

Synthesis of Finely Controllable Sizes of Au Nanoparticles on a Silica Template and Their Nanozyme Properties

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Abstract

The precise synthesis of fine-sized nanoparticles is critical for realizing the advantages of nanoparticles for various applications. We developed a technique for preparing finely controllable sizes of gold nanoparticles (Au NPs) on a silica template using the seed-mediated growth and interval dropping methods. These Au NPs, embedded on silica nanospheres ($\text{SiO}_2\text{@Au}$ NPs), possess peroxidase-like activity as nanozymes and have several advantages over other nanoparticle-based nanozymes. We confirmed their peroxidase activity; in addition, factors affecting the activity were investigated by varying the reaction conditions such as concentrations of tetramethyl benzidine and H_2O_2 , pH, particle amount, reaction time, and termination time. We found that $\text{SiO}_2\text{@Au}$ NPs are highly stable under long-term storage and reusable for five cycles. Our study, therefore, provides a novel method for controlling the properties of nanoparticles and for developing nanoparticle-based nanozymes.

Introduction

Enzymes are biocatalysts that play an important role in living systems. However, they are expensive, difficult to store, laborious to produce, and are easily denatured in external environments from varying temperature, pH, and chemical stressors [1, 2]. These drawbacks critically limit their practical uses [3].

To overcome the above limitations, nanozymes have been developed as a new alternative to enzymes [1, 4]. Nanozymes are nanomaterials that possess an intrinsic enzyme-like activity and have advantages such as stability in external environments, reasonable costs, and good catalytic activity [1, 2, 5–9].

Since the discovery of the unique peroxidase-like activity of Fe_3O_4 magnetic nanoparticles (NPs) by Yan's group in 2007, numerous researchers around the world have gained an interest in nanozymes made of metal nanomaterials [10]. Metal nanomaterials, including gold (Au) NPs, platinum NPs, iron oxide NPs, cerium oxide NPs, manganese oxide NPs, and copper oxide NPs began to be used as nanozymes [5, 6, 10–15]. Most of them have exhibited enzyme-like activities like that of peroxidases, catalases, oxidases, and superoxide oxidases [1–3, 5, 6, 10, 14–22].

Among the various metal nanomaterials, Au NPs have attracted the most attention because of their outstanding catalytic properties and advantages [23–35]. Au NPs can be synthesized easily and are stable [36, 37]. In addition, the physical/chemical properties of Au NPs can be controlled by controlling their size and shape [24, 26, 33, 35]. These NPs are also highly biocompatible and are easy to functionalize [24, 30, 35, 38–42]. However, gold is not cheap, and the catalytic reactions depend on the surface area of the NPs. Fine-sized NPs can be cost-effective, but are difficult to separate for reuse [1, 3, 8–10, 17, 43, 44]. The development of a structure that uses small amounts of gold to acquire a large surface area, while still being easily separated, might prove to be a highly useful material in the field of nanozymes. A nanostructure where Au NPs are assembled onto a silica (SiO_2) sphere core was developed by the Halas group in 1998 [45]. The chemical and optical properties of the Au NP-embedded SiO_2 structure can be changed by simply controlling the diameter of the core and layered nanoparticles [46–

51]. The NP-embedded SiO₂ nanostructures are also cost-effective since only small amounts of expensive Au NPs are embedded onto the silica core, and they are easily separated from the reaction solution with the SiO₂ core.

Au NP-assembled SiO₂ nanostructures have been investigated in various fields [44, 50, 52–62] and have been found to have many merits due to the combined properties of Au NPs and of the silica core, simultaneously utilizing the outstanding and unique features of Au NPs as well as the inert and versatile feature of SiO₂ [53, 55, 56, 63]. In addition, the absorbance spectra of the nanostructures can be tuned across the visible and infrared regions by controlling the size of the Au NPs [38, 39, 64, 65]. Due to these properties, the Au NP-assembled SiO₂ nanostructures have broad applications. While there are many possible approaches for the control of the size and density of Au NPs on a template surface, a method of synthesis for those nanostructures has not yet been established [46, 48–50, 64, 65]. For this reason, the low density and non-uniform morphology of Au NPs on nanostructures remain a considerable challenge [46, 48–50]. There is, therefore, a need for the development of an improved method for preparing Au NP-assembled silica nanostructures.

Our group recently developed SiO₂@Au nanostructures in which Au NPs were densely immobilized on the surface of a SiO₂ nanosphere [66]. For this nanostructure, the SiO₂ nanosphere was used as a template and the Au NPs were uniformly and densely introduced on it using the seed-mediated growth method (Fig. 1). SiO₂@Au nanostructures have enhanced separation and re-dispersion properties and are more stable during surface modification than Au NPs. Also, they have shown potential as effective nanozymes. In this study, besides introducing dense and uniform Au NPs onto the SiO₂ nanospheres, we also developed a facile method to very precisely control the size of Au NPs on the SiO₂ surface and investigated their optical and catalytic characteristics by controlling the size of the Au NPs. Furthermore, various factors affecting the peroxidase-like activity of SiO₂@Au NPs were also studied.

Materials And Methods

Chemicals and reagents

Tetraethylorthosilicate (TEOS), tetrakis(hydroxymethyl)phosphonium chloride (THPC), chloroauric acid (HAuCl₄), 3-aminopropyltriethoxysilane (APTS), AA, PVP (MW 40,000), and TMB were purchased from Sigma-Aldrich (St. Louis, MO, USA). Ammonium hydroxide (NH₄OH, 27%), ethyl alcohol (EtOH, 99.9%), sodium hydroxide (NaOH), and sulfuric acid (H₂SO₄) were purchased from Samchun (Seoul, Korea). Hydrogen peroxide (H₂O₂) was purchased from Daejung (Siheung, Gyeonggi-do, Korea). Phosphate buffer saline containing 0.1% Tween 20 (PBST, pH 7.4) was purchased from Dynebio (Seongnam, Gyeonggi-do, Korea)

Characterization

The transmission electron microscope (TEM) images of the samples were taken using a JEM-F200 multi-purpose electron microscope (JEOL, Akishima, Tokyo, Japan) with a maximum accelerated voltage of 200 kV. The UV-Vis absorption spectra of the sample were measured by an Optizen POP UV/Vis spectrometer (Mecasys, Seoul, Korea). The centrifugation of samples was performed using a microcentrifuge 1730R (LaboGene, Lyngen, Denmark).

Synthesis of gold nanoparticles (Au NPs) assembled SiO₂ nanostructure (SiO₂@Au NPs)

The SiO₂@Au NPs were synthesized according to the previous report [66]. The colloidal Au was prepared by stirring 47.5 mL of water, 0.5 mL of 0.2 M NaOH, 12 μ L of THPC, and 1 mL of 50 mM HAuCl₄ for 1 h. Silica nanospheres (~ 160 nm) were prepared by using the modified Stöber method [83]. Briefly, 40 mL of EtOH, 1.6 mL TEOS, and 3 mL of NH₄OH were allowed to react with each other for 20 h. The amino group was introduced to the surface of 2 mg SiO₂ NPs by treating them with 62 μ L of APTS. The aminated SiO₂ NPs were incubated with colloidal Au (~ 2.5 nm) for 12 h. After several cycles of centrifugation of the mixture at 8500 rpm for 10 min, 2 mg of Au-seeded SiO₂ NPs were obtained and dispersed in 2 mL of PVP solution (1 mg/mL of PVP in water). Subsequently, 200 μ L of Au-seeded SiO₂ NPs suspension was added to 9.8 mL of PVP solution. Twenty microliters of 10 mM HAuCl₄ solution (in water, Au³⁺ precursor) and 40 μ L of AA solution, (10 mM AA in water, reducing agent) were added to the mixture in sequence. The reaction mixture was stirred for 5 min. To control the size of the Au NPs on the surface of the Au-seeded SiO₂ nanospheres, 10 mM of Au³⁺ precursor and AA were added. Until the concentrations of Au³⁺ reached 50, 100, 150, 200 and 300 μ M in the various mixtures, the same volumes of Au³⁺ precursor and AA were repeatedly added every 5 min. The SiO₂@Au NPs were washed several times with centrifugation at 8500 rpm for 10 min. The washed SiO₂@Au NPs were dispersed in 1 mL of 0.1% PBST solution to obtain a 0.2 mg/mL SiO₂@Au NP suspension.

Peroxidase-like activity of SiO₂@Au

To verify the peroxidase-like activity of SiO₂@Au NPs, 100 μ L of TMB solution (1 mM in EtOH) and 100 μ L of the various concentrations of SiO₂@Au NPs (50, 100, 150, 200, and 300 μ M, respectively) were added to 700 μ L of pH 4 buffer. Then, freshly prepared 100 μ L of H₂O₂ solution (2 M in pH 4 buffer) was added and the mixture was incubated for 30 min at room temperature. To terminate the reaction, 500 μ L of 1 M H₂SO₄ was added to each mixture and incubated for 10 min. The absorbance of the mixture at 350–800 nm was measured by using the UV-Vis spectrometer.

Peroxidase-like activity of SiO₂@Au in various reaction conditions

TMB concentration

All assays of peroxidase-like activity were carried out in 1.5 mL Eppendorf tubes at room temperature. The TMB solutions were prepared in EtOH at various concentrations (1, 2, 4, 6, 8, and 10 mM, respectively). Then, 100 μL of each TMB solution, 100 μL of 2 M H_2O_2 , and 100 μL of $\text{SiO}_2@\text{Au}$ (0.2 mg/mL) were added to 700 μL of pH 4 buffer. The final concentrations of TMB in the reaction mixture were 0.1, 0.2, 0.4, 0.6, 0.8, and 1.0 mM. After incubating the mixture for 30 min, 500 μL of 1 M H_2SO_4 was added to terminate the reaction. The absorbance of mixtures was measured using a UV-Vis spectrometer.

H_2O_2 concentration

Various concentrations of H_2O_2 solutions (1, 2, 3, and 4 M) were prepared. Next, 100 μL of 6 mM TMB solution, 100 μL of each H_2O_2 solutions, and 100 μL of $\text{SiO}_2@\text{Au}$ (0.2 mg/mL) were added to 700 μL of pH 4 buffer. The final concentrations of H_2O_2 in the mixtures were 100, 200, 300, and 400 mM respectively. After incubating the mixtures for 30 min at room temperature, 500 μL of 1 M H_2SO_4 was added to each to terminate the reaction.

pH value of the buffer

Buffers of pH 3, 4, 5, 6, 7, and 8 were prepared. Next, 700 μL of prepared each buffer were added, followed by adding 100 μL of TMB solution, 100 μL of 2 M H_2O_2 solution, and 100 μL of $\text{SiO}_2@\text{Au}$ NPs (0.2 mg/mL). The mixtures were incubated for 30 min and the reaction was terminated using 500 μL of 1 M H_2SO_4 .

Amount of $\text{SiO}_2@\text{Au}$ NPs

All reagents including 100 μL of 6 mM TMB solution, 100 μL 2 M H_2O_2 solution, and 700 μL pH 4 buffer were added into tubes. Then 1, 5, 10, 15, 20, 25, 30, 40, and 50 μg of $\text{SiO}_2@\text{Au}$ was dispersed in 100 μL PBST each and added to the mixtures, followed by incubation and termination.

Reaction time

To investigate the effect of reaction time on their peroxidase-like activity, the mixtures containing 100 μL of 6 mM TMB solution, 100 μL of $\text{SiO}_2@\text{Au}$ (0.2 mg/mL), and 100 μL of 2 M H_2O_2 solution were added to 700 μL of pH 4 buffer. Next, the mixtures were incubated for 0, 5, 10, 15, 20, 25, and 30 min respectively and terminated using 1 M H_2SO_4 .

Termination time

Mixtures including 700 μL of pH 4 buffer, 100 μL of 6 mM TMB solution, 100 μL of $\text{SiO}_2\text{@Au}$ (0.2 mg/mL), and 100 μL of 2 M H_2O_2 solution were prepared and incubated for 30 min. After adding 500 μL of 1 M H_2SO_4 , each sample was incubated for 0, 5, 10, 15, 20, 25, and 30 min to terminate the reaction.

Long-term stability of peroxidase-like activity

To investigate the long-term stability of peroxidase-like activity of $\text{SiO}_2\text{@Au}$, the experiment was repeated every day for 2 weeks and on the 31st day after their fabrication. The experimental procedures were conducted as follows: adding 100 μL of 6 M TMB solution, 100 μL of $\text{SiO}_2\text{@Au}$ (0.2 mg/mL), and 100 μL of 2 M H_2O_2 solution to 700 μL of pH 4 buffer; 30 min reaction time; 10 min for termination using 500 μL of 1 M H_2SO_4 .

Reusability as nanozymes

To investigate the reusability of $\text{SiO}_2\text{@Au}$, the peroxidase assay was performed according to the mentioned procedures. After the assay was done, the $\text{SiO}_2\text{@Au}$ NPs were collected using centrifugation at 10,000 rpm for 10 min and the absorbance of the supernatant at 453 nm was measured by using a UV-vis spectrometer. The peroxidase assay was repeated with the collected $\text{SiO}_2\text{@Au}$ NPs.

Results And Discussion

Preparation of size-controlled Au NPs-assembled silica nanostructures ($\text{SiO}_2\text{@Au}$ NPs)

The $\text{SiO}_2\text{@Au}$ NPs were prepared by using seed-mediated growth synthesis, consisting of two steps: embedding Au seeds on the SiO_2 surface, and growth of the Au NPs via the addition of an Au^{3+} precursor and reductant in intervals [45, 53]. First, the Au seeds (~ 2.5 nm) were prepared using an Au^{3+} precursor (HAuCl_4) and tetrakis(hydroxymethyl)phosphonium chloride (THPC). Then, the Au seeds were incubated with aminated SiO_2 nanospheres (~ 160 nm) to obtain Au-seeded SiO_2 nanospheres as previously reported [62, 63, 66–68]. On the Au-seeded SiO_2 nanospheres, the reduction of the Au^{3+} precursor was directly conducted using ascorbic acid (AA) which is a mild reducing agent, in the presence of polyvinylpyrrolidone (PVP) as a stabilizer. In these mildly reducing conditions, the progress of the growth stage is much slower than in strongly reducing conditions, making it easier to control the growth procedure [36]. A low concentration of the Au^{3+} precursor and AA were added onto the Au-seeded SiO_2 in 5 min intervals until the desired concentrations for fine control over the size of the Au NPs are attained. The amounts are indicated in Table 1.

First, numerous small Au NPs (~ 2.5 nm) were attached throughout the SiO_2 surface as seeds for the further growth of Au NPs, as shown in Fig. 1. Subsequently, various concentrations of the Au^{3+} precursor were added to the Au-seeded SiO_2 nanospheres at intervals; the size of the Au NPs grew larger as the

Au^{3+} concentration was increased. Also, the size of the Au NPs on the SiO_2 nanospheres was precisely controlled with high levels of density and uniformity, which were confirmed clearly as shown in Fig. 1. When the size of the Au NPs increased, the color of the solution changed in the order pale brown, then pink, purple, dark blue, and finally black (Fig. 2a). These changes occurred depending on the size and shape of the NPs due to their localized surface plasmon resonance (LSPR) [37, 46, 48]. When nanoparticles are close to one another, the absorption spectra of proximally located nanoparticles red-shifts considerably from that of solitary particles [65, 69]. Increasing the space between the particles reduces the shift [69]. Mie's theory accounts for how increasing nanoparticle diameters induces the absorption spectra to red-shift by changing the electric surface charge density of the NPs [70]. The results of our study showed that the growth of Au NPs on $\text{SiO}_2@Au$ was controlled well by the color change of the particle suspension, transmission electron microscopy (TEM) images, and absorbance according to the above-mentioned theories. The absorption spectra of SiO_2 showed that the absorbance band was red-shifted and broadened upon an increase in the concentration of the Au^{3+} precursor, indicating the formation of larger Au NPs, which change the proximate interparticle distance (Fig. 2b–c). In the absorbance band of $\text{SiO}_2@Au$ NPs-treated 0 μM Au^{3+} precursor, no peak was observed due to the exceedingly tiny size of the Au NPs. On the other hand, 50, 100, 150, 200, and 300 Au^{3+} precursor-treated $\text{SiO}_2@Au$ showed peaks at 543, 571, 593, 619, and 632 nm respectively (Fig. 2c). Also, the absorbance bandwidth broadened as the size of the Au NPs was increased.

Figure 2: (a) Optical images and (b) Size of Au NPs, and (c) UV-Vis absorption spectra of $\text{SiO}_2@Au$ NPs fabricated in various concentrations of Au^{3+} precursor: (i) 0 μM , (ii) 50 μM , (iii) 100 μM , (iv) 150 μM , (v) 200 μM , and (vi) 300 μM .

Table 1

Amount of material controlling TEM) images of gold-embedded silica nanospheres ($\text{SiO}_2\text{@Au}$ NPs) and its effect.

Sample	Au-seeded SiO_2 spheres (mg)	Au precursor (μM)	Ascorbic acid (μM)	Au size (nm)	λ_{max} (nm)	Absorbance intensity at λ_{max} (a.u.)	Suspension color
i	200	0	0	1.41	-	-	Pale brown
ii	200	50	100	4.33	543	0.27	Pink
iii	200	100	200	6.42	571	0.54	Purple
iv	200	150	300	7.33	593	0.87	Dark blue
v	200	200	400	9.56	619	1.20	Dark blue
vi	200	300	600	15.27	632	1.27	Dark blue
Sample numbers correspond to those mentioned in Fig. 1.							

Verification of the peroxidase-like activity of $\text{SiO}_2\text{@Au}$ NPs

The peroxidase-like activity of $\text{SiO}_2\text{@Au}$ NPs was evaluated through oxidation of a 3,3',5,5'-tetramethylbenzidine (TMB) substrate. The TMB oxidation reaction involves the transfer of two electrons that each produce a clear color change. When the first electron is transferred to form TMB^+ via oxidization of TMB, the TMB solution changes from colorless to blue. Since TMB^+ is quite unstable in an acidic environment, it further oxidizes to TMB^{2+} when the second electron is transferred; TMB^{2+} is stable in acidic conditions, exhibiting a yellow color and a maximum absorption peak at 453 nm (Fig. 3a)[71].

To confirm the peroxidase-like activity of the $\text{SiO}_2\text{@Au}$ NPs, TMB + H_2O_2 , TMB + $\text{SiO}_2\text{@Au}$ NPs, and TMB + H_2O_2 + $\text{SiO}_2\text{@Au}$ NPs were prepared in a pH 4 buffer for the peroxidase assay. The TMB + H_2O_2 solution was colorless and an absorbance peak at 453 nm did not appear, as shown in Fig. 2b,c. This result indicated that peroxidase-like activity did not occur in the absence of $\text{SiO}_2\text{@Au}$. Next, the color of the TMB + $\text{SiO}_2\text{@Au}$ NP sample was entirely on account of the $\text{SiO}_2\text{@Au}$ NPs, displaying an absorbance band with a maximum peak at 630 nm. However, a yellow solution and an absorbance band at 453 nm were observed in the TMB + H_2O_2 + $\text{SiO}_2\text{@Au}$ NPs sample. These results showed that $\text{SiO}_2\text{@Au}$ NPs catalyzed TMB oxidation in the presence of H_2O_2 , indicating that a peroxidase-like reaction occurred due to the peroxidase-mimicking property of $\text{SiO}_2\text{@Au}$ NPs.

The peroxidase-like activity depends on the size of the Au NPs of the $\text{SiO}_2\text{@Au}$ NPs

To investigate the correlation between the size of the Au NPs and the peroxidase-like activity of SiO₂@Au NPs, various kinds of SiO₂@Au NPs with different Au NPs sizes were prepared. The concentrations of the treated Au³⁺ precursors were 0, 50, 100, 150, 200, and 300 μM each, resulting in the formation of 1.4, 4.3, 6.4, 7.3, 9.5, and 15 nm diameter Au NPs on the SiO₂@Au structures respectively (Table 1, Fig. 2b). Each of these SiO₂@Au NPs was subjected to the TMB assay to estimate their peroxidase-like activity. An absorbance peak at 453 nm was observed in the UV-Vis absorption spectra of all samples, indicating that all of the SiO₂@Au NPs had peroxidase-like activity, irrespective of the size of the Au NPs. However, the SiO₂@Au NPs produced without any Au³⁺ precursor treatment showed relatively very weak peroxidase-like activity. This may be because the size of the Au NPs on the silica core was too small; there were a large number of vacant spaces on the SiO₂@Au NPs with the given number of Au NPs, providing an insufficient surface area for the reaction between the Au NPs and reactants. On the other hand, SiO₂@Au NPs treated with more than 50 μM Au³⁺ precursor showed high peroxidase-like activity. The size of the Au NPs was rapidly increased when concentrations of Au³⁺ precursor exceeded 50 μM as shown in the TEM images (Fig. 1a). As the size of the Au NPs on the SiO₂@Au NPs was increased, the surface area which can react with reactants also increased. Therefore, the peroxidase-like activity of the SiO₂@Au NPs was increased as the concentrations of the Au³⁺ precursor were increased (Fig. 4a, b). Even though the size of the Au NPs grew as the concentration of the Au³⁺ precursor increased, severe aggregation occurred immediately after the peroxidase reaction in the SiO₂@Au was treated with over 200 μM of the Au³⁺ precursor. Since good dispersibility is an important factor for generating a constant and stable catalytic activity, 150 μM of Au³⁺ precursor-treated SiO₂@Au NPs, which have high peroxidase-like activity without aggregation, was used in the subsequent experiments [72, 73].

Effects of reaction conditions on the peroxidase-like activity of SiO₂@Au NPs

It is known that the catalytic activity of nanozymes is affected by reaction conditions like those associated with an enzyme [10, 32, 74–77]. For this reason, the effect of reaction conditions on the peroxidase-like activity on SiO₂@Au NPs was investigated. The concentration of TMB and H₂O₂, pH of the buffer, the number of SiO₂@Au NPs, reaction time, and termination time were considered in this study. For confirming the effects of TMB concentrations, the concentrations of TMB varied from 0 to 1.0 mM while the other conditions were fixed in the peroxidase assay. The catalytic activity of SiO₂@Au NPs increased until the concentrations of TMB were 0.8 mM; they then decreased at 1.0 mM, because the poor solubility of TMB in an aqueous buffer caused precipitation during the oxidation reaction (Fig. 5a)[78]. In the case of the concentration of hydrogen peroxide, the catalytic activity of SiO₂@Au NPs was increased steeply until reaching 200 mM H₂O₂ and decreased at a concentration of 300 mM H₂O₂. Even though the activity again increased at 400 mM H₂O₂, the rate of increase was lower than 200 mM H₂O₂ (Fig. 5b). Subsequently, various buffers with different pH were subjected to the peroxidase assay. The highest activity was shown under pH 4.0, at which H₂O₂ was more stable and TMB dissolved maximally (Fig. 5c)

[10, 44, 78–81]. The velocity of the catalytic reaction showed the highest value when 20 and 25 μg of $\text{SiO}_2\text{@Au}$ NPs were treated in the sample where the rest of the conditions were fixed (Fig. 5d). About 25 min was required for the TMB^+ oxidation and 5 min for the termination of the TMB^+ oxidation to obtain stable results (Fig. 5e, f).

Long-term stability and reusability test of $\text{SiO}_2\text{@Au}$ as nanozyme

$\text{SiO}_2\text{@Au}$ NPs are substantially more advantageous over enzymes owing to their long-term stability and reusability. Denaturation during storage and their on-off usage are the major defects when using enzymes in practice [1, 2]. To verify the long-term stability of their peroxidase-like activity, the $\text{SiO}_2\text{@Au}$ NPs were examined by repeating the peroxidase assay every day at the same time for 14 days and on the 31st day after they were produced, keeping them under storage at 25°C in the meantime (Fig. 6a). The results showed that the peroxidase-like activity remained highly stable for at least 30 days after the fabrication of the $\text{SiO}_2\text{@Au}$ NPs. In sequence, the reusability of the $\text{SiO}_2\text{@Au}$ NPs as a nanozyme was evaluated through repeated peroxidase assays. Notably, the peroxidase-like activity of $\text{SiO}_2\text{@Au}$ until the fourth round was 98% of the first cycle level and mildly reduced at the fifth cycle to 89% of the first round. The reusability of $\text{SiO}_2\text{@Au}$ NPs was significantly high compared to a previous report on an Au NP-embedded silica nanostructure [82]. These results indicate that $\text{SiO}_2\text{@Au}$ NPs excel not only in being highly reusable but also in separating easily from the reaction mixture.

Conclusions

In summary, we successfully synthesized finely controllably sized Au NPs on the SiO_2 nanosphere ($\text{SiO}_2\text{@Au}$) using the seed-mediated growth method and interval dropping method under mild conditions. The effect of the size of Au NPs on the $\text{SiO}_2\text{@Au}$ NPs was confirmed by the TEM images, color changing of its suspension, and UV-Vis absorption spectra. Moreover, we investigated the factors affecting the peroxidase-like activity of $\text{SiO}_2\text{@Au}$ NPs such as TMB concentration, H_2O_2 concentration, pH, $\text{SiO}_2\text{@Au}$ NPs amount, reaction time, and termination time. Furthermore, $\text{SiO}_2\text{@Au}$ NPs showed high stability during 30 days-long storage time at room temperature and outstanding reusability for five cycles. This work is therefore meaningful for utilizing controllable nanoparticles in various fields and provides a better approach to develop nanoparticle-based nanozymes.

Declarations

Data Availability

All data generated or analyzed during this study are included in this article.

Conflicts of Interest

The authors declare that there is no conflict of interest regarding the publication of this paper.

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Ethics declarations

Ethics approval and consent to participate.

Not applicable for this study.

Consent for publication

We give our consent for the manuscript to be published in Journal of Nanobiotechnology.

Author Contributions

Conceptualization, Xuan-Hung Pham, and Bong-Hyun Jun; methodology, Bomi Seong, and Xuan-Hung Pham; investigation, Bomi Seong, and Wooyeon Kim; formal analysis, Jaehi Kim; software, Sang Hun Lee; writing—original draft preparation, Bomi Seong, Jaehi Kim; writing—review and editing, Xuan-Hung Pham, and Bong-Hyun Jun; supervisor, Bong-Hyun Jun. All authors have read and agreed to the published version of the manuscript.

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Figures

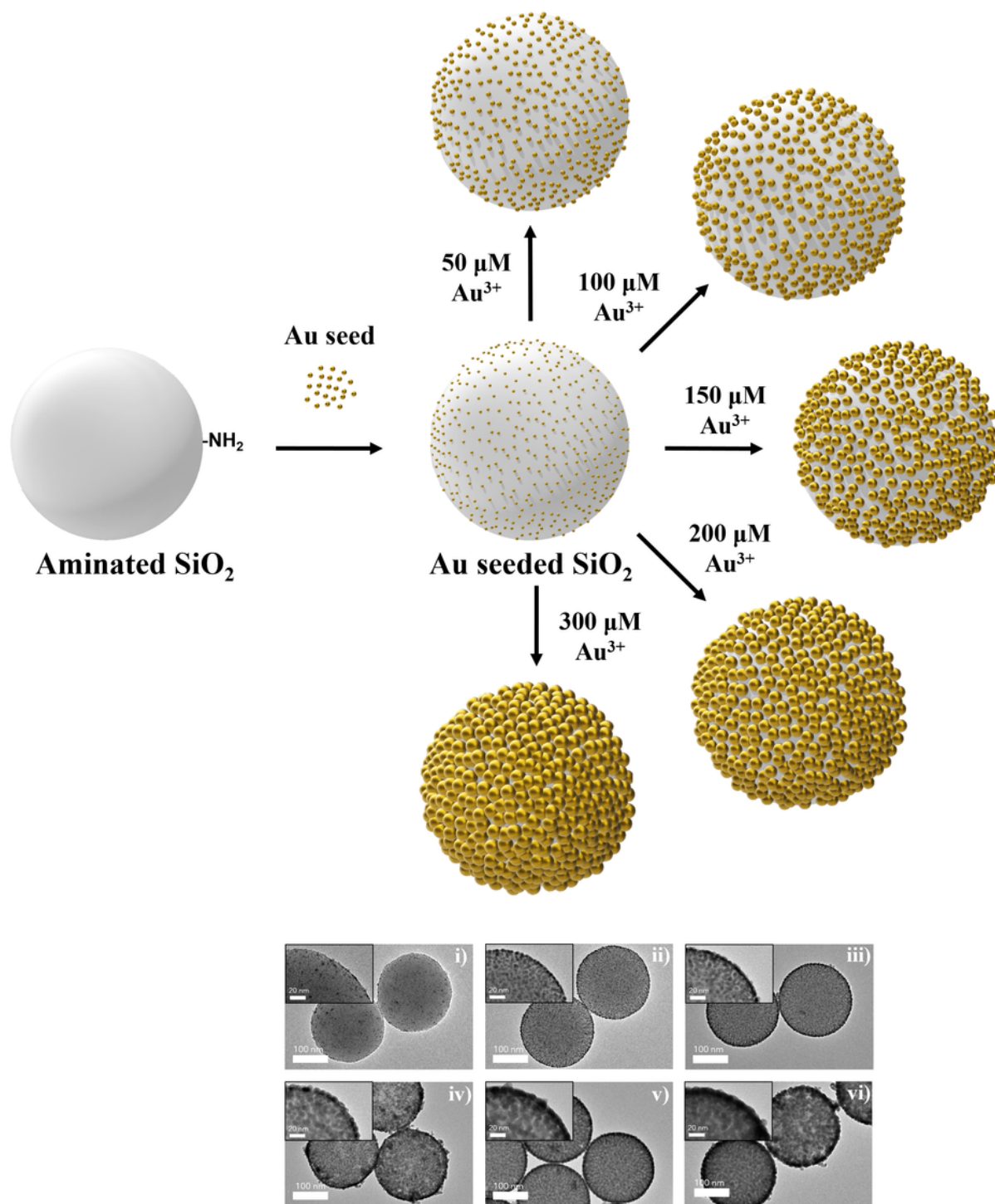


Figure 1

Gold-embedded silica nanospheres (SiO₂@Au NPs). a) A schematic representation of synthesis of SiO₂@Au NPs. b) The transmission electronic microscopy (TEM) images of SiO₂@Au NPs fabricated in various concentrations of Au³⁺: (i) 0 μM , (ii) 50 μM , (iii) 100 μM , (iv) 150 μM , (v) 200 μM , and (vi) 300 μM .

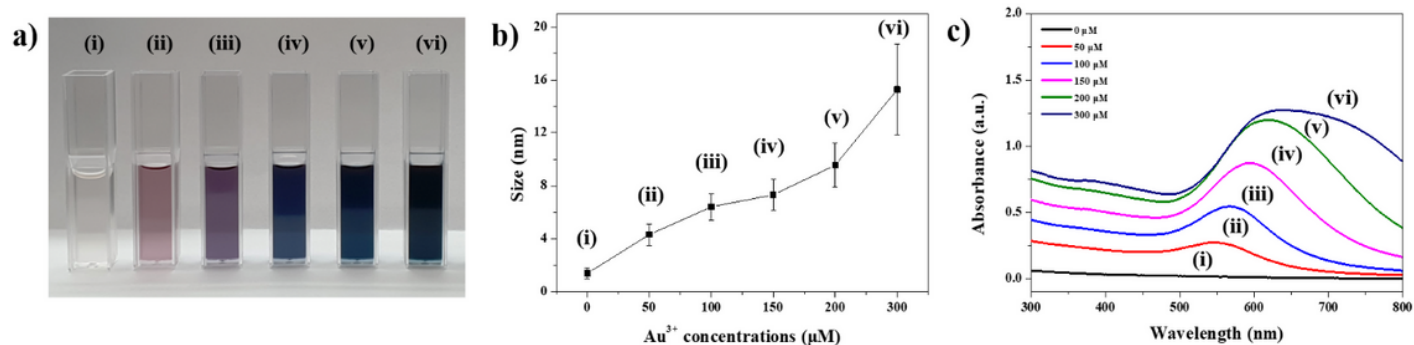


Figure 2

(a) Optical images and (b) Size of Au NPs, and (c) UV-Vis absorption spectra of SiO₂@Au NPs fabricated in various concentrations of Au³⁺ precursor: (i) 0 μM, (ii) 50 μM, (iii) 100 μM, (iv) 150 μM, (v) 200 μM, and (vi) 300 μM.

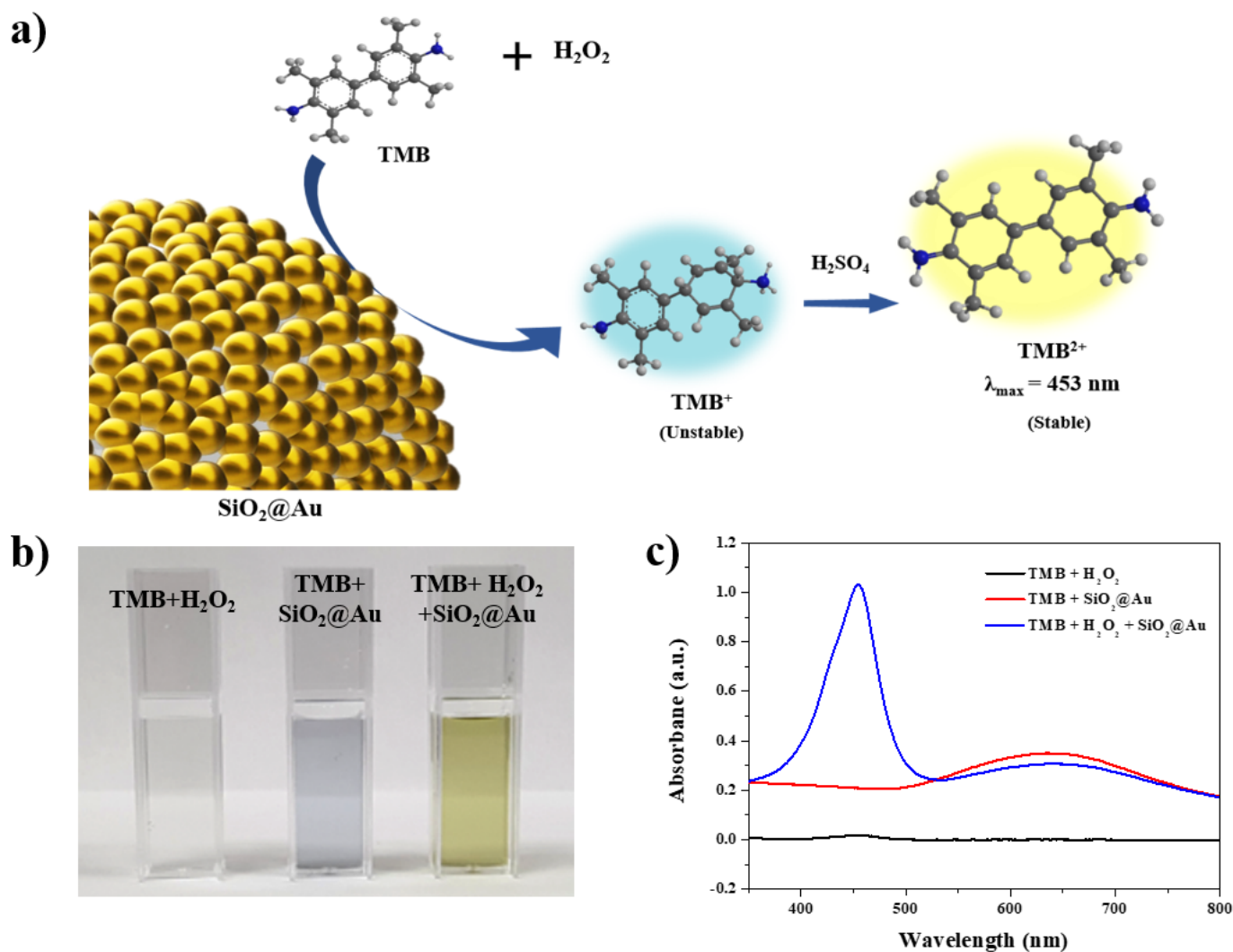


Figure 3

(a) Illustration of the peroxidase-like activity of the SiO₂@Au NPs. (b) Optical image and (c) UV-Vis absorption spectra of tetramethylbenzidine (TMB)+H₂O₂, TMB+SiO₂@Au NPs, and TMB+H₂O₂+SiO₂@Au NPs after termination with 1 M H₂SO₄ in the mixture of 1 mM TMB and 200 mM H₂O₂. The peak which appeared at 453 nm wavelength originated from the oxidized TMB substrate.

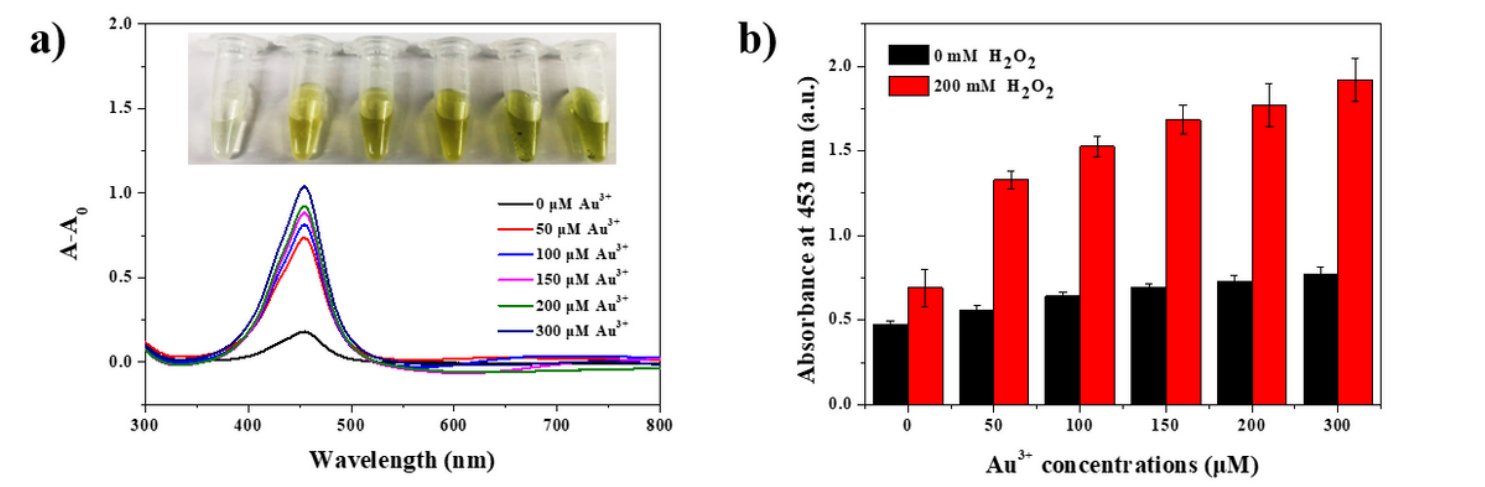


Figure 4

(a) The UV-Vis absorption spectra of subtraction sampled without H₂O₂ from samples with H₂O₂ and (b) Absorbance plots of 1 mM TMB and 200 mM H₂O₂ in the presence of various SiO₂@Au NPs fabricated in different Au³⁺ concentrations in the range of 0 to 300 μM.

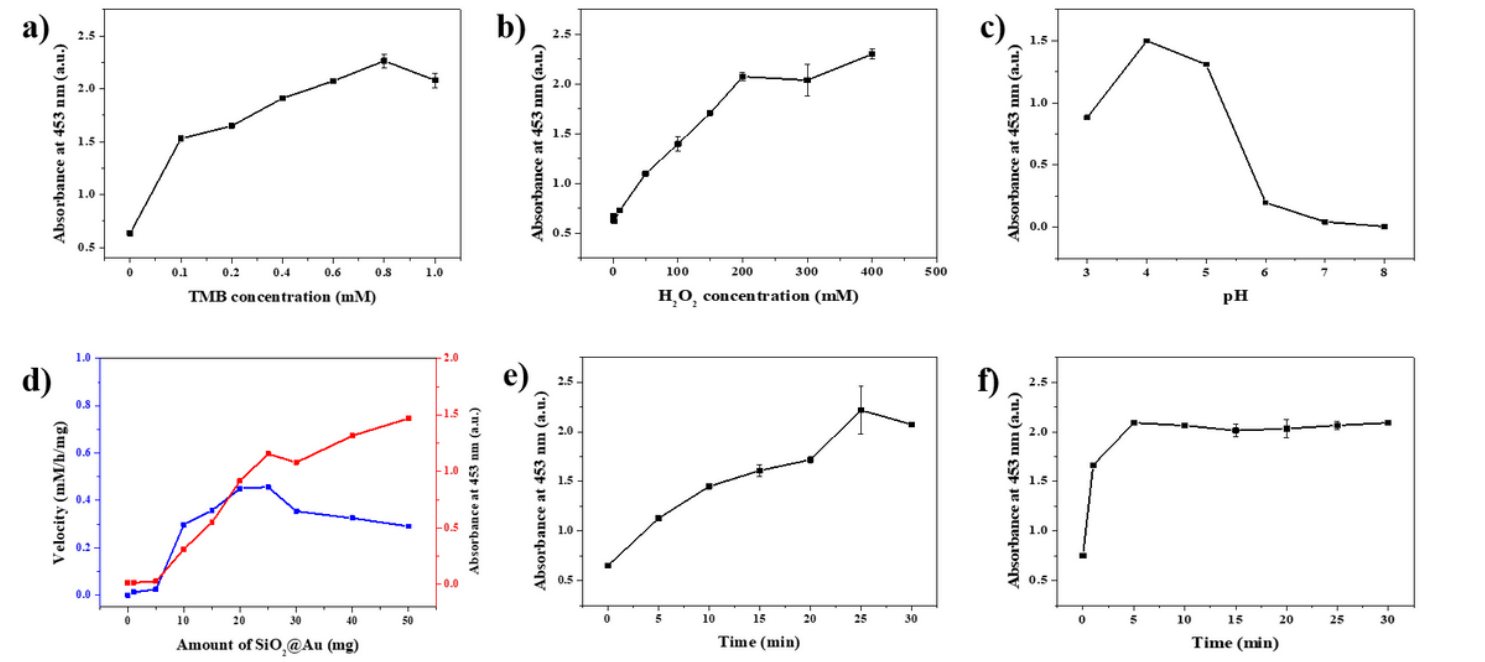


Figure 5

The effects of different reaction conditions on the peroxidase-like activity of SiO₂@Au NPs in a mixture of TMB and H₂O₂. (a) TMB concentration, (b) H₂O₂ concentration, (c) pH value of the solution, (d) the

velocity and amount of SiO₂@Au, (e) reaction time, and (f) termination time.

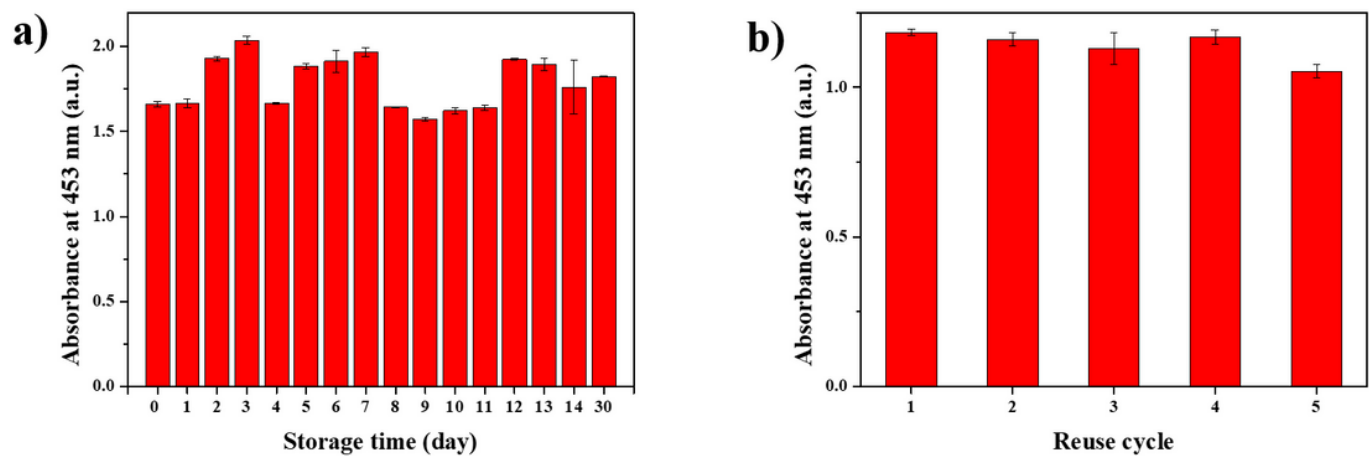


Figure 6

(a) Long-term stability of the peroxidase-like activity and (b) reusability of SiO₂@Au NPs in a mixture of 0.6 mM TMB, 200 mM H₂O₂, and pH 4 buffer. The SiO₂@Au NPs were stored in PBST (0.1%) at room temperature.

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