

Serum iron, prealbumin, D-dimer and prothrombin time are indicators of the diagnosis, severity and outcome of *Salmonella enterica* serovar Enteritidis infection

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Research

Keywords: *Salmonella*; serovar Enteritidis; acute diarrhea; serum iron, coagulation

DOI: <https://doi.org/10.21203/rs.3.rs-60917/v1>

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Abstract

Aims *Salmonella enterica* serovar Enteritidis (*S. Enteritidis*) infection is one of the main causes of foodborne illness among individuals in the military. In this study, we aimed to investigate indicators associated with the diagnosis, severity and outcome of *S. Enteritidis* infection, and to analyze the antimicrobial susceptibility of the isolated *S. Enteritidis* strains.

Methods Routine stool test, fecal occult blood tests (FOBTs), routine blood tests and blood biochemical analyses were completed in our clinical laboratory. The stool samples were inoculated on *Salmonella Shigella* (SS) agar plates, the single bacterial colony was identified with mass spectrometry (MS). The serum agglutination test was performed to identify the *Salmonella* serotype. Antimicrobial susceptibility testing was performed using the minimal inhibitory concentration (MIC) method with the VITEK 2 COMPACT analyzer.

Results Recently, 6 patients from one company visited our department complaining of fever, watery stool, abdominal pain and so on. The patients' white blood cell (WBC) counts (66.7%), neutrophil percentages (100%), C-reactive protein (CRP) levels (100%), prothrombin times (PTs, 100%) and serum D-dimer concentrations (83.3%) were higher or longer than normal, while serum iron (100%) and prealbumin (83.3%) were lower than normal when they were admitted. Stool cultures and serovar identification results indicated *S. Enteritidis* infection. The antimicrobial susceptibility results showed that the 6 isolated bacterial strains were sensitive to trimethoprim-sulfamethoxazole and levofloxacin, and resistant to ampicillin. All patients were administered levofloxacin accompanied by rifaximin and probiotics, and their symptoms and the abovementioned parameters were recovered. During the disease course, the prolonged PT, elevated D-dimer and decreased serum iron were positively correlated with severe symptoms and serum inflammatory indexes.

Conclusion Prolonged PT, elevated D-dimer and hypoferrremia were associated with the diagnosis, severity and outcome of *S. Enteritidis* infection, and a decreased prealbumin level was associated with the diagnosis of *S. Enteritidis* infection. In patients complaining of acute fever, watery stool and abdominal pain, PT, D-dimer, prealbumin and serum iron should be monitored to attain an early diagnosis and initiate appropriate treatment for *S. Enteritidis* infection.

Background

Salmonella enterica serovar Enteritidis (*S. Enteritidis*) infection is one of leading infectious causes of foodborne illness worldwide[1]. Worldwide, 94 million patients with gastroenteritis are diagnosed with *Salmonella* infection, 155,000 of whom die every year[2, 3], and it has been a huge public health issue for both civilians and individuals in the military. Individuals in the military mostly work and live in special environments such as mountains, islands, and ships that are characterized by high humidity, high salinity, high temperatures, and low air pressure. They also suffer from stress due to fatigue, tension, lack of sleep and military-related issues; thus, they are more susceptible to *S. Enteritidis* infection than the general population. The characteristic manifestations of *Salmonella* infection include fever, chills, watery stool, nausea, vomiting, abdominal pain and so on[4], these symptoms can significantly reduce the combat effectiveness of soldiers.

In recent years, because of the abuse of antibiotics in both human beings and poultry, high proportions of *S. Enteritidis* isolates have been reported to be resistant to multiple antibiotics, resulting in increasingly challenging of *S. Enteritidis* treatment. The gold standards for *S. Enteritidis* infection diagnosis and antimicrobial susceptibility analysis rely on fecal/blood cultures, but these processes are time-consuming. Therefore, identifying other parameters correlated with the diagnosis, severity and outcome of *S. Enteritidis* and clarifying the antibiotic resistance status of *S. Enteritidis* are imperative. In recent years, because of the abuse of antibiotics, high proportions of *S. Enteritidis* isolates have been reported to be resistant to multiple antibiotics[5], resulting in increasingly challenging *S. Enteritidis* treatment. The gold standards for *S. Enteritidis* diagnosis and antimicrobial susceptibility analysis rely on fecal/blood cultures, but these processes are time-consuming. Therefore, identifying other parameters correlated with the diagnosis, severity and outcome of *S. Enteritidis* infection and clarifying the antibiotic resistance status of *S. Enteritidis* are imperative.

Iron is critical for both humans and invading pathogens[6], it is correlated with the oxygen transport, the tricarboxylic acid (TCA) cycle, lipid and sterol metabolism, mitochondrial respiration, chromatin remodeling, DNA replication and repair and so on[7]. The invading pathogens and the hosts compete for the use of iron[6]. In acute infectious diseases, including bacteria (such as *S. Typhi*[8]), fungi[7] (such as *C. glabrata*, *C. albicans*, *A. fumigatus* and *C. neoformans*) and virus (such as coronavirus disease 2019, COVID-19[9]), inflammatory factors, such as IL-6, could increase the expression of hepcidin to promote the degradation of the iron exporter ferroportin in duodenal enterocytes and macrophages[10–14], which could decrease the serum iron concentration by restricting iron uptake and macrophage iron release[15]. The dysregulation of iron release by macrophages could contribute to the survival of intracellular, macrophage-tropic bacteria[13], but could be detrimental to the extracellular pathogens[8, 16, 17]. However, its correlation with *S. Enteritidis* infection remains unknown.

Coagulation system disturbance is associated with various infectious diseases such as *S. Typhi*[18], Meningococcal[19], Staphylococcus[20], COVID-19[21] and HBV[22, 23]. *S. Typhi* can activate the coagulation system, and the consumption of coagulation factors could extend the prothrombin time (PT) and the activated partial thromboplastin time (APTT)[18]. D-dimer and PT are significant indicators of severe COVID-19 and a poor prognosis[24, 25]. However, the correlation between the function of the coagulation system and *S. Enteritidis* infection remains unclear.

In this study, we isolated 6 strains of *Salmonella* from stool cultures, and serovar identification indicated *S. Enteritidis*. All 6 strains were sensitive to trimethoprim-sulfamethoxazole and levofloxacin, and resistant to ampicillin. Levofloxacin combined with rifaximin and probiotics could be effective in *S. Enteritidis* therapies. We also found that the white blood cell (WBC) count, neutrophil proportion, C-reactive protein (CRP), PT and D-dimer were elevated, while serum iron and prealbumin were decreased in *S. Enteritidis*-infected patients. Prolonged PT, elevated D-dimer and decreased serum iron were positively correlated with serious symptoms (high body temperature and high diarrhea frequency) and increased concentrations of serum inflammatory indexes (CRP, WBC count and neutrophil proportion). Thus, we concluded that prolonged PT, upregulated D-dimer and decreased serum iron were associated with the diagnosis, severity and outcome of *S. Enteritidis* infection, and decreased serum prealbumin was associated with the diagnosis of *S. Enteritidis* infection.

Methods

Blood sample collection and testing

Blood samples were collected from the patients every day by a skilled nurse according to the doctor's order; the first blood samples were collected before antibiotic treatment was initiated. Routine blood tests (Blood cell automated analyzer, XN-1500, Japan SYSMEX, Japan), blood biochemical analyses (Blood biochemical automated analyzer, Becman, American), serum CRP analyses, coagulation function analyses (Blood coagulation analyzer, ACL TPO 700, IL company, American), and serum folate analyses were conducted by our clinical laboratory. Serum hepcidin was detected by ELISA (ELISA Kit for Hepsidin, CEB979Hu, Cloud-Clone Corp, American) according to the manufacturer's instructions.

Faecal Sample Collecting And Test

Fecal samples were collected from the patients every day, and routine stool tests, FOBTs, stool *Clostridium difficile* (*C. difficile*) toxin and antigen detection, stool fungal detection by smear, stool bacterial cultures, and *Salmonella* serovar identification were conducted by our clinical laboratory.

The stool samples were inoculated on SS agar medium. After incubation at 35°C for 24 hours, the single bacterial colony was identified with MALDI TOF MS (Microflex series of Bruker, XJ-058, German). The Serum agglutination test was performed to identify the serotype with the diagnostic sera for *Salmonella* (NingBo, TianRun Bio-Parmaceutical, co.ltd, China), which could identify 60 kinds of *Salmonella* serovar. Antimicrobial susceptibility testing was performed using the MIC method with the VITEK 2 COMPACT analyzer (BioMerieux, France).

Bristol Stool Form Scale

The Bristol Stool Form Scale was used to analyze stool consistency. According to the form of the stool, human feces could be classified into 7 categories: type 1 (separate hard lumps); type 2 (sausage-like but lumpy); type 3 (sausage-like with cracks on the surface); type 4 (sausage- or snake-like, smooth and soft); type 5 (soft blobs with clear edges); type 6 (fluffy pieces with ragged edges, mushy stool); type 7 (watery, no solid pieces, entirely liquid).

Statistical Analyses

All statistical analyses were performed with SPSS 20.0. Pearson correlation coefficients were calculated to analyze correlations between different characteristics. The comparison of quantitative data between different groups was conducted with paired or unpaired Student's t-tests. A value of $p < 0.05$ was considered statistically significant.

Results

The clinical characteristics of the patients infected with *S. Enteritidis*

Recently, 6 patients from one company visited our department, they all complained of fever (body temperatures ranged from 38.5°C to 39.2°C), watery stool (Bristol type 7, defecation frequency ranged from 3 times/day to 10 times/day), nausea, abdominal pain, chills and fatigue; some reported vomiting (1 out of 6, 16.7%), lower back pain (2/6, 33.3%), headache (2/6, 33.3%) and knee pain (2/6, 33.3%) (Table 1). They were all males aged between 18 and 23 years.

Table 1
The clinical/pathological variables of the patients when they were admitted.

Symptoms										
Patient	Temperature (°C)	Defection frequency (times/day)	Stool Bristol type	Nausea	Abdominal pain	Chills	Fatigue	Vomit	Lower back pain	Headache
Case 1	39.1	5	7	+	+	+	+	-	+	+
Case 2	38.8	3	7	+	+	+	+	+	-	-
Case 3	39	8	7	+	+	+	+	-	-	-
Case 4	39	4	7	+	+	+	+	-	-	-
Case 5	38	4	7	+	+	+	+	-	+	+
Case 6	38.8	10	7	+	+	+	+	-	-	-
Blood routine test					Coagulation function				Inflammatory	
Patient	WBC	Neutrophil	Lymphocyte	Hb	Platelet	PT	APTT	Fibrinogen	D-dimer	CRP
	(3.5–9.5*10 ⁹ /L)	(40–75%)	(20–50%)	(130–175 g/L)	(125–350*10 ⁹ /L)	(9.4–12.5 s)	(25.1–36.5 s)	(2–4 g/L)	(0–299 mg/ml)	(0–5 mg/L)
Case 1	13.75	91.9	4.1	144	194	14.1	35.9	2.17	512	41.9
Case 2	8.37	86.7	9.2	153	172	16.5	27.7	2.21	428	7.2
Case 3	15.72	91.3	4.8	154	178	17.3	17.8	2.59	479	73.8
Case 4	11.54	88.5	6.3	153	161	13.2	31.8	2.06	422	14.3
Case 5	7.1	83.2	11.9	153	187	12.7	36.3	2.18	384	23.2
Case 6	10.03	78.4	14.5	163	179	13.1	34.6	2.21	296	8.3
Liver function					Renal function					
Patient	ALT	AST	Albumin	Prealbumin	ALP	GGT	Total bile acid	Cr	BUN	Serum iron
	(9–50 U/L)	(8–40 U/L)	(40–55 g/L)	(250–400 g/L)	(40–150 U/L)	(11–50 U/L)	(<10 μmol/L)	(62–115 μmol/L)	(2.9–8.2 mmol/L)	(10.6–36.7 μmol/L)
Case 1	15.6	21.8	48.6	218	63.3	15.2	1	89.6	4.5	1.3
Case 2	14.2	16.9	47.1	264	73.5	10.6	3.6	90.9	4.7	3.6
Case 3	8.5	16.6	49.1	219	95.5	20.9	1.3	101.1	5.9	3
Case 4	13.3	19.9	44.3	215	78	10	1	91.9	3.3	2.8
Case 5	9.9	16.1	46.5	241	102.3	11.9	2.5	78.2	3.7	4.8
Case 6	16.5	16.1	45.2	237	69.4	20	1.2	87.8	6.2	3.5

To make a definitive diagnosis, the blood tests were accomplished. The results of the inflammatory parameters showed that the WBC counts in 4 patients (4/6, 66.7%) and the neutrophil proportions and CRP levels in all patients (6/6, 100%) were higher than normal (Table 1). Acute infection may result in multiorgan functional disturbance; thus, we investigated the parameters of liver function, renal function, myocardial enzymes, arterial blood gas analysis results, coagulation function and so on. The results showed that the patients' serum prealbumin levels (5/6, 83.3%) were lower than normal, and patients' PTs (6/6, 100%) and D-dimer (5/6, 83.3%) levels were significantly elevated. Moreover, the patients' alanine aminotransferase (ALT), aspartate aminotransferase (AST) and albumin levels, APTTs, fibrinogen levels, renal function parameters (blood urea nitrogen [BUN], creatinine [Cr]), myocardial enzymes (Tnl) and arterial blood gas analyses results (pH, PaO₂, PaCO₂) were normal (Table 1). Interestingly, we also found that the serum iron concentrations in the 6 patients were all lower than normal.

Ultrasonography and intestinal computed tomography (CT) examinations revealed intestinal mucosal edema in 3 patients (3/6, 50%), and ascites in 0 patient.

Routine stool examinations revealed red blood cells (RBCs) and WBCs in the stool of 2 patients (2/6, 33.3%); however, coccus to bacillus (C/B) ratio in all 6 patients could not be analyzed, because the bacterial loads were not sufficient (Table 2). In accordance with the routine fecal test results, fecal occult blood tests (FOBTs) were positive in 2 patients (2/6, 33.3%) (Table 2). The detection of *C. difficile* toxins and antigens in the stool were negative (Table 2). Stool culture on *Salmonella Shigella* (SS) agar plates were all positive, the single bacterial colonies were identified with mass spectrometry (MS) and the results indicated *Salmonella*. Serovar testing revealed all isolated strains were *S. enterica* serovar Enteritidis (*S. Enteritidis*) as detected by the Serum agglutination test (Table 2). Taken all, prolonged PT, elevated D-dimer, decreased serum iron and decreased serum prealbumin were associated with the diagnosis of *S. Enteritidis* infection.

Table 2
The fecal test results of the 6 patients when they were admitted.

	stool routine test (cells/per high-power field)		FOBT	C. difficile		Stool culture	Serovar
	WBC	RBC		antigen	toxin A&B		
Case 1	8–12	10–15	+	-	-	+	+
Case 2	0–1	0	-	-	-	+	+
Case 3	3–5	20–30	+	-	-	+	+
Case 4	0	0	-	-	-	+	+
Case 5	0	0	-	-	-	+	+
Case 6	0	0	-	-	-	+	+

The Kinetics Of The Patients'; Symptoms And Clinicopathological Variables

Antimicrobial susceptibility testing was performed using the MIC method with the VITEK 2 COMPACT analyzer. All the isolated *Salmonella* strains were sensitive to trimethoprim-sulfamethoxazole and levofloxacin and resistant to ampicillin.

Before fecal cultures and antibacterial susceptibility results were obtained, levofloxacin and *Bacillus licheniformis* capsules were administered according to experience, accompanied by body temperature control, fluid infusion and correction of electrolyte imbalance. According to the antibacterial susceptibility results, we added rifaximin and live combined *B. subtilis* and *Enterococcus faecium* enteric-coated capsules to destroy the *Salmonella* and to modulate the imbalance in the gut microbiota. During the disease course, the body temperatures of 5 patients returned normal on the second day of hospitalization, and the body temperatures of the other 2 patients were controlled on the third day (Fig. 1a). The defecation frequency and stool characteristics (Bristol type) kept decreasing to normal after admission (Fig. 1b). CRP levels increased on the second day and then returned to normal (Fig. 1c); WBC counts (Fig. 1d) and neutrophil percentages (Fig. 1e) continued to decrease after admission. Serum iron levels decreased on the second day and then increased (Fig. 1f). Serum prealbumin levels decreased in the first 3 days and then increased (Fig. 1g). PT increased on the second day and then decreased (Fig. 1h). D-dimer continued to decrease to normal after admission (Fig. 1i). The bowel wall was not detected to be thickening on the 8th day. After hospitalization, 3 additional fecal cultures (on the 3rd, 4th and 5th day respectively) were performed, and the results were all negative. Levofloxacin was withdrawn when a patient's body temperature remained normal for 3 days and the defecation frequency was less than 3 times per day. Patients were discharged on the 10th day.

Positive associations between decreased serum iron and the severity/outcome of *S. Enteritidis* infection

In this study, we found that serum iron levels in all patients was lower than normal, and patients with a higher body temperature, higher WBC count and higher neutrophil concentration had lower serum iron level, and with the remission of symptoms and inflammatory parameters, serum iron increased (Table 1 and Fig. 1). We then analyzed correlations between serum iron level and inflammatory parameters and found that the serum iron concentration was negatively associated with body temperature (Pearson correlation coefficient $r = -0.458$, $p = 0.002$, Fig. 2a), defecation frequency (Pearson $r = -0.524$, $p < 0.001$, Fig. 2b), CRP level (Pearson $r = -0.604$, $p < 0.001$, Fig. 2c), WBC count (Pearson $r = -0.429$, $p = 0.005$, Fig. 2d), neutrophil proportion (Pearson $r = -0.658$, $p < 0.001$, Fig. 2e) and PT (Pearson $r = -0.531$, $p < 0.001$, Fig. 2f).

Hepcidin is a cytokine synthesized by liver and plays a critical role in iron metabolism. We then investigated whether hepcidin is involved in iron down-regulation during *S. Enteritidis* infection. Our results showed that the patients' serum hepcidin levels (6/6, 100%) were higher than normal when they were admitted, they increased on the second day, and then decreased with symptom remission (Fig. 3a). Serum hepcidin was negatively correlated with serum iron (Pearson $r = -0.889$, $p < 0.001$, Fig. 3b). Above all, our results indicated that the decreased serum iron was associated with the severity and outcome of *S. Enteritidis* infection, and that the elevated serum hepcidin was associated with the decreased serum iron during *S. Enteritidis* infection.

Positive associations between the prolonged PT/elevated D-dimer and the severity /outcome of the *S. Enteritidis* infection

We then investigate the correlations between the coagulation system disturbance and the severity and outcome of *S. Enteritidis* infection. The patient with severe symptoms had a higher D-dimer and longer PT than the other patients, and with the remission of symptoms and inflammatory parameters, PTs and D-dimer levels were decreased (Table 1 and Fig. 1). PT was positively associated with body temperature (Pearson $r = 0.447$, $p = 0.003$, Fig. 4a), defecation frequency (Pearson $r = 0.566$, $p < 0.001$, Fig. 4b), CRP level (Pearson $r = 0.69$, $p < 0.001$, Fig. 4c), WBC count (Pearson $r = 0.544$, $p < 0.001$, Fig. 4d) and neutrophil proportion (Pearson $r = 0.612$, $p < 0.001$, Fig. 4e); serum D-Dimer concentration was positively associated with body temperature (Pearson $r = 0.823$, $p < 0.001$, Fig. 4f), defecation frequency (Pearson $r = 0.693$, $p < 0.001$, Fig. 4g), CRP level (Pearson $r = 0.512$, $p = 0.001$, Fig. 4h), WBC count (Pearson $r = 0.752$, $p < 0.001$, Fig. 4i) and neutrophil proportion (Pearson $r = 0.884$, $p < 0.001$, Fig. 4j). Taken together, the prolonged PT and elevated D-Dimer were correlated with the severity and outcome of the *S. Enteritidis* infection.

The Correlation Between Serum Prealbumin And Acute Inflammation Status

Serum prealbumin and albumin are indicators of liver synthesis function, and prealbumin may decrease earlier than albumin when liver function is damaged. As shown above, the decreased prealbumin was associated the diagnosis of *S. Enteritidis* infection. We then investigated the association between serum prealbumin and the severity and outcome of *S. Enteritidis* infection, and found that it was negatively correlated with CRP level (Pearson $r = -0.678$, $p < 0.001$, Fig. 5a) and PT (Pearson $r = -0.353$, $p = 0.022$, Fig. 5b); there was no significant correlations between serum prealbumin and body temperature (Pearson $r =$

0.208, $p = 0.187$, Fig. 5c), defecation frequency (Pearson $r = -0.138$, $p = 0.383$, Fig. 5d), WBC count (Pearson $r = -0.073$, $p = 0.646$, Fig. 5e) and neutrophil proportion (Pearson $r = -0.093$, $p = 0.559$, Fig. 5f).

As the serum prealbumin continued to decrease in the first 3 days, and begun to increase at the 4th day, while other parameters of inflammation begun to restore at the 2th or 3th day. We then analyzed the correlation between serum prealbumin and inflammatory parameters of the first day, and found that although the correlation coefficients were larger than 0.2 or smaller than -0.2 , the p values were all larger than 0.05 (Fig. 5g-5 l), this could be reasoned by the small case number. Taken together, our results indicated that a decreased prealbumin was correlated with the diagnosis of the *S. Enteritidis* infection, but more solid proofs were needed to verify its association with the severity and outcome of *S. Enteritidis* infection.

The kinetics of serum homocysteine during *S. Enteritidis* infection

When the patients were admitted, all patients had symptoms of diarrhea, thus, a low-fat and low-protein diet was provided. A normal diet was reintroduced when a patient' defecation frequency was less than 3 times per day and a patient' fecal Bristol type returned to type 5. However, during the disease course in these 6 patients, we found that the patients' homocysteine levels continued to increase beginning on the 6th day of hospitalization, and 2 patients' homocysteine levels were higher than normal on the 7th day (Fig. 6a). The patients were aged from 18 to 23 years, their blood pressures were normal, and none had a family history of high blood pressure; therefore, other mechanisms responsible for the increased serum homocysteine may exist. Serum folate plays a critical role in homocysteine metabolism. Accompanied by methyltetrahydrofolic acid, homocysteine can be transformed into methionine and tetrahydrofolic acid by methylenetetrahydrofolate (MTHFR). We found that the patients' serum folate levels were either lower than normal or at the lower normal limit (7 nmol/L \sim 45.6 nmol/L) on the 8th day. We provided folate tablets (5 mg, QD) to the patients with high serum homocysteine and asked the other patients to improve their nutrition and consume more green vegetables for folic acid supplementation. Then, serum folate increased (Fig. 6b), while serum homocysteine decreased (Fig. 6a). Thus, we suggest that a normal diet should be reintroduced early.

Discussion

Salmonella infection is a foodborne disease, and meat products are the main sources of human salmonellosis. In this study, our epidemiological investigation revealed that all 6 patients had a history of consuming the same improperly stored, cold meals that were purchased online. Samples were collected and sent to the Center for Disease Control (CDC); the specific source needs further investigation.

To date, more than 2600 *Salmonella* serotypes have been reported, among which *S. Enteritidis* and *S. Typhi* are the most common[5]. *S. Enteritidis* infection is generally self-limiting, but patients can present with severe manifestations, and these patients should receive antibiotics therapy. In this study, the 6 patients suffered from fever, chills, watery stool, nausea and fatigue, which needed appropriate treatment. However, because of antibiotic abuse, high proportions of *Salmonella* isolates are resistant to tetracycline (53.9%), ciprofloxacin (47.2%), ampicillin (44.4%), nalidixic acid (42.7%), and trimethoprim-sulfamethoxazole (38.8%)[26]. In this study, all 6 isolates were sensitive to trimethoprim-sulfamethoxazole and levofloxacin and resistant to ampicillin. A combination of levofloxacin, rifaximin, and live combined *B. subtilis* and *E. faecium* enteric-coated capsules were administered, and the patients' symptoms began to improve on the second day, indicating that this treatment strategy was effective. Besides medicine treatment, we also recommend strengthening nutrition and reintroducing the normal diet early, however, the specific time to reintroduce the normal diet needs further investigation.

As stool culture and blood culture examinations are time-consuming, identifying other parameters correlated with *S. Enteritidis* infection is meaningful. In this study, we found that in addition to the elevated inflammatory indicators such as body temperature, CRP, WBC count and neutrophil cell proportion, the patients' serum iron and prealbumin levels were significantly decreased, while PT and D-dimer levels were elevated.

Iron metabolism disturbance has been widely reported in various infectious diseases. *S. Typhi* competes with humans for iron utilization by increasing the expression of hepcidin, which can restrict iron uptake and macrophage iron release, and the high serum hepcidin leads to hypoferrremia[8]; COVID-19 infection resulted in hyperferritinemia due to cytokine storms, and serum iron transferrin saturation in intensive care unit (ICU) patients infected with COVID-19 was extremely reduced for the first 2 days and then increased from days 3 to 6[9, 27, 28]. In this study, we found that serum iron was significantly decreased in *S. Enteritidis* patients, and it seemed to be lower in patients with more serious symptoms; with the recovery of patients' symptoms, serum iron increased. Serum iron was negatively associated with body temperature, defecation frequency, CRP, WBC count, neutrophil proportion and PT. We also found that in the patients, serum hepcidin was elevated and that its concentration was negatively associated with serum iron. Thus, we hypothesized that *S. Enteritidis* infection downregulated serum iron by increasing serum hepcidin, and the decreased serum iron was correlated with the diagnosis, severity and outcome of *S. Enteritidis* infection.

Coagulation system activation is a characteristic parameter of acute infectious disease. Acute inflammation-mediated endothelial cell damage could result in the activation of the coagulation system, and the consumption of coagulation factors leads to disseminated intravascular coagulation (DIC)[29]. *S. Typhi* infection has been reported to activate the coagulation system, increase serum fibrinogen and D-dimer and prolong PT and APTT[18]. COVID-19 infection may lead to disturbances in the procoagulation, anticoagulation and fibrinolytic systems[30–34]; a high concentration of D-dimer and prolonged PT are associated with severe COVID-19 and a poor prognosis[24]. In this study, we found that RBCs were detected in the stool of 2 patients, PT was prolonged and D-dimer was elevated in all 6 patients, while APTTs and fibrinogen levels were normal. There was no obvious change in the APTT during the disease course (Additional file 1: Fig. 1a). Fibrinogen concentrations increased for the first 4 days and then decreased, but during the disease course, serum fibrinogen in only 1 patient was higher than normal (4 g/L) on days 4, 5 and 6 (Additional file 1: Fig. 1b). Prolonged PT and elevated D-dimer were positively correlated with more serious symptoms (high body temperature and high diarrhea frequency) and high concentrations of serum inflammatory indexes (CRP, WBC count and neutrophil proportion), thus, we reported that a prolonged PT and an increased D-dimer concentration were associated with the diagnosis, severity and outcome of *S. Enteritidis* infection.

Acute infection may cause a strike and suppress the function of multiple organs. Gamma-glutamyltransferase (GGT) and prealbumin were reported to have critical values in the diagnosis of COVID-19, and prealbumin and albumin were downregulated as the disease severity increased[25]. The decreased prealbumin is a marker of disease activity in cholerae infection[35]. In this study, we found that serum prealbumin in 5 patients was lower than normal when they were admitted, and it continued to decrease for the first 3 days. It finally began to increase on the 4th day of admission. However, analyzing the associations between serum prealbumin concentration and the index of the severity of the disease, there was no significant correlation, which may be explained by bowel wall edema-mediated intestinal absorption disorder and liver functional reserve. In all, we reported that decreased prealbumin was correlated with the diagnosis of *S. Enteritidis* infection, but more solid proofs are needed to verify its associations with the severity and the outcome of the *S. Enteritidis* infection.

Conclusion

In this study, we isolated 6 strains of *Salmonella* from stool cultures, and serovar testing indicated *S. Enteritidis*. All 6 strains were sensitive to trimethoprim-sulfamethoxazole and levofloxacin and resistant to ampicillin. Levofloxacin combined with rifaximin and probiotics could be an effective *S. Enteritidis* therapy. The prolonged PT, elevated D-dimer and decreased serum iron were associated with the diagnosis, severity and outcome of the *S. Enteritidis* infection, the decreased serum prealbumin was associated with the diagnosis of *S. Enteritidis* infection. In patients with symptoms of acute fever, watery stool, and abdominal pain, PT, D-dimer, serum iron and serum prealbumin should be monitored to attain an early diagnosis and initiate appropriate treatment for *S. Enteritidis* infection.

Abbreviations

S. Enteritidis: *Salmonella enterica* serovar Enteritidis; FOBTs: fecal occult blood tests; SS: *Salmonella Shigella*; MS: mass spectrometry; MIC: minimal inhibitory concentration; WBC: white blood cell; RBC: red blood cell; CRP: C-reactive protein; PT: prothrombin time; TCA: tricarboxylic acid; APTT: activated partial thromboplastin time; COVID-19: coronavirus disease 2019; *difficile*: *Clostridium difficile*; ALT: alanine aminotransferase; AST: aspartate aminotransferase; GGT: Gamma-glutamyltransferase; BUN: blood urea nitrogen; CR: creatinine; Tnl: myocardial enzymes; CT: computed tomography; C/B: coccus to bacillus; CDC: Center for Disease Control.

Declarations

Ethics approval and consent to participate

The researchers collected the data after the project was certified from the Research Ethics Committee, the Sixth Medical Center of PLA General Hospital.

Consent for publication

Not applicable.

Availability of data and materials

All data generated or analyzed during this study are included in this published article [and its supplementary information files].

Competing interests

The authors declare that they have no competing interests.

Funding

This study was sponsored by the foundation of the logistics department of Chinese Navy (BHJ14L010).

Authors' contributions

LHC designed the project, provided the funding, corrected the manuscript and approved the final version; XWL analyzed the data, accomplished the Elisa study, wrote the original draft; SXW, LY and XNL helped with the data collection and literature research; XHW, ZHY and ZY helped with the statistical analyses; YJL accomplished the main laboratory test; JR helped with the sample collection.

Acknowledgements

We would like to thank the military nurse team at the the Sixth Medical Center of PLA General Hospital (Bing Li, Tong Wu, Zhaoxia Jiang, Jinli Tang, Qi Zhang, Xiaoxiao Chen, Liping Yang, Shuye Wang, Ke Zhan, Shuaimeng Cai, Liyun Zang, Yuanyuan Zhao, Wenxiao Ding), they helped collect the blood and fecal samples. We are also grateful to the laboratory team at the Sixth Medical Center of PLA General Hospital who help with the clinical testing.

References

1. Phoba MF, Barbe B, Ley B, Van Puyvelde S, Post A, Mattheus W, et al: High genetic similarity between non-typhoidal Salmonella isolated from paired blood and stool samples of children in the Democratic Republic of the Congo. *PLoS Negl Trop Dis* 2020, 14:e0008377.

2. Majowicz SE, Musto J, Scallan E, Angulo FJ, Kirk M, O'Brien SJ, et al: The global burden of nontyphoidal Salmonella gastroenteritis. *Clin Infect Dis* 2010, 50:882-889.
3. Yang X, Wu Q, Zhang J, Huang J, Chen L, Wu S, et al: Prevalence, Bacterial Load, and Antimicrobial Resistance of Salmonella Serovars Isolated From Retail Meat and Meat Products in China. *Front Microbiol* 2019, 10:2121.
4. Lee KS, Kim D, Lee H, Lee K, Yong D: Isolation of Non-Hydrogen Sulfide-Producing Salmonella enterica Serovar Infantis from a Clinical Sample: the First Case in Korea. *Ann Lab Med* 2020, 40:334-336.
5. Xu Z, Wang M, Zhou C, Gu G, Liang J, Hou X, et al: Prevalence and antimicrobial resistance of retail-meat-borne Salmonella in southern China during the years 2009-2016: The diversity of contamination and the resistance evolution of multidrug-resistant isolates. *Int J Food Microbiol* 2020, 333:108790.
6. Sousa Geros A, Simmons A, Drakesmith H, Aulicino A, Frost JN: The battle for iron in enteric infections. *Immunology* 2020.
7. Martinez-Pastor MT, Puig S: Adaptation to iron deficiency in human pathogenic fungi. *Biochim Biophys Acta Mol Cell Res* 2020, 1867:118797.
8. Darton TC, Blohmke CJ, Giannoulitou E, Waddington CS, Jones C, Sturges P, et al: Rapidly Escalating Hepsidin and Associated Serum Iron Starvation Are Features of the Acute Response to Typhoid Infection in Humans. *PLoS Negl Trop Dis* 2015, 9:e0004029.
9. Edeas M, Saleh J, Peyssonnaud C: Iron: Innocent bystander or vicious culprit in COVID-19 pathogenesis? *Int J Infect Dis* 2020, 97:303-305.
10. Nemeth E, Rivera S, Gabayan V, Keller C, Taudorf S, Pedersen BK, et al: IL-6 mediates hypoferrremia of inflammation by inducing the synthesis of the iron regulatory hormone hepcidin. *J Clin Invest* 2004, 113:1271-1276.
11. Shu W, Pang Z, Xu C, Lin J, Li G, Wu W, et al: Anti-TNF-alpha Monoclonal Antibody Therapy Improves Anemia through Downregulating Hepatocyte Hepsidin Expression in Inflammatory Bowel Disease. *Mediators Inflamm* 2019, 2019:4038619.
12. Ganz T: Iron in innate immunity: starve the invaders. *Curr Opin Immunol* 2009, 21:63-67.
13. Nairz M, Weiss G: Iron in infection and immunity. *Mol Aspects Med* 2020:100864.
14. Hood MI, Skaar EP: Nutritional immunity: transition metals at the pathogen-host interface. *Nat Rev Microbiol* 2012, 10:525-537.
15. Nairz M, Dichtl S, Schroll A, Haschka D, Tymoszuk P, Theurl I, et al: Iron and innate antimicrobial immunity-Depriving the pathogen, defending the host. *J Trace Elem Med Biol* 2018, 48:118-133.
16. Drakesmith H, Prentice AM: Hepsidin and the iron-infection axis. *Science* 2012, 338:768-772.
17. Arezes J, Jung G, Gabayan V, Valore E, Ruchala P, Gulig PA, et al: Hepsidin-induced hypoferrremia is a critical host defense mechanism against the siderophilic bacterium *Vibrio vulnificus*. *Cell Host Microbe* 2015, 17:47-57.
18. de Jong HK, Parry CM, van der Vaart TW, Kager LM, van den Ende SJ, Maude RR, et al: Activation of coagulation and endothelium with concurrent impairment of anticoagulant mechanisms in patients with typhoid fever. *J Infect* 2018, 77:60-67.
19. Aladag Ciftdemir N, Duran R, Vatansever Ozbek U, Hancerli Torun S, Acunas B, Unal Sahin N: [Meningococcemia: Different Serotypes in the Same Region]. *Mikrobiyol Bul* 2020, 54:163-170.
20. Krogh AKH, Brunse A, Thymann T, Bochsén L, Kristensen AT: Staphylococcus epidermidis sepsis induces hypercoagulability in preterm pigs. *Res Vet Sci* 2019, 127:122-129.
21. Iavarone M, D'Ambrosio R, Soria A, Triolo M, Pugliese N, Del Poggio P, et al: High rates of 30-day mortality in patients with cirrhosis and COVID-19. *J Hepatol* 2020.
22. Chang ML, Cheng JS, Chien RN, Liaw YF: Hepatitis Flares Are Associated With Better Outcomes Than No Flare in Patients With Decompensated Cirrhosis and Chronic Hepatitis B Virus Infection. *Clin Gastroenterol Hepatol* 2020, 18:2064-2072 e2062.
23. Shanshan W, Xinfang D, Shuihong Y, Kecong L, Jinjin Q, Zhi C, et al: Pathological changes of liver one year later in CHB patients with negative HBV DNA. *Infect Agent Cancer* 2019, 14:48.
24. Long H, Nie L, Xiang X, Li H, Zhang X, Fu X, et al: D-Dimer and Prothrombin Time Are the Significant Indicators of Severe COVID-19 and Poor Prognosis. *Biomed Res Int* 2020, 2020:6159720.
25. Wei XY, Jing D, Jia B, Li Q, Zhou XQ, Gong MF, et al: Characteristics of in peripheral blood of 70 hospitalized patients and 8 diarrhea patients with COVID-19. *Int J Med Sci* 2020, 17:1142-1146.
26. Peruzzy MF, Capuano F, Proroga YTR, Cristiano D, Carullo MR, Murru N: Antimicrobial Susceptibility Testing for Salmonella Serovars Isolated from Food Samples: Five-Year Monitoring (2015-2019). *Antibiotics (Basel)* 2020, 9.
27. Bolondi G, Russo E, Gamberini E, Circelli A, Meca MCC, Brogi E, et al: Iron metabolism and lymphocyte characterisation during Covid-19 infection in ICU patients: an observational cohort study. *World J Emerg Surg* 2020, 15:41.
28. Zhao K, Huang J, Dai D, Feng Y, Liu L, Nie S: Serum Iron Level as a Potential Predictor of Coronavirus Disease 2019 Severity and Mortality: A Retrospective Study. *Open Forum Infect Dis* 2020, 7:ofaa250.
29. Cohen J: The immunopathogenesis of sepsis. *Nature* 2002, 420:885-891.
30. Zhu J, Pang J, Ji P, Zhong Z, Li H, Li B, et al: Coagulation dysfunction is associated with severity of COVID-19: a meta-analysis. *J Med Virol* 2020.
31. Puzzitiello RN, Pagani NR, Moverman MA, Moon AS, Menendez ME, Ryan SP: Inflammatory and Coagulative Considerations for the Management of Orthopaedic Trauma Patients With COVID-19: A Review of the Current Evidence and Our Surgical Experience. *J Orthop Trauma* 2020, 34:389-394.
32. Lin J, Yan H, Chen H, He C, Lin C, He H, et al: COVID-19 and Coagulation Dysfunction in Adults: A Systematic Review and Meta-analysis. *J Med Virol* 2020.
33. Al-Samkari H, Karp Leaf RS, Dzik WH, Carlson JCT, Fogerty AE, Waheed A, et al: COVID-19 and coagulation: bleeding and thrombotic manifestations of SARS-CoV-2 infection. *Blood* 2020, 136:489-500.

34. Aloisio E, Serafini L, Chibireva M, Dolci A, Panteghini M: Hypoalbuminemia and elevated D-dimer in COVID-19 patients: a call for result harmonization. Clin Chem Lab Med 2020.

35. Khan WA, Salam MA, Bennis ML: C reactive protein and prealbumin as markers of disease activity in shigellosis. Gut 1995, 37:402-405.

Figures

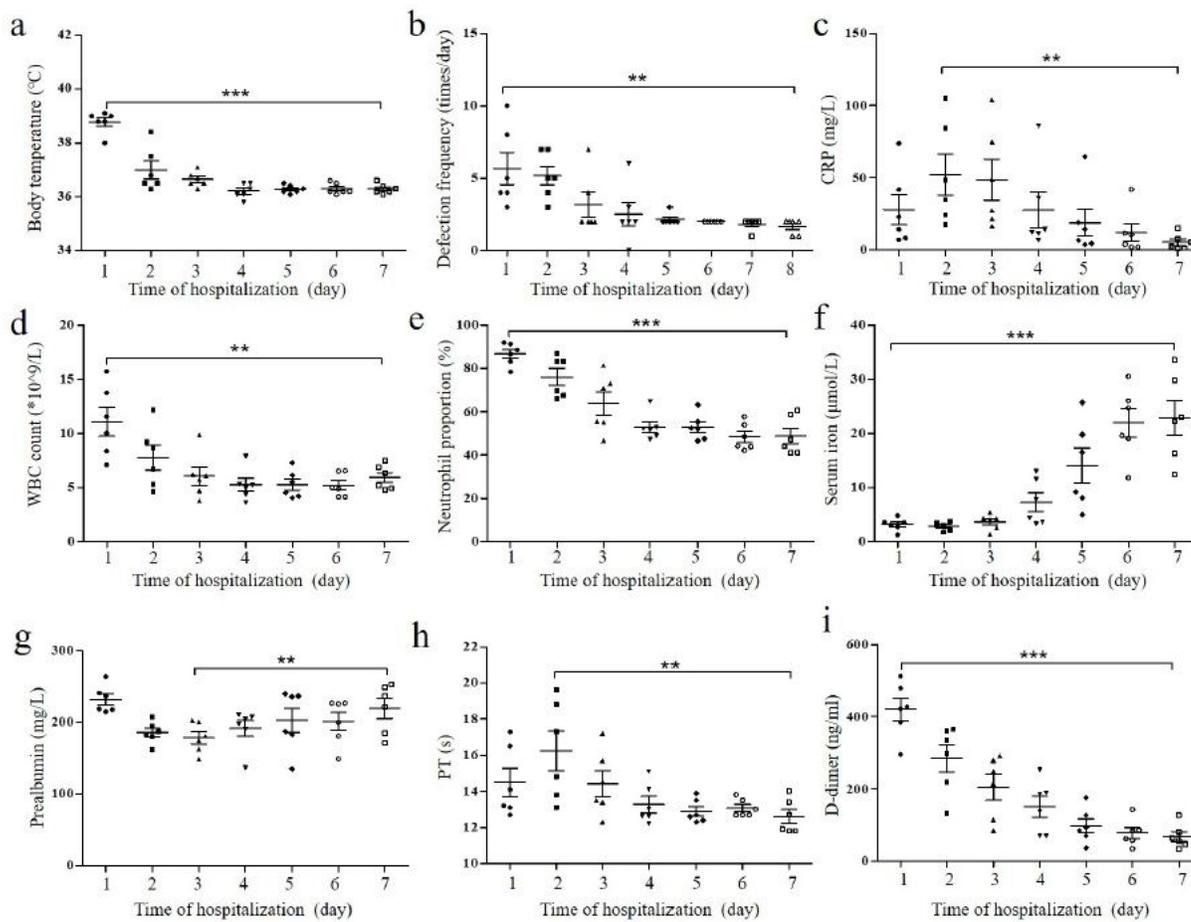


Figure 1
The kinetics of the patients' symptoms and clinicopathological variables. a Body temperatures, b Defection frequency, c CRP levels, d WBC counts, e: Neutrophil percentage, f Serum iron levels, g Prealbumin levels, h PT, i D-dimer levels were measured in the 6 patients infected with *S. Enteritidis*. **: $p < 0.01$, ***: $p < 0.001$.

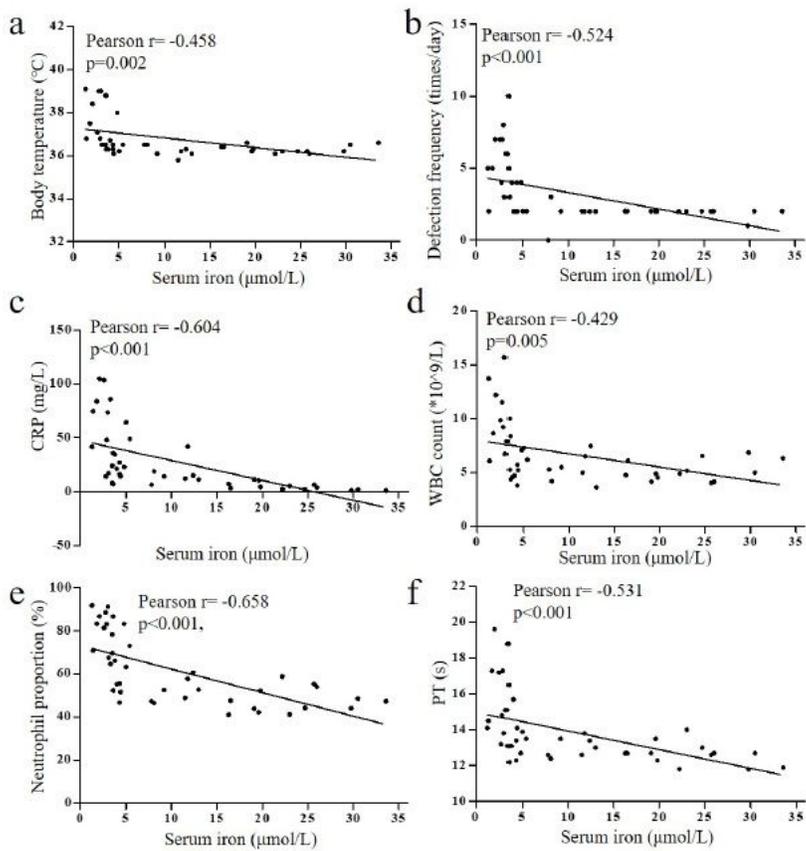


Figure 2

Correlations between serum iron levels and the severity/outcome of the *S. Enteritidis* infection. The correlation between a Serum iron levels and body temperatures, b Serum iron levels and defecation frequency, c Serum iron levels and CRP levels, d Serum iron levels and WBC counts, e Serum iron levels and neutrophil percentage, f Serum iron levels and PT were analyzed. The data of each parameter in the first 7 days were included. The Pearson correlation coefficient and the p values are stated in the corresponding Figure.

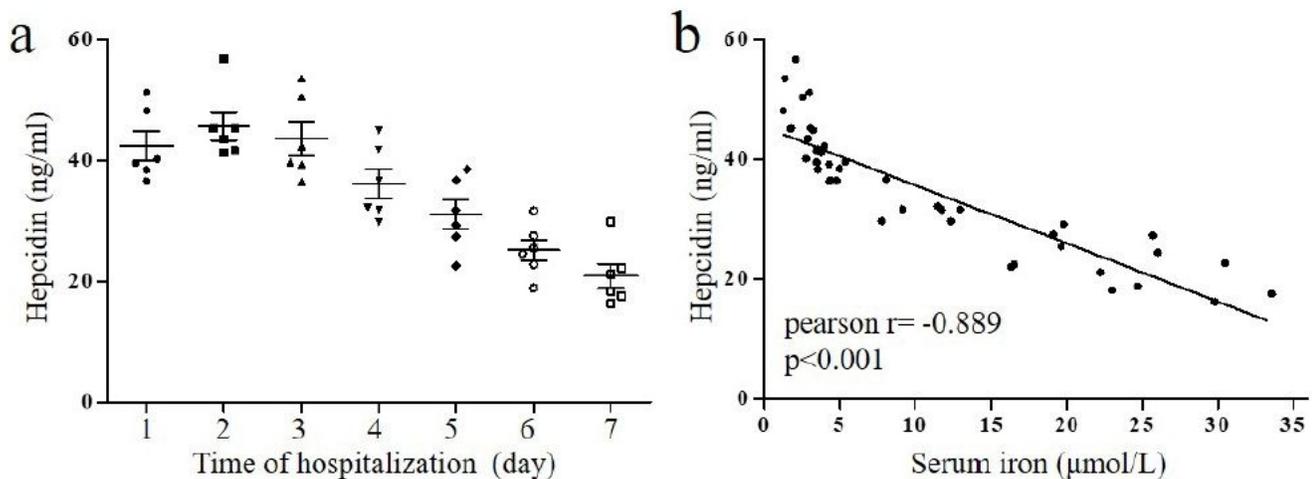


Figure 3

The elevated serum hepcidin was associated with the decreased serum iron. a The kinetics of the patients' serum hepcidin levels were measured in the 6 patients infected with *S. Enteritidis*. ***: $p < 0.001$. b The correlation between serum iron levels and hepcidin levels in the first 7 days were analyzed. The Pearson correlation coefficient and the p values are stated in the corresponding Figure.

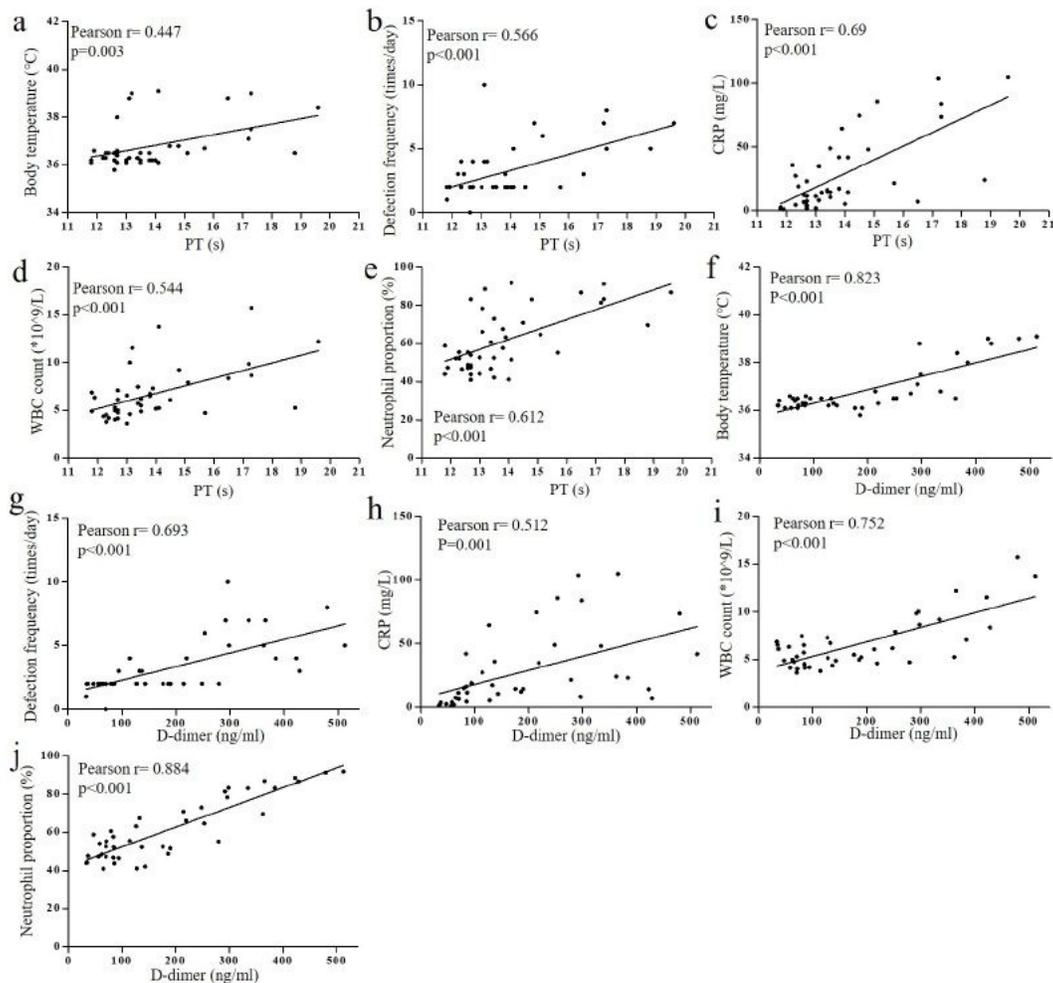


Figure 4

Correlations between PT/D-dimer and the severity/outcome of the *S. Enteritidis* infection. The correlation between a PT and body temperatures, b PT and defecation frequency, c PT and CRP levels, d PT and WBC counts, e PT and neutrophil percentage, f D-dimer levels and body temperatures, g D-dimer levels and defecation frequency, h D-dimer levels and CRP levels, i D-dimer levels and WBC counts, j D-dimer levels and neutrophil percentage were analyzed. The data of each parameter in the first 7 days were included. The Pearson correlation coefficient and the p values are stated in the corresponding figure.

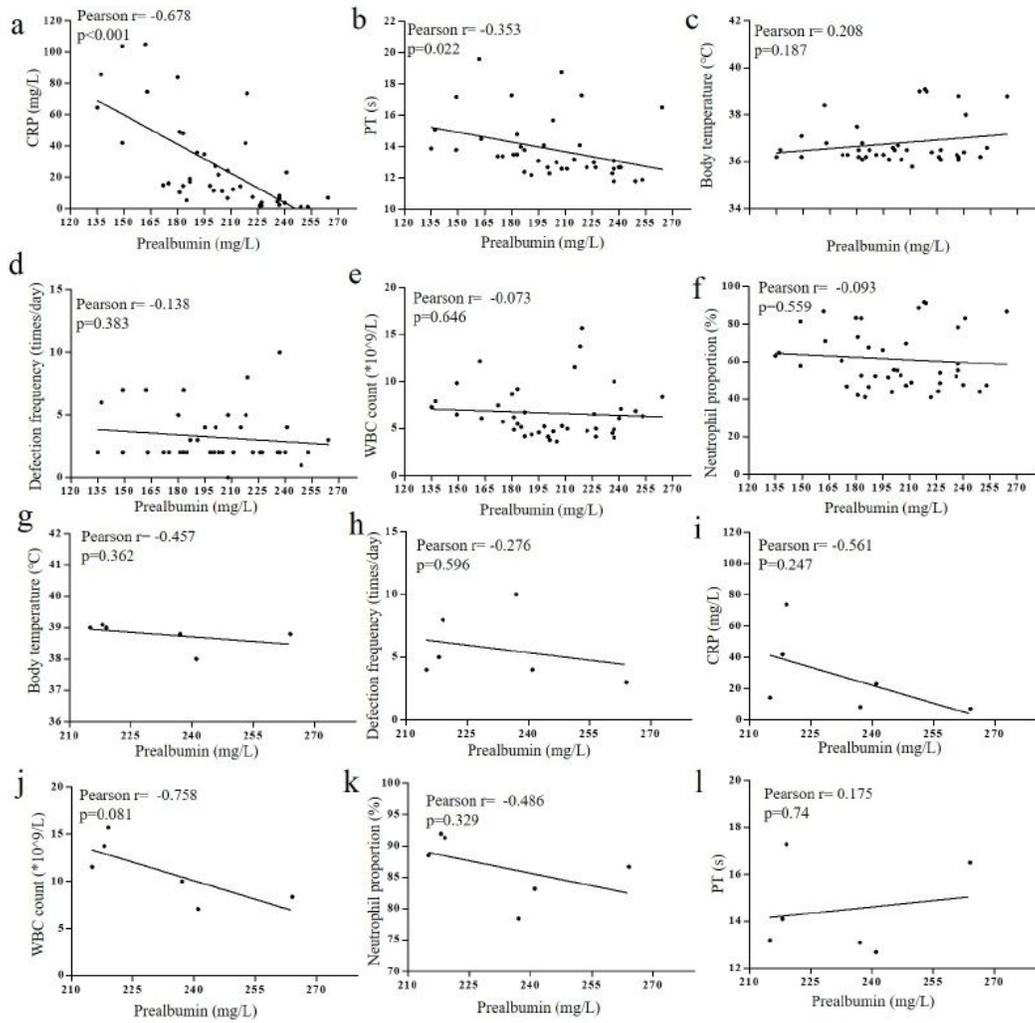


Figure 5
 Correlations between serum prealbumin and the acute inflammation status. The correlation between a Serum prealbumin levels and CRP levels, b Serum prealbumin levels and PT, c Serum prealbumin levels and body temperatures, d Serum prealbumin levels and defection frequency, e Serum prealbumin levels and WBC counts, f Serum prealbumin levels and Neutrophil percentage were analyzed. Fig. 5 a-f contained the data in the first 7 days. The correlation between g Serum prealbumin levels and body temperatures, h Serum prealbumin levels and defection frequency, i Serum prealbumin levels and CRP levels, j Serum prealbumin levels and WBC counts, k Serum prealbumin levels and neutrophil percentage, l Serum prealbumin levels and PT were analyzed. Fig. 5 g-l contained the data on the first 1 day. The Pearson correlation coefficient and the p values are stated in the corresponding figure.

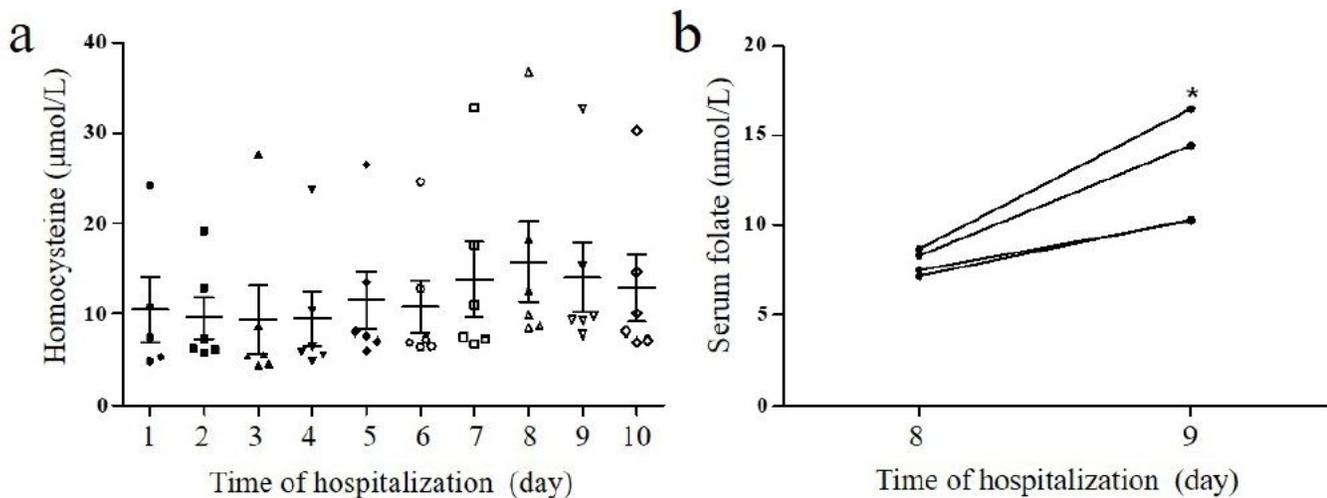


Figure 6
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The kinetics of patients' serum homocysteine during S. Enteritidis infection. a Serum homocysteine were measured in the 6 patients infected with S. Enteritidis during the disease course. b Serum homocysteine on the 8th and 9th days were measured in the patients. **: $p < 0.01$. Note: Fig. 6B didn't conclude the folate of the two patients who were administered folate plate, their folate on the 8th day was 5.8 and 7.3 nmol/L respectively, and their serum folate on the 9th day were larger than the upper limit that our machine could measure (54.6 nmol/L).

Supplementary Files

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