

Severity of radiation pneumonitis, from clinical, dosimetric and biological features: a pilot study.

Samantha Aso

Hospital Universitari de Bellvitge

Arturo Navarro-Martin (✉ artnama79@gmail.com)

Institut Catala d' Oncologia <https://orcid.org/0000-0002-1327-5367>

Richard Castillo

Emory University Winship Cancer Institute

Susana Padrones

Hospital Universitari de Bellvitge

Edward Castillo

Beaumont Health

Ana Montes

Universitat de Barcelona

Jose Ignacio Martinez Ballarin

Hospital Universitari de Bellvitge

Noelia Cubero

Hospital Universitari de Bellvitge

Rosa Lopez

Hospital Universitari de Bellvitge

Laura Rodriguez

Hospital Universitari de Bellvitge

Ramon Palmero

Institut Catala d' Oncologia

Federico Manresa

Hospital Universitari de Bellvitge

Thomas Guerrero

Beaumont Health

Maria Molina

Hospital Universitari de Bellvitge

Research

Keywords: non-small cell lung cancer, radiation pneumonitis, lung function, wound healing.

Posted Date: August 21st, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-57934/v1>

License:  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Version of Record: A version of this preprint was published on October 27th, 2020. See the published version at <https://doi.org/10.1186/s13014-020-01694-1>.

Abstract

Background and objective: Radiation pneumonitis (RP) could be a lethal complication of lung cancer treatment. No reliable predictors of RP severity have been recognized. This prospective pilot study was performed to identify early predictors of high grade lung toxicity and to evaluate clinical, biological or dosimetric features associated with different grades of toxicity.

Method: Sixteen patients with non-small cell lung cancer with indication of concurrent chemoradiotherapy using 60Gy/2Gy/fraction starting at cycle one of platinum based chemotherapy were included. Bronchoalveolar lavage (BAL), pulmonary function testing (PFT), and ¹⁸F-2-fluoro-2-deoxy-D-glucose positron-emission tomography was performed before radiotherapy (RT), after three weeks of treatment, and two months post-RT. For analysis, patients were grouped by grade (low [G1-G2] vs. high [G3-G5]). The two groups were compared to identify predictors of RP. Protein expression BAL and lung tissue metabolism was evaluated in two patients (RP-G1 vs. RP-G3). Categorical variables such as comorbidities, stages and locations were summarized as percentages. Radiation doses, pulmonary function values and time to RP were summarized by medians with ranges or as means with standard deviation. Longitudinal analysis PFT was performed by a T-test.

Results: All 16 patients developed RP, as follows: G1 (5 pts; 31.3%); G2 (5 pts; 31.3%); G3 (5 pts; 31.3%); and G5 (1 pts; 6.1%). Patients with high grade RP presented significant decrease ($p=0.02$) in diffusing lung capacity for carbon monoxide (DLCO) after three week of RT. No correlation between dosimetric values and RP grades was observed. BAL analysis of the selected patients showed that CXCL-1, CD154, IL-1ra, IL-23, MIF, PAI-1 and IFN-g were overexpressed in the lungs of the RP-G3 patient, even before treatment. The pre-RT SUVmax value in the RP-G3 patient was non-significantly higher than in the patient with RP-G1.

Conclusions: RT induces some degree of RP. Our data suggest that decrease in DLCO% is the most sensitive parameter for the early detection of RP. Moreover, we detect biological differences between the two grades of pneumonitis, highlighting the potential value of some cytokines as a prognostic marker for developing high grade lung toxicity. Further multicenter studies with larger sample size are essential to validate these findings.

Introduction

Radiotherapy (RT) is a mainstay treatment for non-small cell lung cancer (NSCLC). Several studies have showed a benefit in local control and survival increasing biological equivalent doses¹ However, its effectiveness is limited by the risk of radiation-induced lung injury (RILI). RILI is the result of an abnormal healing response to lung irradiation caused by damage to parenchymal cells, vasculature, and/or stroma followed by inflammatory cytokine release.² Diagnosis of RILI is based on nonspecific symptoms with or without abnormalities in pulmonary function tests (PFT). Radiographic changes usually reveal parenchymal abnormalities.

Radiation pneumonitis (RP), and pulmonary fibrosis (PF) represent, the acute and late phase of RILI, which has been described in 30% of cases, with mortality rates as high as 2%.³⁻⁶ Distinctions between these phases is arbitrary because early and late effects of RT are a continuous spectrum of the same biological event. Early-RILI or RP is considered when symptoms appear within 12 week after lung RT and up to 6 months post-RT. X-rays is characterized by inhomogeneous opacity inside or outside the irradiation field and increased density of septal structures. Late-RILI or PF is a chronic lung damage that usually evolves over 6 to 24 months after RT. X-rays shows contracted, dense scar that occupies a much smaller volume than the originally irradiated volume. Also fibroelastosis pleuroparenchymal changes can be observed do to RT.⁷

The relationship between the development of RILI and baseline patient characteristics, lung function parameters and radiation dose have been retrospectively investigated.^{3,8,9} Some molecular biomarkers in blood and bronchoalveolar lavage (BAL) have been proposed.¹⁰⁻¹² In addition, imaging technologies such as ¹⁸F-2-fluoro-2-deoxy-D-glucose (¹⁸F-FDG) positron-emission computed tomography (PET/CT) are able to quantify the uptake of ¹⁸F-FDG in the lung as a marker of pulmonary inflammation.¹³⁻¹⁷ Despite these advances, scarce data is available to show reliable predictive factors of RP, the effects of radiation on the lung, or the mechanisms leading to fibrosis and death in the context of RILI.

This prospective pilot study was conducted to identify early predictive factors of severity in RP and to evaluate the possible features associated with different grades of RP.

Methods And Material

Patients

Patients diagnosed with NSCLC at our Institution with indication of concurrent chemoradiotherapy regimen using 60 Gy 2Gy/fraction starting at cycle one of platinum based chemotherapy were prospectively included from January 2011 to March 2013. Inclusion criteria comprised: histologically confirmed NSCLC, inoperable locally advanced NSCLC, no previous thoracic RT. Exclusion criteria included: Karnofsky index<70, interstitial lung disease (ILD), forced expiratory volume at first second (FEV1) <30%, chronic respiratory failure, oral corticosteroid treatment, contraindication for bronchoscopy, or refusal to participate. The Ethics Committee of the University Hospital of Bellvitge and the Catalan Institute of Oncology approved the study protocol (PR206/08). Patients signed a written informed consent prior to inclusion.

Patients underwent BAL by fiberoptic-bronchoscopy, lung function testing, and ¹⁸F-FDG-PET/CT prior to initiation of RT, at the end of the third week of RT, and at two months post-RT. Patient consultations were once weekly from the time of study inclusion until RT completion. Thereafter, patients were evaluated every 15 days for 6 months and then monthly for one year. The follow-up visits included: medical history, physical examination and monthly chest X-rays. RP diagnosis was based on the appearance or worsening of dyspnea and cough, which may associate fever or chest pain, accompanied with changes

of radiological images. RP diagnosis and imaging evaluation were made by the multidisciplinary clinical team (medical oncologist, radiation oncologist and thoracic radiologist). RP grade was scored according to the Common Terminology Criteria for Adverse Events version 4.0. (CTCAEv4.0).¹⁸ Patients were divided into 2 groups (low-grade RP [G1 and G2], and high-grade RP [G3-G5]), according to the CTCAEv4.0. The two groups were compared to identify early predictors for high-grade RP development.

Radiotherapy treatment

Treatment planning for the RT used a 3D technique. An specific CT scan over the thorax and upper abdomen with intravenous contrast was obtained.¹⁹ Gross tumor volume (GTV) was contoured according to the PET/CT and diagnostic CT scan. No prophylactic nodal irradiation was performed. To cover subclinical disease, the GTV was expanded according to histological findings. The GTV was increased by 0.6 cm for squamous cell carcinomas and by 0.8 cm for adenocarcinomas to provide the clinical target volume (CTV).²⁰ The planning target volume (PTV) was determined by adding 0.7 cm to the CTV in the lateral and anterior posterior direction and 1.5 cm in the cranio-caudal direction.²¹ The mean dose to the PTV ranged from 60 Gy to 66 Gy according to standard protocols.²² Organs at risk were contoured in accordance with treatment guidelines.²³ Dose constrains to the lungs were V20 < 35% (i.e., 35% of the healthy lung should receive \leq 20Gy) with a mean dose < 19Gy. Radiotherapy and chemotherapy were started at the same time.

Pulmonary Function Testing

PFT parameters were measured according to European Respiratory Society guidelines²⁴ using computerized lung function testing equipment (Body Box 5500; Morgan Scientific). The parameters assessed included forced vital capacity (FVC), forced expiratory volume at first second (FEV1), and diffusing lung capacity for carbon monoxide (DLCO). The same technician performed the PFT at all follow-up consultations.\

BAL sample collection

BAL was performed in both lungs (i.e., the irradiated and non-irradiated) by using four 40 mL aliquots of 0.45% saline solution per wash through a fiberoptic-bronchoscope (Olympus BF-160). The first sample was discarded; the second and the third samples were mixed and sent for cytological evaluation. The fourth aliquot was centrifuged (543.6 g x 5 min) into cellular fraction and supernatant, which was aliquoted for cytokine determination; both fractions were frozen at -80 °C.

Protein array analysis of cytokine in BAL supernatant

To evaluate differences in individual predisposition to lung damage and tissue repair response, we evaluate BAL samples. In this preliminary report, we chose one representative patient from each study group: one from low grade RP (RP-G1) and another from high grade RP (RP-G3). Expression protein was assessed using the Human Cytokine Array Panel A (R&D Systems, Minneapolis, MN; USA). Protein concentration was measured in each sample.²⁵ Pixel density of the spots was analysed using the Multi Gauge V3.0 (FujiFilm, Palo Alto, CA; USA). Three independent readers compared the mean pixel density values in the spots.

Positron-Emission Computed Tomography

Patients underwent PET/CT imaging with ^{18}F -FDG according to standard practice.²⁶ They were asked to fast 6 hours prior to the imaging session to ensure fasting blood glucose levels within the normal range (3.3–5.6 mmol/L). Patients received an intravenous administration of FDG per kg of body weight. The ^{18}F -FDG-PET/CT scan was performed with a hybrid PET/CT scanner (General Electric Discovery ST). The whole-body acquisition protocol included a CT scan and a PET scan in a three-dimensional mode. No iodine intravenous contrast was administered. The CT data were used for attenuation correction and anatomic location of PET findings. The standardized uptake value (SUV) was used to measure uptake in the lungs.

Image analysis

Pre-treatment PET/CT image analyses of the two patients (i.e., RP-G1 and RP-G3) were performed to screen for possible differences in the SUV. This analysis was processed by three independent readers and evaluated using custom Matlab software (v2011a, Mathworks, Inc; Natick, MA; USA). The lung region of interest (ROI) was segmented semi-automatically. Overlap of central airway, liver, heart, diaphragm, and tumor in the lung ROI were manually removed. The resulting binary lung ROI was used for the analysis. The SUV was calculated from the PET attenuation corrected emission images.²⁷ SUV of voxels in the lung ROI were binned into histograms; the maximum SUV (SUVmax) was calculated according to the formula described by Petit et al.¹⁵

Statistical Analysis

Since the prevalence of RP is variable due to differences in diagnostic scales⁴, sample size was calculated using the “observed versus a reference mean”²⁸, which includes the reported prevalence of RP among the interstitial lung disease.²⁹ To detect a relevant clinical difference (alpha = 0.05 and beta = 0.1), 17 patients were required (assuming a follow-up loss rate of 20%).

Patients were divided into 2 groups (low-grade [G1 and G2], and high-grade [G3-G5]), according to the CTCAEv4.0. The two groups were compared to identify predictors for the early identification of RILI. Categorical variables were summarized as percentages. Ordinal categorical variables were summarized by medians with ranges or as means with standard deviation (SD). Longitudinal analysis of FEV1(%) and DLCO(%) was performed by a T-test. Differences were considered statistically significant for $p < 0.05$. All plots and analyses were performed using the statistical software R version 3.2.1 for Windows.

Results

Population

Seventeen patients were invited to participate and one refused. A total of 16 patients were included. Table 1 shows the characteristics of the cohort. All patients developed RP, with different grades of severity distributed as follows: 5 patients, G1 (31.3%); 5 patients, G2 (31.3%); 5 patients, G3 (31.3%); and

1 patient, G5 (6.1%). Patients were grouped by RP grade (low [G1-G2] vs. high [G3-G5]), with 10 and 6 patients in each group, respectively. Four patients from the high-grade group developed RILI in both lungs.

Table 1
Clinical and treatment characteristics of the sample

Characteristics	
No. of patients	16
Sex	14 (87.5%)
Male	2 (12.5%)
Female	
Age (range)	63 (58.8–76)
Smoking history	10 (62.5%)
Current	5 (31.2%)
Former	1 (6.2%)
Never	
Pulmonary Function (range)	101 (87-105.8)
FVC (%)	85.5 (71.5–93.3)
FEV1 (%)	67.9 (60.5–73)
FEV1/FVC	71 (57.2–87.5)
DLCO (%)	
Comorbidities	5 (31.2%)
Hypertension	3 (18.8%)
Diabetes	11 (68.8%)
COPD	2 (12.5%)
Heart disease	2 (12.5%)
Vascular disease	

FVC: forced vital capacity FEV1: forced expiratory volume in one second

DLCO: diffusing lung capacity for carbon monoxide COPD: chronic

obstructive pulmonary disease NSCLC: non-small cell lung cancer

Characteristics	
Histologic type	4 (25%)
Adenocarcinoma	9 (56.2%)
Squamous cell carcinoma	2 (12.5%)
Large cell neuroendocrine carcinoma	1 (6.2%)
NSCLC	
Location	1 (6.2%)
Mediastinum	3 (18.8%)
Hilar	6 (37.5%)
Right upper lobe	2 (12.5%)
Right inferior lobe	3 (18.8%)
Left superior lobe	1 (6.2%)
Left inferior lobe	
Radiation doses (range)	17.2 (12.7–22.9)
Mean dose (Gy)	30 (20.3–35.8)
V20 (%)	60 (47.5–65.8)
V5 (%)	
Pneumonitis grades	5 (31.3%)
1	5 (31.3%)
2	5 (31.3%)
3	0 (0%)
4	1 (6.1%)
5	
FVC: forced vital capacity FEV1: forced expiratory volume in one second	
DLCO: diffusing lung capacity for carbon monoxide COPD: chronic	
obstructive pulmonary disease NSCLC: non-small cell lung cancer	

Table 2 shows the patient characteristics by RP group (low vs. high grade). There were no significance differences between the groups in terms of age, gender, comorbidities, PFT baseline values, cancer histology, stage and tumor localization. No differences were observed in the time of onset of RP and

severity ($p = 0,6642$). The mean radiation dose was higher in the high-grade group [18(15.2–20.1) vs. 16.1 (12-22.2)]; however, V20 was lower in the high-grade group [29.5 (95% CI 23–30) vs. 32 (95% CI 21–35)]. No significant correlation between dosimetric values and RP grades was observed.

Table 2
Patient characteristics according to pneumonitis grade: low vs. high grade

Characteristics	Low grade (G1-G2)	High grade (G3-G5)
No. of patients	10 (62.5%)	6 (37.5%)
Sex	9 (90%)	5 (83.3%)
Male	1 (10%)	1 (16.7%)
Female		
Age (range)	66.0 (59.2–67)	59.5 (58.2–65.2)
Smoking history	8 (80%)	2 (33.3%)
Current	2 (20%)	3 (50%)
Former	0 (0%)	1 (16.7%)
Never		
Pulmonary Function (range)	100,5 (88.5- 106.5)	102.5(82.5-104.8)
FVC (%)	81.5 (59–93)	89 (77-90.5)
FEV1 (%)	64 (55-71.6)	69.5 (67.9–80.3)
FEV1/FVC	71 (46.8–83.5)	71.5 (63.5–92.2)
DLCO (%)		
Comorbidities	3 (30%)	2 (33.3%)
Hypertension	0 (0%)	3 (50%)
Diabetes	7 (70%)	4 (66.7%)
COPD	1 (10%)	1 (16.7%)
Heart disease	2 (20%)	0 (0%)
Vascular disease		
Histologic type	3 (30%)	1 (16.7%)
Adenocarcinoma	5 (50%)	4 (66.7%)
Squamous cell carcinoma	2 (20%)	0 (0%)
Large cell neuroendocrine carcinoma	0 (0%)	1 (16.7%)
NSCLC		

FVC: forced vital capacity FEV1: forced expiratory volume in one second DLCO: diffusing lung capacity for carbon monoxide COPD: chronic obstructive pulmonary disease NSCLC: non- small cell lung cancer

Characteristics	Low grade (G1-G2)	High grade (G3-G5)
Clinical Stage	3 (30%)	-
IIB	6 (60%)	3 (50%)
IIIA	1 (10%)	5 (50%)
IIIB		
Location	1 (10%)	0 (0%)
Mediastinum	2 (20%)	1 (16.7%)
Hilar	4 (40%)	2 (33.3%)
Right upper lobe	1 (10%)	1 (16.7%)
Right inferior lobe	1 (10%)	2 (33.3%)
Left superior lobe	1 (10%)	0 (0%)
Left inferior lobe		
Chemotherapy agents	1 (10%)	1 (16.7%)
Carboplatin-Etoposide	1 (10%)	0 (0%)
Carboplatin-Gemcitabina	2 (20%)	0 (0%)
Cisplatin-Vinorelbine	6 (60%)	5 (83.3%)
Cisplatina-Etoposide		
Radiation doses (range)	16.1 (12-22.2)	18 (15.2–20.1)
Mean dose(Gy)	32 (21-35.8)	29.5(23–30)
V20 (%)		
Onset of radiation pneumonitis (median)	68,5 days	111 days
FVC: forced vital capacity FEV1: forced expiratory volume in one second DLCO: diffusing lung capacity for carbon monoxide COPD: chronic obstructive pulmonary disease NSCLC: non- small cell lung cancer		

Pulmonary Function Testing

Baseline FEV1(%) and DLCO(%) at diagnosis was not associated with the different grade of RP development. No significant differences in mean values were found between the groups at the three time points (baseline, end of week three, and at two month post-RT) (Tables 3 and 4). However, by the end of the third week of RT, the DLCO(%) had decreased substantially in those cases that developed high-grade group (p = 0.0203) (Fig. 1-B). Furthermore, the DLCO(%) decline was even worse after 2 months post-RT in

that same group ($p = 0.0342$) (Fig. 1-B). A FEV1(%) decline was observed after 2 months of RT but not earlier during the treatment (Fig. 1-A). Therefore, the DLCO(%) decrease during the RT allows to predict a high-grade RP even before starting respiratory symptoms.

Table 3
Mean values in FEV1 between CTCAEv4.0 groups at baseline, end of third week, and two month post-RT

Mean values	Low grade (G1-G2)	High grade (G3-G5)	p-value
Baseline	78.2%(SD 21.2)	88.5% (SD: 19.1)	0.2803
3 weeks of RT	93.4% (SD 17.5)	86.2% (SD: 22.4)	0.0668
2 months post- RT	78.7% (SD: 12.0)	76.4% (SD 11.8)	0.3015
FEV1: forced expiratory volume in one second. CTCAEv4.0: Common Terminology Criteria for Adverse Events version 4.0. RT: radiotherapy. SD: standard deviation.			

Table 4

Mean values in DLCO between CTCAEv4.0 groups at baseline, end of third week, and two month post-RT

Mean values	Low grade (G1-G2)	High grade (G3-G5)	p-value
Baseline	67.7%(SD 22.4)	83.3% (SD: 30.2)	0.0719
3 weeks of RT	62.8% (SD 11.5)	65.6% (SD: 21.6)	0.2136
2 months post- RT	57% (SD: 13.6)	55.2% (SD 16.7)	0.0595
DLCO: diffusing lung capacity for carbon monoxide. CTCAEv4.0: Common Terminology Criteria for Adverse Events version 4.0. RT: radiotherapy. SD: standard deviation.			

Protein array analysis in the BAL supernatant

Different cytokine and chemokine expression profile (pre-RT and week 3) was found in a patient with RP-G1 compared to another with RP-G3 (Fig. 2). Before RT (Fig. 2-A), the only protein expressed in the tumor-free lung of the RP-G1 patient was ICAM while the tumor-free lung of the RP-G3 patient presented an overexpression of CD154, CXCL-1, ICAM, IFN- γ , IL-1ra, IL-23, MIF and PAI-1. In the lung with tumor (Fig. 2-A), the RP-G1 patient expressed CXCL-1, ICAM, IL-1ra and MIF while the RP-G3 patient showed those same cytokines but also expressed CD154, IL-23, IFN- γ and PAI-1. At the end of third week of RT (Fig. 2-B), a change in the cytokine and chemokine patterns was detected in both cases. RT induced expression of CD154, CXCL-1, IL-1ra, IL-23, MIF, and PAI-1 while reducing ICAM expression in both lungs of the RP-G1 patient. RT increased cytokine response of the most overexpressed proteins in the tumor and tumor-free

lungs of the RP-G3 patient, with a higher expression of CD154, CXCL1, IL-1ra, IFN- γ , IL-23 and PAI-1 (Fig. 2-B).

PET/CT Image Analysis

The pre-RT SUVmax value was calculated for the RP-G1 and RP-G3 patients. In both patients, the pre-treatment SUVmax was higher than normal (standardized lung SUVmax values, 0.05 ± 0.17).³⁰ However, the RP-G3 patient had a non-significantly higher SUVmax (2.20 vs. 2, respectively).

Discussion

The present study demonstrates that RT induces RILI in all patients who undergo external RT with variable clinical manifestations and different degrees of lung damage. This variability in the degree of RILI suggests that severe RP may be associated with differences in individual predisposition. The early decrease of DLCO(%) was a predicting factor of severe RP development and thus could serve as a marker for early diagnosis and treatment modification.

The reported prevalence of RP in lung cancer patients ranges from 0–58%.⁴ This variation is likely due to differences in diagnostic scales, non-specific symptoms, and the lack of standardized assessment protocols.⁴ In the present study, standard follow-up protocols detected RILI in all of the patients but with different severity presentation. This finding suggests that all patients treated with RT are likely to develop RILI to a greater or lesser extent, which may depend on individual biological characteristics.

Previous studies have shown that subtle, local changes in PFT after RT can be used as indicators of acute and chronic lung damage, although published results are not always consistent.^{3,9} The largest and most consistent changes in PFT values after RT are observed in DLCO, which has been directly associated with respiratory morbidity.³¹ In the present study, we evaluated the mean differences in FEV1(%) and DLCO(%) between low-grade RP patients [G1-G2] and high-grade RP patients [G3-G5]. We found no statistically significant differences between the two groups in FEV1(%) values at the different time points, thus leading us to conclude that FEV1(%) is not a predictor of RP, a finding that is consistent with other reports.^{9,31} By contrast, we found that a decrease in DLCO(%) 2 month post-RT was predictive of RP severity, in line with some previous reports.^{9,31} Importantly, the decline in DLCO(%) after three weeks of RT was an early predictor of severe RP. The explanatory power of this variable could be that histopathological alterations present during the latency phase (i.e., without clinical manifestations) for RILI may alter gas exchange.³ Clearly, the ability to early predict severe RILI would be helpful to optimize therapeutic options.

Classically, RP grade has been associated with the radiation dose.^{3,8} In our study, the high-grade group received a higher mean radiation dose but lower V20 than the low-grade group. This finding is contrast with the meta-analysis published by Palma et al.³² The absence of a significant association in this study between RP grade and radiation parameters could be due to the limited sample size, the small differences among patients with regards to the radiation dose and biological predisposition.⁹

RP increases cellular metabolism. ¹⁸F-FDG-PET/CT quantify this metabolic increase; indeed, ¹⁸F-FDG uptake during RT and post-treatment is a marker of symptomatic RP.¹³⁻¹⁷ Our results suggest that a high pre-treatment SUVmax is associated with high grade RP. Accordingly, Castillo et al demonstrated the predictive value of pre-treatment ¹⁸F-FDG lung uptake in the subsequent development of RP symptoms.¹⁶⁻¹⁷

RILI produce an imbalance between type 1 and type 2 helper T-cells and abnormal fibroproliferative wound healing³³. Variations among patients in terms of RP severity and lung repair capacity could be related to individual pre-treatment lung biomolecular conditions and genetic factors³⁴. In the present study, increased expression of some mediators were obtained in BAL of tumor lungs in both patients (RP-G1 and RP-G3), although these proteins were also expressed in the tumor-free lung of the RP-G3 patient before RT. Previous studies indicate that IL-1ra is involved in acute inflammation³⁵; MIF modulates RILI³⁶; and CXCL1 promotes angiogenesis and thus contribute to the pathogenesis of PF.³³ Interestingly, CD154, IFN- γ , IL-23 and PAI-1 were expressed in both lungs (tumor and tumor-free lung) before RT only in the RP-G3 patient. These four cytokines have been described in animal models of lung fibrosis.^{37, 38} Furthermore, a recent study reported that a truncated PAI-1 protein protects against RILI in a murine model.³⁹ Finally, Liu et al. found that rs7242 GT/GG genotypes located in the 3'UTR of PAI-1 were associated with a significantly increased risk of RP.⁴⁰ Overall, our findings suggest a potential biological predisposition to lung damage and altered wound healing in RILI development, which would deserve a depth study to better understand pathogenesis.

Study strengths and limitations.

We have to recognize some limitations: First, the small sample size which was calculated using "observed versus a reference mean" and even although we have enrolled sixteen patients, instead of including seventeen, we have not had any loss of follow up. Secondly, the low power of the biological lung analysis (only two patients). In this sense, this is a pilot study to identify if there are differences in biological features associated with different grades of RP. Our findings, warrant further investigation in a larger sample. The main strength of the study is that it is the first prospective study to evaluate patients with NSCLC through a longitudinal clinical and biological follow-up that demonstrate RT induces RILI in all cases but in some of them with a high-grade of lung injury and consequent altered wound repair.

Conclusion

RT treatment always induces some degree of lung injury and the extent of the damage is variable. Our data suggest that decrease in DLCO% is the most sensitive parameter for the early detection of severe RP. Moreover, we detect biological differences between the two grades of pneumonitis, highlighting the potential value of cytokines such as CXCL-1, CD154, IL-1ra, IL-23, MIF, PAI-1 and IFN- γ as a prognostic marker for developing high grade of lung toxicity. Further multicenter studies with larger sample size are essential to validate these preliminary findings.

Abbreviations List

Common Terminology Criteria for Adverse Events version 4.0: CTCAEv4.0

Chemokine (C-X-C motif) ligand 1: CXCL1

Clinical target volume: CTV

Grade 1: G1

Grade 2: G2

Grade 3: G3

Grade 5: G5

Gross tumour volume: GTV

Intercellular Adhesion Molecular: ICAM

Interferon-gamma: IFN-g

Interleukin-23: IL-23

Interleukin-1 receptor antagonist: IL-1ra

Macrophage migration inhibitory factor: MIF

Planning target volume: PTV

Plasminogen activator inhibitor type 1: PAI-1

Radiotherapy: RT

Radiation pneumonitis grade 1: RP-G1

Radiation pneumonitis grade 3: RP-G3

Region of interest: ROI

Transforming growth factor β -1: TGF- β 1

V20: healthy lung that should receive \leq 20Gy

Declarations

Ethics approval and consent to participate: The Ethics Committee of the University Hospital of Bellvitge and the Catalan Institute of Oncology approved the study protocol (PR206/08). Patients signed a written informed consent prior to inclusion.

Consent for publication: not applicable

Availability of data and materials: the datasets are available to all interested researchers on reasonable request from corresponding author.

Competing interests: **M.M.**: she's consulting of Esteve-Teijin, Boehringer Ingelheim, Roche, Chiesi, GSK and Pfizer. **Rest of authors**: the authors declare that they have no competing interests.

Funding: The *Sociedad Española de Neumología y Cirugía Torácica (SEPAR)* supported this project by a grant to finance the material expenses for the protein array analysis of cytokine, for statistical analysis and English corrector. *The Insitut d'Investigació de Bellvitge (IDIBELL)* supported this project by a grant to finance Samantha Aso PhD graduated student. These institutions haven't participated in the design or development of the study or in the writing of the document.

Author's contributions: S.A., A.N and M.M. takes responsibility of the content and writing of manuscript. S.A., A.N., J.I.M., F.M, and M.M. designed the study. S.A., A.N, S.P, R.P, collected data. N.C and R.L performed the bronchoscopy S.A., L.R., R.C., E.C., and T.G., performed image analysis, S.A., A.M. and M.M., performed cytokine analysis A.N., R.C., S.P, E.C., A.M., J.I.M., N.C., R.L., L.R., R.P, F.M., J.D., and T.G., reviewed manuscript and approved final manuscript.

Acknowledgements: authors wish to thank Bradley Londres for editing the manuscript. Pilar Bayo and Gabriel Reynés (Department of Nuclear Medicine, Bellvitge University Hospital and Physic Department, Catalan Institute of Oncology) for image analysis.

References

1. Ma L, Men Y, Feng L, Kang J, Sun X, Yuan M, Jiang W and Hui Z. A current review of dose-escalated radiotherapy in locally advanced non-small cell lung cancer. *Radiol Oncol.* 2019 Mar 3;53(1):6–14. DOI:[10.2478/raon-2019-0006](https://doi.org/10.2478/raon-2019-0006)
2. Rodemann HP. Molecular radiation biology: Perspectives for radiation oncology. *Radiotherapy and Oncology.* 2009; 92:293–8. DOI:[10.1016/j.radonc.2009.08.023](https://doi.org/10.1016/j.radonc.2009.08.023)
3. Madani I, De Ruyck K, Goeminne H, De Neve W, Thierens H, Van Meerbeeck J. Predicting risk of radiation-induced lung injury. *J Thorac Oncol.* 2007;**2** :864–74. DOI:[10.1097/JTO.0b013e318145b2c6](https://doi.org/10.1097/JTO.0b013e318145b2c6)
4. Inoue A, Kunitoh H, Sekine I, Sumi M, Tokuyue K and Saijo N. Radiation pneumonitis in lung cancer patients: a retrospective study of risk factors and the long-term prognosis. *Int J Radiat Oncol Biol Phys.* 2001; 49:649–55. DOI: [10.1016/s0360-3016\(00\)00783-5](https://doi.org/10.1016/s0360-3016(00)00783-5)

5. Kong FM, Ten Haken R, Eisbruch A, Lawrence TS. Non-small cell lung cancer therapy-related pulmonary toxicity: An update on radiation pneumonitis and fibrosis. *Seminars in Oncology*. 2005; 32:S42-S54. DOI:[10.1053/j.seminoncol.2005.03.009](https://doi.org/10.1053/j.seminoncol.2005.03.009)
6. Wang JY, Chen KY, Wang JT, Chen JH, Lin JW, Wang HC, Lee LN and Yang PC. Outcome and prognostic factors for patients with non-small-cell lung cancer and severe radiation pneumonitis. *Int J Radiat Oncol Biol Phys*. 2002; 54:735–41. DOI: [10.1016/s0360-3016\(02\)02994-2](https://doi.org/10.1016/s0360-3016(02)02994-2)
7. Portillo K, Arriaga I and Ruiz-Manzano J. Fibroelastosis pleuropulmonar: ¿es también una entidad idiopática? *Arch Bronconeumol*. 2015; 51:509-514. DOI: [10.1016/j.arbres.2015.05.002](https://doi.org/10.1016/j.arbres.2015.05.002)
8. Zhang XJ, Sun JG, Sun J, Ming H, Wang XX, Wu L and Chen ZT. Prediction of radiation pneumonitis in lung cancer patients: a systematic review. *J Cancer Res Clin Oncol*. 2012; 138:2103–16. DOI: [10.1007/s00432-012-1284-1](https://doi.org/10.1007/s00432-012-1284-1)
9. Kong F-MS, Wang S. Nondosimetric risk factors for radiation-induced lung toxicity. *Semin Radiat Oncol*. 2015; 25:100–9. DOI: [10.1016/j.semradonc.2014.12.003](https://doi.org/10.1016/j.semradonc.2014.12.003)
10. Barthelemy-Brichant N, Bosquée L, Cataldo D, Corhay J-L, Gustin M, Seidel L, Thiry A, Ghaye B, Nizet M, Albert A, Deneufbourg L-M, Bartsch P and Nusgens B. Increased IL-6 and TGF-beta1 concentrations in bronchoalveolar lavage fluid associated with thoracic radiotherapy. *Int J Radiat Oncol Biol Phys*. 2004; 58:758–67. DOI: [10.1016/S0360-3016\(03\)01614-6](https://doi.org/10.1016/S0360-3016(03)01614-6)
11. Rube CE, Palm J, Erren M, Fleckenstein J, König J, Remberger K and Rube C. Cytokine plasma levels: Reliable predictors for radiation pneumonitis? *PLoS One*. 2008; 3:e2898. DOI: [10.1371/journal.pone.0002898](https://doi.org/10.1371/journal.pone.0002898)
12. Kong FM, Ao X, Wang L, Lawrence TS. The use of blood biomarkers to predict radiation lung toxicity: A potential strategy to individualize thoracic radiation therapy. Vol. 15, *Cancer Control*. 2008. p. 140–50. DOI: [10.1177/107327480801500206](https://doi.org/10.1177/107327480801500206)
13. Guerrero T, Johnson V, Hart J, Pan T, Khan M, Luo D, Liao Z, Ajani J, Stevens C and Komaki R. Radiation Pneumonitis: Local Dose Versus [18F] Fluorodeoxyglucose Uptake Response in Irradiated Lung. *Int J Radiat Oncol Biol Phys*. 2007; 68:1030–5. DOI:[10.1016/j.ijrobp.2007.01.031](https://doi.org/10.1016/j.ijrobp.2007.01.031)
14. Ruyscher D De, Houben A, Aerts HJWL, Dehing C, Wanders R, Öllers M, Dingemans A-M C, Hochstenbag M, Boersma L, Borger J, Dekker A and Lambin P. Increased 18F-deoxyglucose uptake in the lung during the first weeks of radiotherapy is correlated with subsequent Radiation-Induced Lung Toxicity (RILT): A prospective pilot study. *Radiother Oncol*. 2009; 91:415–20. DOI:[10.1016/j.radonc.2009.01.004](https://doi.org/10.1016/j.radonc.2009.01.004)
15. Petit SF, Van Elmpt WJC, Oberije CJG, Vegt E, Dingemans AMC, Lambin P, Dekker AL.A. and De Ruyscher D. [18F]fluorodeoxyglucose uptake patterns in lung before radiotherapy identify areas more susceptible to radiation-induced lung toxicity in non-small-cell lung cancer patients. *Int J Radiat Oncol Biol Phys*. 2011; 81:698–705. DOI:[10.1016/j.ijrobp.2010.06.016](https://doi.org/10.1016/j.ijrobp.2010.06.016)
16. Castillo R, Pham N, Ansari S, Meshkov D, Castillo S, Li M, Olanrewaju A, Hobbs B, Castillo E and Guerrero TM. Pre-radiotherapy FDG PET predicts radiation pneumonitis in lung cancer. *Radiation Oncology*. 2014; 9:74. DOI: [10.1186/1748-717X-9-74](https://doi.org/10.1186/1748-717X-9-74)

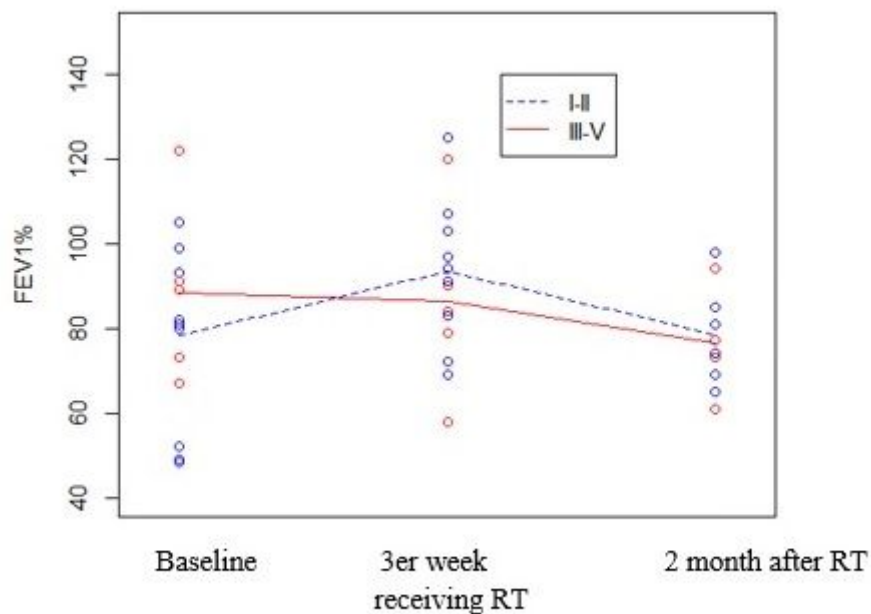
17. Castillo R, Pham N, Castillo E, Aso-Gonzales S, Ansari S, Hobbs B, Palacio D, Skinner H and Guerrero TM. Pre-Radiation Therapy Fluorine Helps Identify Patients with Esophageal Cancer at High Risk for Radiation Pneumonitis. *Radiology*. 2015; 275:822-31. DOI: [10.1148/radiol.14140457](https://doi.org/10.1148/radiol.14140457)
18. National Institute of Cancer. Common Terminology Criteria for Adverse Events (CTCAE), Version 4.0, DCTD, CTI, NIH, DHHS. NIH Publication. 2009. 0-71 p. DOI: [10.1186/s12955-016-0426-6](https://doi.org/10.1186/s12955-016-0426-6)
19. De Ruyscher D, Faivre-Finn C, Nestle U, Hurkmans CW, Le Péchoux C, Price A, and Senan S. European organisation for research and treatment of cancer recommendations for planning and delivery of high-dose, high-precision radiotherapy for lung cancer. *J Clin Oncol*. 2010; 28:5301–10. DOI: [10.1200/JCO.2010.30.3271](https://doi.org/10.1200/JCO.2010.30.3271)
20. Giraud P, Antoine M, Larrouy A, Milleron B, Callard P, De Rycke Y, Carette MF, Rosenwald JC, Cosset JM, Housset M and Touboul E. Evaluation of microscopic tumor extension in non-small-cell lung cancer for three-dimensional conformal radiotherapy planning. *Int J Radiat Oncol Biol Phys*. 2000; 48:1015–24. DOI: [10.1016/s0360-3016\(00\)00750-1](https://doi.org/10.1016/s0360-3016(00)00750-1)
21. Senan S, Chapet O, Lagerwaard FJ, Haken RK Ten. Defining Target Volumes for Non-small Cell Lung Carcinoma. *Semin Radiat Oncol*. 2004;14:308–14. DOI: [10.1016/j.semradonc.2004.07.004](https://doi.org/10.1016/j.semradonc.2004.07.004)
22. Ramnath N, Dilling TJ, Harris LJ, Kim AW, Michaud GC, Balekian AA, Diekemper R, Detterbeck FC and Arenberg DA. Treatment of stage III non-small cell lung cancer: Diagnosis and management of lung cancer, 3rd ed: American college of chest physicians evidence-based clinical practice guidelines. *Chest*. 2013; 143:e314S-340S. DOI: [10.1378/chest.12-2360](https://doi.org/10.1378/chest.12-2360)
23. Kong FM, Ritter T, Quint DJ, Senan S, Gaspar LE, Komaki RU, Hurkmans CW, Timmerman R, Bezjak A, Bradley JD, Movsas B, Marsh L, Okunieff P, Choy H and Curran WJ Jr. et al. Consideration of dose limits for organs at risk of thoracic radiotherapy: Atlas for lung, proximal bronchial tree, esophagus, spinal cord, ribs, and brachial plexus. *Int J Radiat Oncol Biol Phys*. 2011; 81:1442–57. DOI: [10.1016/j.ijrobp.2010.07.1977](https://doi.org/10.1016/j.ijrobp.2010.07.1977)
24. Brusasco V, Crapo R, Viegi G. Coming together: The ATS/ERS consensus on clinical pulmonary function testing. *European Respiratory Journal*. 2005; 26:1–2. DOI: [10.1183/09031936.05.00034205](https://doi.org/10.1183/09031936.05.00034205)
25. Lowry OH, Rosebrough, Farr AL, Randall RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem*. 1951; 193:265–75.
26. Boellaard R, Doherty MJO, Weber WA, Mottaghy FM, Lonsdale MN, Stroobants SG, Oyen WJ, Kotzerke J, Hoekstra OS, Pruim J, Marsden PK, Tatsch K, Hoekstra CJ, Visser EP, Arends B, Verzijlbergen FJ, Zijlstra JM, Comans EF, Lammertsma AA, Paans AM, Willemsen AT, Beyer T, Bockisch A, Schaefer-Prokpo C, Delbeke D, Baum RP, Chiti A and Krause BJ. FDG PET and PET/CT: EANM procedure guidelines for tumour PET imaging: version 1.0. *Eur J Nucl Med Mol Imaging*. 2010; 37:181–200. DOI:[10.1007/s00259-009-1297-4](https://doi.org/10.1007/s00259-009-1297-4)
27. Strauss LG, Conti PS. The Applications of PET in Clinical Oncology. *J Nucl Med*. 1991; 32:623–48.
28. Marrugat J, Vila J, Pavesi m and Sanz F. Estimation of the sample size in clinical and epidemiological investigations. *Med Clin (Barc)*. 1988; 111(7):267-76.

29. Xaubet A, Ancochea J, Morell F, Rodríguez-Arias JM, Villena V, Blanquer R, Montero C, Sueiro A, Disdier C, Vendrell M, Spanish Group on Interstitial Lung Disease, SEPAR. Report on the Incidence of Interstitial Lung Diseases in Spain. *Sarcoidosis Vasc Diffuse Lung Dis.* 2004;21(1):64-70.
30. Ley JA, Borbón GA, Ochoa Carrillo FJ, Escobar RV, Rojas SH, Estrada G. Valor estandarizado de captación máximo, determinado con Tomografía Computarizada. "Primera experiencia en México" (Spanish). *An Radiol Mex.* 2007; 6:113–9.
31. Lopez Guerra JL, Gomez DR, Zhuang Y, Levy LB, Eapen G, Liu H, Mohan R, Komaki R, Cox JD and Liao Z. Changes in pulmonary function after three-dimensional conformal radiotherapy, intensity-modulated radiotherapy, or proton beam therapy for non-small-cell lung cancer. *Int J Radiat Oncol Biol Phys.* 2012; 83:E537-43. DOI: [10.1016/j.ijrobp.2012.01.019](https://doi.org/10.1016/j.ijrobp.2012.01.019)
32. Palma DA, Senan S, Tsujino K, Barriger RB, Rengan R, Moreno M, Bradley JD, Kim TH, Ramella S, Marks LB, De Petris L, Stitt Larry and Rodrigues G. Predicting radiation pneumonitis after chemoradiation therapy for lung cancer: an international individual patient data meta-analysis. *Int J Radiat Oncol Biol Phys.* 2013; 85(2): 444–50. DOI:[10.1016/j.ijrobp.2012.04.043](https://doi.org/10.1016/j.ijrobp.2012.04.043)
33. Keane MP. The role of chemokines and cytokines in lung fibrosis. Vol. 17, *European Respiratory Review.* 2008. p. 151–6.
34. Tang W, Yang L, Qin W, Yi MX, Liu B, Yuan XL. Impact of genetic variant of HIPK2 on the risk of severe radiation pneumonitis in lung cancer patients treated with radiation therapy. *Radiat Oncol.* 2020; 15(1):9. DOI:[10.1186/s13014-019-1456-0](https://doi.org/10.1186/s13014-019-1456-0)
35. Gasse P, Mary C, Guenon I, Noulin N, Charron S, Schnyder-Candrian S, Scsnyder B, Akira S, Quesniaux V.F.J, Lagente V, Ryffel B and Couillin I. IL-1R1/MyD88 signaling and the inflammasome are essential in pulmonary inflammation and fibrosis in mice. *J Clin Invest.* 2007; 117:3786–99. DOI: [10.1172/JCI32285](https://doi.org/10.1172/JCI32285)
36. Mathew B, Jacobson JR, Siegler JH, Moitra J, Blasco M, Xie L, Unzueta C, Zhou T, Evenoski C, Al-Sakka M, Sharma R, Huey B, Bulent A, Smith B, Jayaraman S, Reddy NM, Reddy SP, Fingerle-Rowson G, Bucala R, Dudek SM, Natarajan V, Weichselbaum RR and Garcia J.G.N. Role of migratory inhibition factor in age-related susceptibility to radiation lung injury via NF-E2-related factor-2 and antioxidant regulation. *Am J Respir Cell Mol Biol.* 2013; 49:269–78. DOI: [10.1165/rcmb.2012-02910C](https://doi.org/10.1165/rcmb.2012-02910C)
37. Gasse P, Riteau N, Vacher R, Michel ML, Fautrel A, di Padova F, Fick L, Charron S, Lagente V, Eberl G, Le Bert M, Quesniaux VF, Huaux F, Leite-de-Moraes M, Ryffel B and Couillin I. IL-1 and IL-23 mediate early IL-17A production in pulmonary inflammation leading to late fibrosis. *PLoS One.* 2011; 6:e23185. DOI: [10.1371/journal.pone.0023185](https://doi.org/10.1371/journal.pone.0023185)
38. Kaufman J, Sime PJ, Phipps RP. Expression of CD154 (CD40 ligand) by human lung fibroblasts: differential regulation by IFN-gamma and IL-13, and implications for fibrosis. *J Immunol.* 2004; 172:1862–71. DOI: [10.4049/jimmunol.172.3.1862](https://doi.org/10.4049/jimmunol.172.3.1862)
39. Senoo T, Hattori N, Tanimoto T, Furonaka M, Ishikawa N, Fujitaka K, Haruta Y, Murai H, Yokoyama A and Kohno N. Suppression of plasminogen activator inhibitor-1 by RNA interference attenuates pulmonary fibrosis. *Thorax.* 2010; 65:334–40. DOI: [10.1136/thx.2009.119974](https://doi.org/10.1136/thx.2009.119974)

40. Liu B, Tang Y, Yi M, Liu Q, Xiong H, Hu G and Yuan X. Genetic variants in the plasminogen activator inhibitor-1 gene are associated with an increased risk of radiation pneumonitis in lung cancer patients. *Cancer Med.* 2017; 6:681–8. DOI: [10.1002/cam4.1011](https://doi.org/10.1002/cam4.1011)

Figures

A. - Evolution of FEV1



B. - Evolution of DLCO values

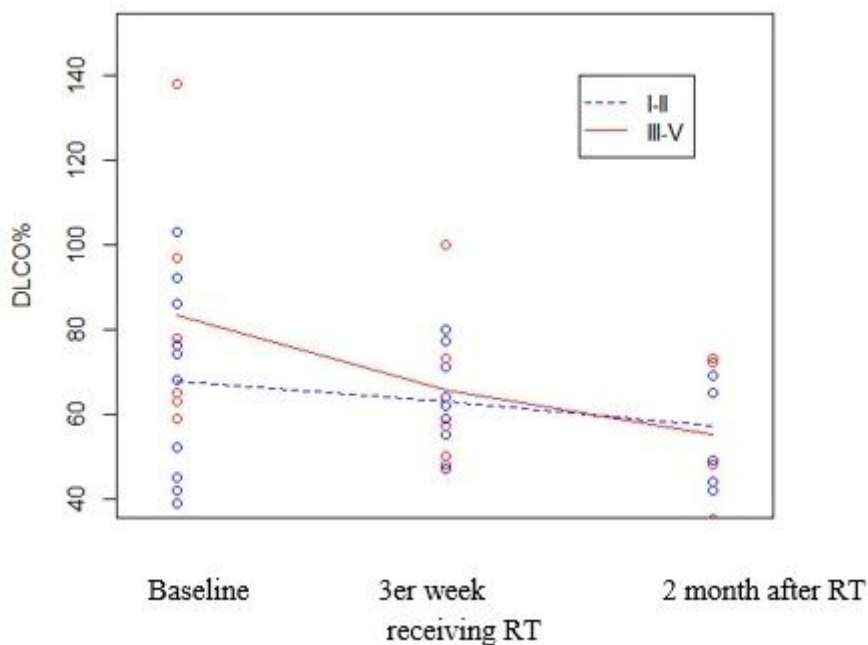
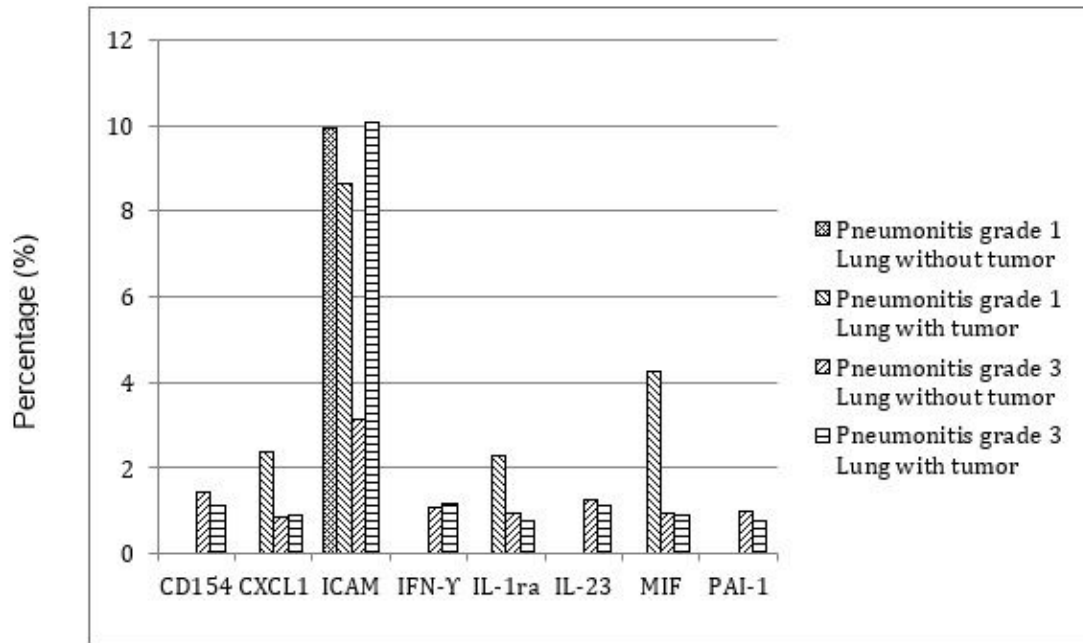


Figure 1

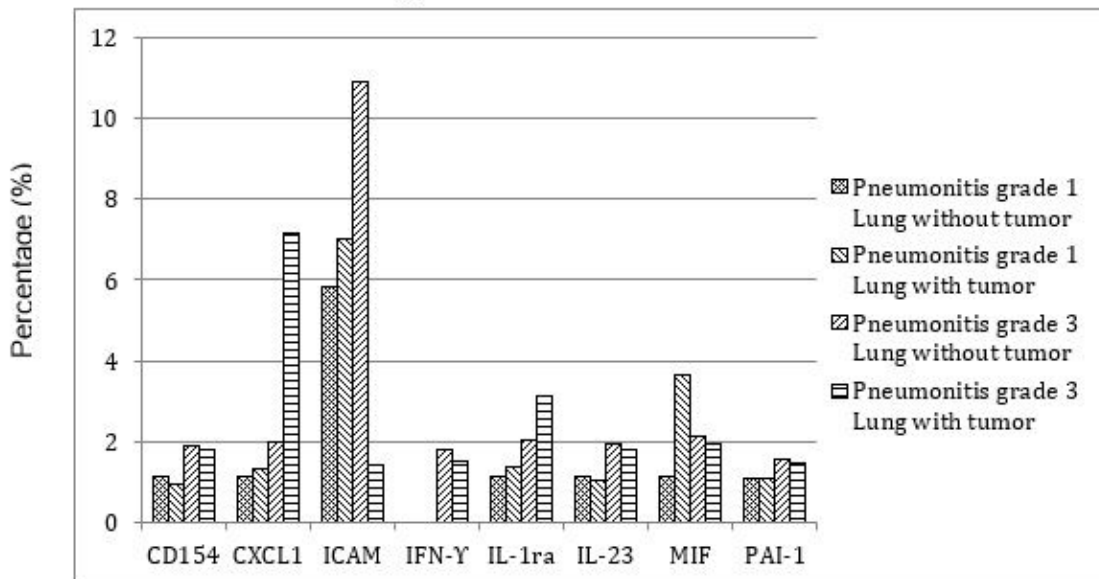
Pulmonary function test values at different time-points by CTCAEv4.0 groups (low-grade [GI and GII] and high-grade [GIII-GV]). A. - Evolution of FEV1: The points represent the FEV1 (%) value of each patient at the three different observation times. The straight, is the mean FEV1 (%) value at each observation time. CTCAEv4.0: Common Terminology Criteria for Adverse Events version 4.0 FEV1: forced expiratory volume in one second RT: radiotherapy. B. - Evolution of DLCO values: The points represent the DLCO (%) value of each patient at three different observation times. The straight line is the mean DLCO (%) value at each observation time. CTCAEv4.0: Common Terminology Criteria for Adverse Events version 4.0 DLCO: diffusing lung capacity for carbon monoxide RT: radiotherapy

A. - Before radiotherapy



Chemokines and cytokine

B. - Third week with radiotherapy



Chemokines and cytokine

Figure 2

Cytokine and chemokines in bronchoalveolar lavage in patients with grade 1 and grade 3 radiation pneumonitis. A. - Before radiotherapy. B. - Third week with radiotherapy