly, the NAFLD-HBV group presented the BMI much lower than that of NAFLD group (NAFLD group vs NAFLD-HBV group:  $27.4 \pm 3.2$  vs  $25.7 \pm 2.6$ ,  $P = 0.023$ ) (Table 1). Significant amelioration of NAFLD-specific pathological characteristics, including hepatocyte steatosis (NAFLD group vs NAFLD-HBV group:  $2.32 \pm 0.65$  vs  $1.33 \pm 0.49$ ,  $P < 0.001$ ) and ballooning (NAFLD group vs NAFLD-HBV group:  $1.84 \pm 0.37$  vs  $1.13 \pm 0.52$ ,  $P < 0.001$ ), was documented in the NAFLD-HBV instead of NAFLD group (Table 1). In contrast to the comparability in most biochemical indices, there was a statistically decreased ALT activity in the NAFLD-HBV group in comparison to that of the NAFLD group (NAFLD group vs NAFLD-HBV group: 64.1 IU/L (39.5 IU/L, 110.3 IU/L) vs 53 IU/L (23.4 IU/L, 69.5 IU/L),  $P = 0.025$ ) (Table 1). Thus, the beneficial impact of chronic HBV infection, especially on the lipid metabolism and related hepatic injury, was indicated in the NAFLD patients.

Table 1  
Demographic, clinical and pathological data of all patients

Variable	NAFLD (n = 41)	NAFLD-HBV (n = 21)	P
Male, n(%)	26 (63.4)	15 (71.4)	0.528
Age (years)	40 ± 14	37 ± 14	0.439
Body mass index (kg/m <sup>2</sup> )	27.4 ± 3.2	25.7 ± 2.6	0.023 *
Waist-to-hip ratio	0.93 ± 0.05	0.92 ± 0.07	0.569
Total bilirubin (µmol/L)	17.3 ± 15.6	14.2 ± 4.3	0.379
Alkaline phosphatase (IU/L)	84.6 (62.3, 100.7)	78 (63, 96.4)	0.693
Alanine aminotransferase (IU/L)	64.1 (39.5, 110.3)	53 (23.4, 69.5)	0.025 *
Aspartate aminotransferase (IU/L)	37.7 (25, 65.4)	28.6(20.3, 45.8)	0.14
Gamma-glutamyl transferase (IU/L)	60.6 (38, 91.1)	31 (22.2, 60)	0.058
Fasting glucose (mmol/L)	5.4 (4.6, 6.5)	4.8 (4.7, 5.5)	0.797
Total cholesterol (mmol/L)	4.5 (4.3, 5.2)	5.2 (4.1, 5.7)	0.322
HDL-cholesterol (mmol/L)	1.2 (1.1, 1.3)	1.2 (1, 1.6)	0.451
LDL-cholesterol (mmol/L)	2.9 (2.6, 3.2)	2.8 (2, 3.5)	0.7
Total triglycerides (mmol/L)	1.6 (1.1, 2.4)	1.2 (1, 2.2)	0.612
Hepatocyte steatosis	2.32 ± 0.65	1.33 ± 0.49	< 0.001 **
Lobular inflammation	0.65 ± 0.66	0.67 ± 0.62	0.915
Ballooning	1.84 ± 0.37	1.13 ± 0.52	< 0.001 **
Liver fibrosis	1.68 ± 1.14	1.13 ± 1.13	0.134
The continuous variables were presented as mean ± SD or median (interquartile range). HDL, high density lipoprotein; LDL, low density lipoprotein. * <i>P</i> < 0.05, ** <i>P</i> < 0.01.			

### Serum lipidomics differentiated NAFLD patients with or without chronic HBV infection

Multivariate analyses were employed in our study to take an overview of the serum lipidomics between NAFLD and NAFLD-CHB groups. Dramatically, 3D PCA score plot of serum lipidomics distinctly differentiated the NAFLD patients with or without chronic HBV infection (Fig. 1A). Similar group discrimination was also obtained by the OPLS-DA score plot (Fig. 1B).

To reveal the role of chronic HBV infection in lipid metabolism, a total of 239 serum lipids was exposed to UPLC-MS/MS in both NAFLD and NAFLD-HBV groups. In result, 64 lipids among these ones (26.78%) were

filtered to be statistically different by unpaired Student's t-test. Detailedly, the profile of differential serum lipids comprised 17 free fatty acid (FFAs), 8 lysophosphatidylcholine (LPCs), 3 lysophosphatidylcholine plasmalogen (LPC-Os), 1 lysophosphatidylethanolamine (LPE), 2 lysophosphatidylethanolamine plasmalogen (LPE-Os), 2 lysophosphatidylinositol (LPIs), 3 phosphatidylcholine (PCs), 18 cholineplasmalogen (PC-Os), 5 phosphatidylethanolamine (PEs), 2 ethanolamine plasmalogen (PE-Os), 1 phosphatidylinositol (PI) and 2 sphingomyelin (SMs) (Table 2). On the other hand, S-plot put forward 31 differential serum lipids with VIP > 1.0, including 7 free fatty acid (FFAs), 3 lysophosphatidylcholine (LPCs), 10 phosphatidylcholine (PCs), 8 cholineplasmalogen (PC-Os), 1 sphingomyelin (SM) and 2 triacylglycerol (TGs) (Fig. 2).

Table 2

Differential serum lipids between NAFLD patients with or without chronic HBV infection by unpaired Student's *t* test

Lipids	NAFLD-HBV/NAFLD	<i>P</i>	Lipids	NAFLD-HBV/NAFLD	<i>P</i>	Lipids	NAFLD-HBV/NAFLD	<i>P</i>
FFA 12:0	0.81	0.021	LPC 20:3	0.67	0.016	PC-O 36:5	1.33	0.002
FFA 14:0	0.70	0.002	LPC 22:6	0.63	0.001	PC-O 38:4	1.31	< 0.001
FFA 14:1	0.59	0.009	LPC 24:0	1.35	0.014	PC-O 38:5	1.35	0.001
FFA 16:0	0.75	0.001	LPC-O 16:0	0.68	0.008	PC-O 40:4	1.40	< 0.001
FFA16:1	0.61	0.001	LPC-O 16:1	0.70	0.008	PC-O 40:5	1.37	0.001
FFA 18:1	0.67	< 0.001	LPC-O 18:1	0.68	0.024	PC-O 42:4	1.38	< 0.001
FFA18:2	0.61	< 0.001	LPE 22:6	0.83	0.047	PC-O 42:5	1.45	< 0.001
FFA 18:3	0.58	< 0.001	LPE-O 16:1	0.56	0.006	PC-O 42:6	1.42	0.001
FFA 20:1	0.66	0.003	LPE-O 18:1	0.62	0.019	PC-O 44:5	1.42	0.001
FFA 20:2	0.59	< 0.001	LPI 18:0	0.30	0.004	PC-O 44:6	1.48	0.001
FFA 20:3	0.44	0.001	LPI 20:4	0.66	< 0.001	PE 34:1	1.47	0.016
FFA20:4	0.44	0.002	PC 34:2	1.16	0.049	PE 34:2	1.43	0.012
FFA20:5	0.20	0.012	PC 34:3	1.25	0.043	PE 36:2	1.33	0.044
FFA 22:2	0.59	0.043	PC 36:2	1.25	0.029	PE 36:3	1.49	0.009
FFA 22:4	0.63	0.004	PC-O 32:0	1.28	0.001	PE 38:5	1.28	0.021

The serum lipids are expressed in a pattern of name carbon numbers: double bond numbers. FFA, free fatty acids; LPC, lysophosphatidylcholine; LPC-O, lysophosphatidylcholine plasmalogen; LPE, lysophosphatidylethanolamine; LPI, lysophosphatidylinositol; PC, phosphatidylcholine; PC-O, choline plasmalogen; PE, phosphatidylethanolamine; PI, phosphatidylinositol; SM, sphingomyelin.



Lipids	NAFLD-HBV/NAFLD	<i>P</i>	Lipids	NAFLD-HBV/NAFLD	<i>P</i>	Lipids	NAFLD-HBV/NAFLD	<i>P</i>
FFA 22:5	0.50	< 0.001	PC-O 34:0	1.25	0.003	PE-O 34:3	1.28	0.017
FFA 22:6	0.41	0.001	PC-O 34:1	1.28	0.001	PE-O 36:3	1.29	0.017
LPC 16:0	0.72	0.004	PC-O 34:2	1.29	0.031	PI 34:1	1.30	0.035
LPC 17:0	0.68	0.007	PC-O 34:3	1.34	0.003	SM 34:0:3	1.26	0.014
LPC 18:0	0.73	0.018	PC-O 36:2	1.27	0.003	SM 36:0:2	0.66	0.005
LPC 18:3	0.70	0.003	PC-O 36:3	1.28	0.012			
LPC 20:2	0.64	0.008	PC-O 36:4	1.31	0.012			

The serum lipids are expressed in a pattern of name carbon numbers: double bond numbers. FFA, free fatty acids; LPC, lysophosphatidylcholine; LPC-O, lysophosphatidylcholine plasmalogen; LPE, lysophosphatidylethanolamine; LPI, lysophosphatidylinositol; PC, phosphatidylcholine; PC-O, choline plasmalogen; PE, phosphatidylethanolamine; PI, phosphatidylinositol; SM, sphingomyelin.

### FFAs, LPCs, PCs and PC-Os alteration characterized the effects of chronic HBV infection on serum lipidomics

Integrating unpaired Student's t-test and S-plot, 17 differential serum lipids with  $P < 0.05$  and  $VIP > 1.0$  were identified to characterize the lipidomics of NAFLD patients upon chronic HBV infection. They were classified into FFAs (FFA 16:0, FFA 16:1, FFA 18:1, FFA 18:2, FFA 20:4, FFA 22:6), LPCs (LPC 16:0, LPC 18:0, LPC 18:3), PCs (PC 34:2, PC 36:2), and PC-Os (PC-O 34:2, PC-O 34:3, PC-O 36:4, PC-O 36:5, PC-O 38:4, PC-O 38:5), respectively (Table 3). When compared to those of the NAFLD group, serum PCs (NAFLD-HBV/NAFLD: 1.21–1.25) and PC-Os levels (NAFLD-HBV/NAFLD: 1.29–1.35) in the NAFLD-HBV group exhibited significant upregulation (Table 3, Fig. 3). Contrastively, serum levels of FFAs (NAFLD-HBV/NAFLD: 0.41–0.75) and LPCs (NAFLD-HBV/NAFLD: 0.70–0.73) experienced statistical downregulation in the NAFLD patients with concurrent chronic HBV infection (Table 3, Fig. 3). These characteristics convinced the dominating role of FFAs, LPCs, PCs and PC-Os in differential lipid profile.

Table 3  
Differential serum lipids between NAFLD and NAFLD-HBV groups ( $P < 0.05$ , VIP  $> 1$ )

lipids	Ratio of concentration (NAFLD-HBV/NAFLD)			
	Fold	Trend	P	VIP
FFA16:0	0.75	↓	0.010	2.85
FFA16:1	0.61	↓	0.005	1.18
FFA18:1	0.67	↓	0.001	2.98
FFA18:2	0.61	↓	0.001	2.93
FFA20:4	0.44	↓	0.022	1.14
FFA22:6	0.41	↓	0.015	1.10
LPC16:0	0.72	↓	0.014	3.80
LPC18:0	0.73	↓	0.018	1.85
LPC18:3	0.70	↓	0.013	1.47
PC34:2	1.21	↑	0.049	7.76
PC36:2	1.25	↑	0.029	7.37
PC-O34:2	1.29	↑	0.009	1.23
PC-O34:3	1.34	↑	0.001	1.32
PC-O36:4	1.31	↑	0.002	1.97
PC-O36:5	1.33	↑	0.002	1.41
PC-O38:4	1.31	↑	$< 0.001$	1.15
PC-O38:5	1.35	↑	$< 0.001$	1.74

### Low-level FFAs upon chronic HBV infection associated with improvements in hepatocyte steatosis and related impairments

Correlation between differential serum lipids (FFAs, LPCs, PCs and PC-Os) and biochemical indices (TBIL, DBIL, ALP,  $\gamma$ -GT, ALT and AST), together with NAFLD-specific pathological characteristics (hepatocyte steatosis, ballooning, lobular inflammation, fibrosis), was subjected to assessment in this study.

Noticeably, low-level FFAs among these lipids showed close association with lessened hepatocyte steatosis (FFA16: 1,  $r = 0.39$ ,  $P = 0.01$ ; FFA18:1,  $r = 0.39$ ,  $P = 0.015$ ; FFA18:2,  $r = 0.33$ ,  $P = 0.036$ ) and ballooning (FFA16:0,  $r = 0.31$ ,  $P = 0.037$ ; FFA16:1,  $r = 0.45$ ,  $P < 0.001$ ; FFA18:1,  $r = 0.37$ ,  $P = 0.005$ ; FFA18:2,  $r = 0.30$ ,  $P = 0.013$ ) (Fig. 4). Furthermore, the reduction of ALT (FFA16: 0,  $r = 0.16$ ,  $P = 0.043$ ; FFA16: 1,  $r =$

0.05,  $P=0.02$ ; FFA18:1,  $r=0.16$ ,  $P=0.004$ ; FFA18:2,  $r=0.13$ ,  $P=0.017$ ) and AST activities (FFA16: 0,  $r=0.34$ ,  $P=0.003$ ; FFA16: 1,  $r=0.37$ ,  $P<0.001$ ; FFA18:1,  $r=0.33$ ,  $P<0.001$ ; FFA18:2,  $r=0.42$ ,  $P<0.001$ ) in patients with low-level FFAs reflected an attenuation of steatosis-related hepatic impairments (Fig. 4). The down-regulatory effect of chronic HBV infection on FFAs, and the association of low-level FFAs and steatosis improvements, resultantly convinced a beneficial role of chronic HBV infection in the lipid profile and related NAFLD.

## Discussion

Nowadays, accumulating proofs shed light on the fact that chronic HBV infection deeply involves in the metabolic profiles and, subsequently, affects multiple components of MetS[3–8, 18, 19]. In contrast to the uninfected control subjects, those with chronic HBV infection demonstrate a decreased prevalence of hypertriglyceridemia and lowered level of serum TG[3, 6]. Whereas hypertriglyceridemia inversely associates with the viral load in HBeAg seropositives[19]. There is also a negative correlation between HBV viral load (HBV-DNA) and serum TG level[6]. After adjusting for demographic and metabolic factors, HBV infection is now recognized to be the independent factor associated with lower risk of NAFLD, mainly attributed to the HBV-related reduction of serum and intrahepatic TG concentration[7, 8, 19]. Some other MetS components, including hypercholesterolemia and high blood pressure, are likely to be improved in the patients with HBV seropositivity[3, 8, 20, 21]. Furthermore, both Third Korean National Health and Nutrition Examination Survey (KNHANES III) and cross-sectional population study in Hong Kong Chinese uncovered an association of HBsAg positivity and low prevalence of MetS[4, 5, 7]. X protein expression and adipokines (e.g., adiponectin) secretion may underlie the interaction of HBV and host metabolism[22–24].

Clinical trials have recently shown that NAFLD takes place on the basis of chronic HBV infection with an increasing annual prevalence[25], yet the viral impact on lipid metabolism, hepatic steatosis and related impairments remains to be explored. In the present study, we identified 26.78% (64/239) differential lipids in the serum lipidomics of NAFLD-HBV group in comparison to that of NAFLD group. The wide-range alterations verified by  $P<0.05$  and/or  $VIP>1.0$ , including FFAs, LPCs, LPEs, LPIs, PCs, PC-Os, PEs, PE-Os, and SMs, confirmed a global metabolic effect of chronic HBV infection in the NAFLD patients. Moreover, both 3D PCA and OPLS-DA score plots for serum lipidomics distinctly differentiated NAFLD patients with or without chronic HBV infection. Thus HBV carriers with or without concurrent NAFLD are suggested to share the HBV-based modulation of systemic lipid metabolism, with some differences in their features.

To take further insight into the HBV-specific serum lipidomics, we integrated the differential lipids obtained from Student's *t*-test and S-plot. When compared to those of the NAFLD group, upregulation of PCs, PC-Os and downregulation of FFAs, LPCs resultantly characterized the lipidomics of NAFLD-HBV group with statistical significance. Serving as inhibitors of hepatic lipogenesis, PCs induce the alleviation of orotic acid-induced rodent hepatocyte steatosis[26]. On the other hand, PCs take an essential place in the membrane integrity[27, 28]. They prevent the membrane leakage to abolish hepatocyte injury and, subsequently, lobular inflammation and liver fibrosis[29, 30]. PC-Os are a class of phospholipids that

contain a vinyl ether linkage at the sn-1 position and highly arachidonic acid at the sn-2 position. They have been reported to act as potential protector against oxidative stress[31, 32]. Contrastively, NAFLD patients demonstrate high serum level of FFAs, which are described to be cytotoxic and potential in the early diagnosis[33, 34]. LPCs, a kind of lipid intermediate elevated in rodent and human nonalcoholic steatohepatitis (NASH), mediate the interaction of saturated fatty acid and insulin resistance[35, 36]. Given their hepatic activities, these HBV-impacted differential lipids are conferred to interact with lipid metabolism and related pathological alterations in the liver.

We further investigate the interaction between lipid profile of PCs, PC-Os, FFAs, LPCs and NAFLD-related biochemical and pathological indices in patients with concurrent chronic HBV infection and NAFLD. In the multivariate model of our study, low-level FFAs of 16:1, 18:1, and 18:2 showed significant association with alleviated hepatocyte steatosis. Consistently, FFAs reduction was accompanied by the improvements in steatosis-based hepatic injury (ballooning, down-regulated aminotransferase activities). With their reflux from adipose tissue to the liver, FFAs lead to lipotoxicity that contributes to hepatic steatosis and related impairments[37]. FFAs reduction upon chronic HBV infection abrogates these lipotoxicity-induced abnormalities. Our study had uncovered an interesting phenomenon that was similar to previous reports[38], namely significantly lowered hepatocyte steatosis occurred in the NAFLD-HBV group as compared to that of NAFLD group. Taken together, chronic HBV infection is presented to have beneficial impact on the lipid metabolism and steatosis-related liver impairments, mainly on the basis of lipidomic improvements.

There are some limitations in this study. First, sampling bias could not be excluded for the limited number of participants in both NAFLD and NAFLD-HBV groups. Second, the non-target method of lipidomic analysis kept serum lipids from absolute quantification. Third, the position of double bonds has not been identified in various kinds of multiple unsaturated serum lipids. These limitations should be taken into consideration for an interpretation of our findings.

## Conclusion

In summary, chronic HBV infection exerts global effect on serum lipidomics of NAFLD patients. Alteration of FFAs, LPCs, PCs and PC-Os dominates the HBV-related lipidomic characteristics. Low-level FFAs (FFA16: 0, FFA16:1, FFA18:1, FFA18:2) upon chronic HBV infection demonstrate association with significant improvements in hepatocyte steatosis, ballooning, and decreased aminotransferase activities. Therefore, chronic HBV infection could be beneficial to the serum lipid profile, hepatic steatosis and related impairments in NAFLD patients.

## Abbreviations

**ALP:** Alkaline phosphatase

**ALT:** Alanine aminotransferase

**AST:** Aspartate aminotransferase

**BMI:** Body mass index

**DBIL:** Direct bilirubin

**FPG:** Fasting plasma glucose

**FFA:** Free fatty acid

**HBV:** Hepatitis B virus

**HDL:** High density lipoprotein

**LDL:** Low density lipoprotein

**LPC:** Lysophosphatidylcholine

**LPC-O:** Lysophosphatidylcholine plasmalogen

**LPE:** Lysophosphatidylethanolamine

**LPE-O:** Lysophosphatidylethanolamine plasmalogen

**LPI:** Lysophosphatidylinositol

**MetS:** Metabolic syndrome

**NAFLD:** Nonalcoholic fatty liver disease

**OPLS-DA:** Orthogonal partial least squares-discriminant analysis

**PCA:** Principal component analysis

**PC:** Phosphatidylcholine

**PC-O:** Cholineplasmalogen

**PE:** Phosphatidylethanolamine

**PE-O:** Ethanolamine plasmalogen

**PI:** Phosphatidylinositol

**SM:** Sphingomyelin

**TC:** Total cholesterol

**TG:** Triacylglycerol

**UPLC-MS/MS:** Ultra-performance liquid chromatography–tandem mass spectrometry

**VIP:** Variable importance in the projection

**γ-GT:** Gamma-glutamyl transferase

## Declarations

### Ethics approval and consent to participate

Our study was approved by the Research Ethics Committee of Xinhua Hospital. Informed contents were obtained from all the patients. The trial registration number is ChiCTR-DDT-13003983, Registered on 13 May 2013 (Retrospectively registered). Relevant information available at:

<http://www.chictr.org.cn/enIndex.aspx?proj=5584>

### Consent for publication

All authors have approved the contents of this manuscript and agreed on the manuscript be published.

### Availability of data and materials

The data will be available on request.

### Competing interests

The authors declare that they have no competing interests.

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### Authors’ contributions

HL and QYX contributed to study concept, study design, data interpretation and original draft writing. YX and JJL contributed to data collection, data analysis and data interpretation. HXC and QP contributed to resources, conceptualization, funding acquisition, supervision, writing—review and editing.

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## Figures

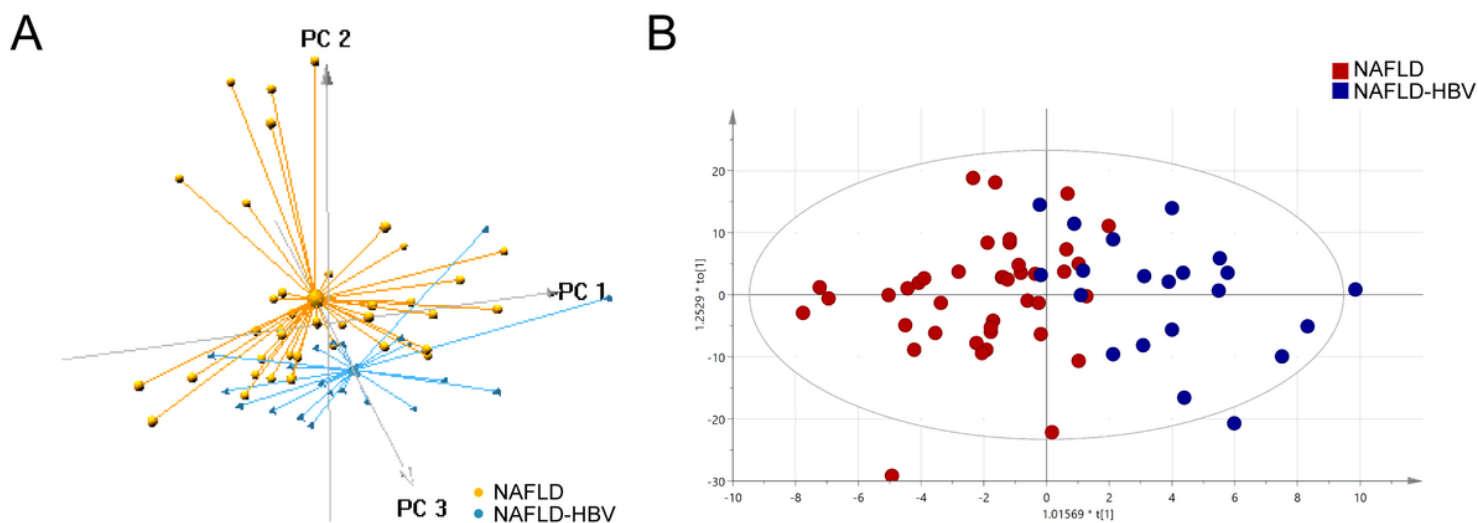
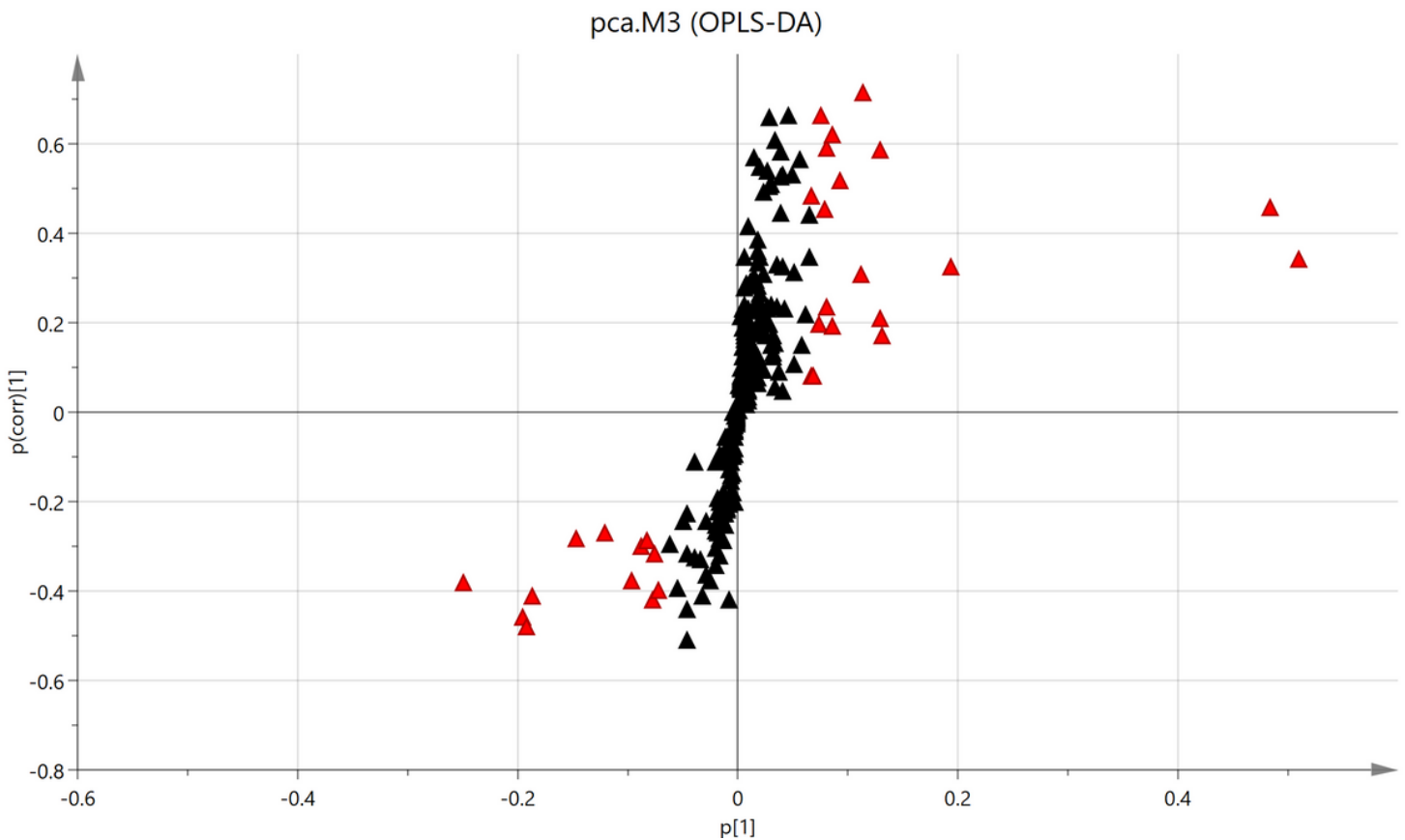


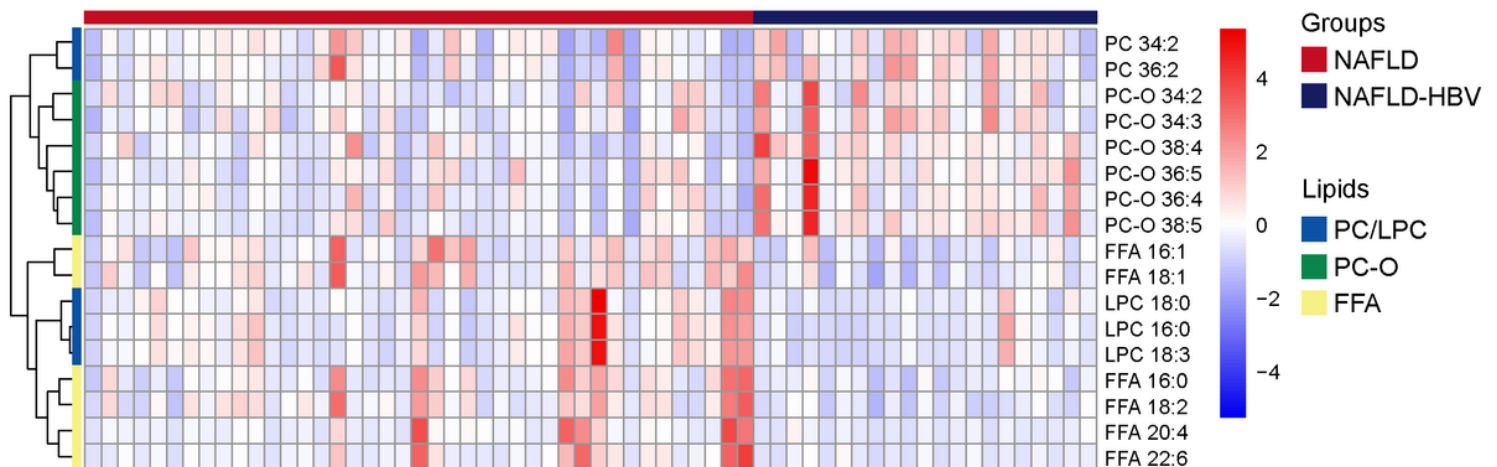
Figure 1

Lipidomics differentiates patients with or without chronic HBV infection. (A) Score plot of 3D principal component analysis (PCA) for the patients with (NAFLD-HBV group) or without chronic HBV infection (NAFLD group) [R2X(cum) = 0.562, Q2(cum) = 0.297]. (B) Score plot of orthogonal partial least squares-discriminant analysis (OPLS-DA) for the NAFLD and NAFLD-HBV groups [R2X(cum) = 0.796, Q2(cum) = 0.183].



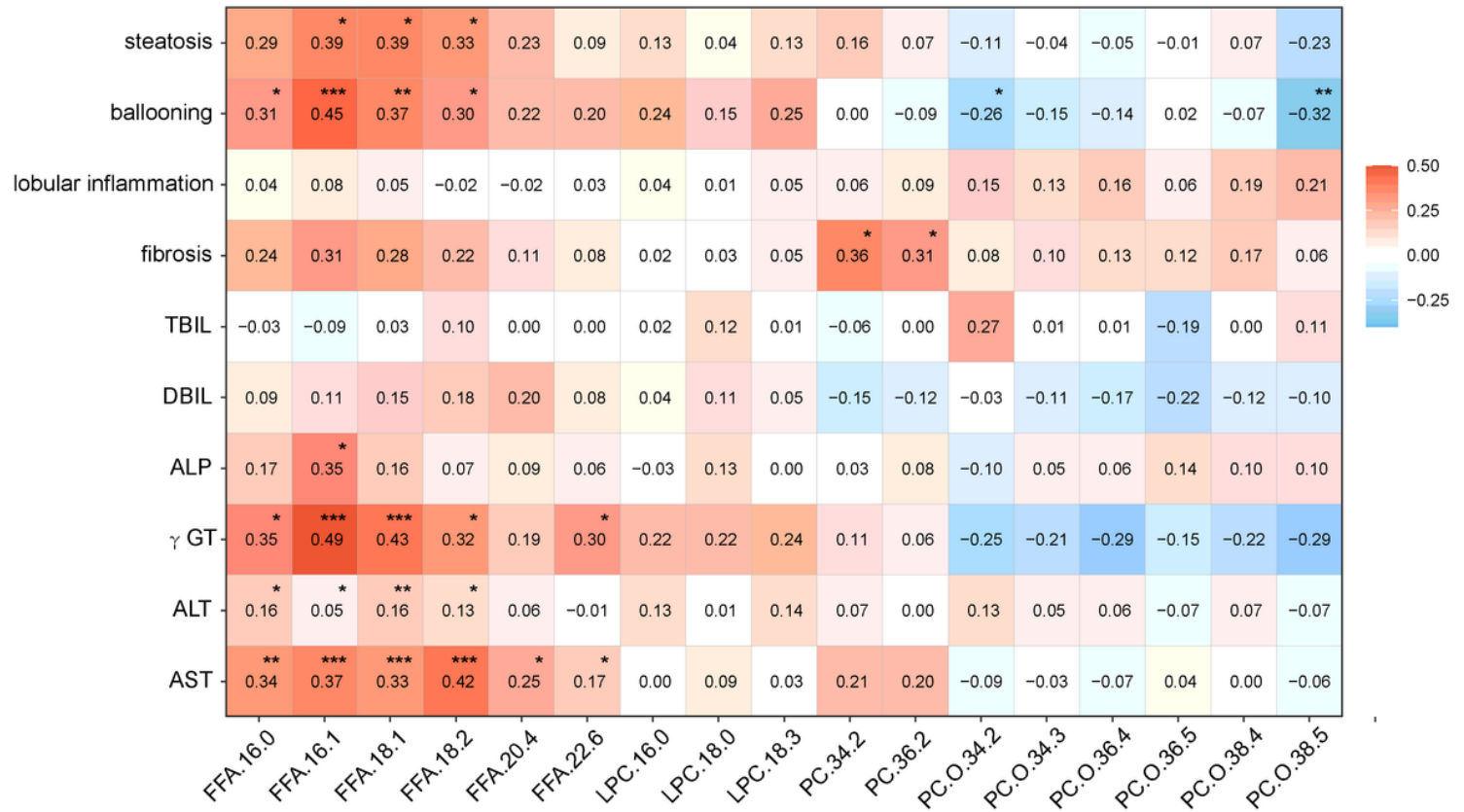
**Figure 2**

S-plot identifies the differential serum lipids between NAFLD patients with or without chronic HBV infection. Differential serum lipids with VIP > 1 are labeled in red.



**Figure 3**

Heatmap of the differential serum lipid between NAFLD and NAFLD-HBV groups. Red denotes a relative increase, and blue denotes a relative decrease.



**Figure 4**

Correlations between differential serum lipids and pathological and clinical data in NAFLD patients. TBIL, total bilirubin; DBIL, direct bilirubin; ALP, Alkaline phosphatase; γGT, Gamma-glutamyl transferase; ALT, alanine aminotransferase; AST, aspartate aminotransferase. \* P < 0.05, \*\* P < 0.01, \*\*\* P < 0.001.