

Identification of Candidate Genes Controlling Fiber Quality Traits in Upland Cotton through Integration of Meta-QTL, Significant SNP and Transcriptomic Data

XU Shudi

Shihezi University College of Agriculture

Zhenyuan Pan

Shihezi University College of Agriculture

Feifan Yin

Huazhong Agriculture University

Qingyong Yang

Huazhong Agriculture University

Zhongxu Lin

Huazhong Agriculture University

Tianwang Wen

Huazhong Agriculture University

Longfu Zhu

Huazhong Agriculture University

Dawei Zhang

Xinjiang Academy of Agricultural Sciences

Xinhui Nie (✉ xjnxh2004130@126.com)

Shihezi University College of Agriculture

Research

Keywords: Fiber quality traits, Meta-QTL, Significant SNPs, Candidate genes, Transcriptomic data

DOI: <https://doi.org/10.21203/rs.3.rs-42429/v2>

License:   This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Abstract

Background: Meta-analysis of quantitative trait locus (QTL) is a computational technique to identify consensus QTL and refine QTL positions on the consensus map from multiple mapping studies. The combination of meta-QTL intervals, significant SNPs and transcriptome analysis has been widely used to identify candidate genes in various plants.

Results: In our study, 884 QTLs associated with cotton fiber quality traits from 12 studies were used for meta-QTL analysis based on reference genome TM-1, as a result, 74 meta-QTLs were identified, including 19 meta-QTLs for fiber length (FL); 18 meta-QTLs for fiber strength (FS); 11 meta-QTLs for fiber uniformity (FU); 11 meta-QTLs for fiber elongation (FE); and 15 meta-QTLs for micronaire (MIC). Combined with 8,589 significant SNPs associated with fiber quality traits collected from 15 studies, 297 candidate genes were identified in the meta-QTL intervals, 20 of which showed high expression specifically in the developing fibers. According to the function annotations, some of the 20 key candidate genes are associated with the fiber development.

Conclusions: This study provides not only stable QTLs used for marker-assisted selection (MAS), but also candidate genes to uncover the molecular mechanisms for cotton fiber development.

Background

As a natural and renewable resource, cotton fiber has been the most important raw material in the cotton textile and processing industry all over the world. With the improvement of people's living standard and advancements in techniques and diversified methods of spinning, demand for high quality cotton fiber is increasing. Cotton fibers are derived from ovule epidermal cells -2 to ~0 d post anthesis (DPA), and ultimately reach 2.5–3.5 cm in the mature period (Stewart 1975), which consists of four stages: fiber initiation; cell elongation; secondary cell wall (SCW) biosynthesis; and maturation (Li et al. 2018). The development mechanism of cotton fiber contributed to the fiber quality improvement. Cotton fiber quality traits are complex quantitative traits, which are influenced by environments and the fiber development, and controlled by many QTLs, including fiber length (FL), fiber strength (FS), fiber uniformity (FU), fiber elongation (FE), and micronaire (MIC), etc (Ademe et al. 2017; Wang et al. 2016). FL and FS are considered as the most important traits affecting yarn quality, and FS is important for advanced spinning technologies in the textile industry (Yang et al. 2016). The MIC is a measure of fiber fineness and fiber maturity, which influences the fiber processing and dyeing consistency (Rodgers et al. 2017). Hundreds of QTLs contributing to fiber quality traits have been previously mapped for cotton using a variety of populations, which were evenly distributed throughout the cotton genome (Qin et al. 2008; Shen *et al.* 2007; Wang et al. 2020; Yu et al. 2013). A total of 104 QTLs for fiber quality traits were detected by using 180 recombinant inbred lines derived from Herein and Yumian 1, and 25 QTLs were detected in all three environments (Tan et al. 2018). A total of 134 QTLs for fiber quality traits were detected using 231 F_{6:8} recombinant inbred lines, which were derived from an intraspecific cross between Xinluzao24 and Lumianyan28 (Liu et al. 2018b). Seventy-four QTLs were detected to be associated with five fiber quality

traits (30 QTLs) and eight yield traits (44 QTLs) using 107 introgression lines, which were developed with an interspecific cross using *G. hirsutum* acc. 4105 as the recurrent parent and *G. tomentosum* as the donor parent (Keerio et al. 2018). One hundred and eighty-six additive QTLs were obtained for five fiber quality traits using 137 recombinant inbred lines (Jia et al. 2018).

MAS has been successfully applied in genetic improvement of varieties of crops, especially for the major QTLs/genes, such as improvement of rice blast resistance by pyramiding three genes in rice (Xiao et al. 2019), improvement of drought adaptation in maize (Ribaut and Ragot, 2006), improvement of yield traits in soybean (Reyna and Sneller 2001; Sebastian et al. 2010), improvement of Fusarium head blight resistance in wheat (Anderson 2007), and improvement of Verticillium Wilt resistance in cotton (Zhang et al. 2014). QTL mapping with molecular markers provides a powerful approach to dissect the molecular mechanism underlying complex fiber quality traits (Ijaz et al. 2019). There were thousands of QTLs for cotton fiber quality traits identified in different mapping populations such as RILs, bi-parental segregating populations, and BC populations (Said et al. 2015b), which provide the potential to be manipulated by MAS for the improvement of cotton fiber quality traits. However, only the stable QTLs for cotton fiber quality traits in various environments and populations can be used in the MAS breeding. In order to make these mapped QTLs more useful to plant breeding and gene cloning, a further analysis of all these loci has to be carried out. In this regard, meta-analysis of QTLs has been proven as an efficient approach to establish the occurrence of QTL “hotspots” in a consensus map, which correspond to the more precise region where these loci represent under analysis (Goffinet and Gerber 2000; Salvi and Tuberosa 2015). More than three overlapped or location similar QTLs reported in multiple documents of the same trait is considered as a meta-QTL. This approach was already applied to various crops and complex traits, such as fusarium head blight resistance in bread wheat (Venske et al. 2019), grain weight in tetraploid wheat (Avni et al. 2018), cyst nematode resistance in soybean (Guo et al. 2006a), resistance to white mold in common bean (Vasconcellos et al. 2017), yield under drought in rice (Swamy et al. 2011), yield in maize (Martinez et al. 2016), and multiple traits in cotton (Said et al. 2013).

Several strategies were combined to identify candidate genes, such as combination of association mapping and linkage analysis (Cui et al. 2018; Mahuku et al. 2016; Zhang et al. 2019a) and combination of QTLs and transcriptome analysis (Chen and Bian 2018; Shimono et al. 2016; Wang et al. 2020). A sucrose synthesis-related gene (*Gh_D03G1338*) associated with FL was identified by the combination of genome-wide association and linkage analyses (Zhang et al. 2019a). Three genes, *Gh_D05G1077* and *Gh_D13G1571* for SY, and *Gh_A11G0775* for LY, were identified using genome-wide association mapping. Five candidate genes were identified by the combination of QTL mapping and transcriptome analysis, which regulated pericarp thickness in sweet corn (Wu et al. 2020). A peroxidase gene (*GhPRXR1*) required for oil content in upland cotton was identified by the combination of genome-wide association and transcriptome analysis (Ma et al. 2019). Two candidate genes for fiber elongation and developmental were identified by the combination of genome-wide association and transcriptome analysis (Ma et al. 2018a).

Some novel genes functioning in fiber initiation and elongation have been verified by molecular biology methods, for example, an R2R3 MYB transcription factor gene (*GhMYB25-like*)(Walford et al. 2011), a homeodomain leucine zipper gene (*GhHD-1*)(Walford et al. 2012), a vacuolar invertase gene (*GhVIN1*) (Wang et al. 2014), a cotton actin gene (*GhACT1*)(Li et al. 2005), cotton annexin genes (*AnxGb6* and *GhAnn2*)(Huang et al. 2013; Tang et al. 2014), an fiber-specific profiling gene (*GhPFN2*)(Wang et al. 2010), and an actin-depolymerizing factor gene (*GhADF1*) (Wang et al. 2009). In addition, cellulose synthases genes *GhCesA1*, *WLIM1a* and *GhADF1* are responsible for the secondary cell wall (SCW) in cotton fibers (Han et al. 2013; Salnikov et al. 2003; Wang et al. 2009).

In this study, 884 QTLs associated with cotton fiber traits from 12 studies (Ali et al. 2018; Diouf et al. 2018; Huang et al. 2017; Jia et al. 2018; Keerio et al. 2018; Li et al. 2016a; Liu et al. 2018b; Ma et al. 2018a; Tan et al. 2018; Wang et al. 2015; Zhang et al. 2015b; Zou et al. 2018) were used for meta-QTL analysis based on upland cotton reference genome TM-1 (Zhang et al. 2015a), and 74 meta-QTLs were identified. Combined with 8589 significant SNP loci associated to cotton fiber quality traits collected from 15 previous publications (Chandnani et al. 2018; Fang et al. 2017; Gapare et al. 2017; Handi et al. 2017; Huang et al. 2017; Islam et al. 2016; Li et al. 2018; Li et al. 2017b; Liu et al. 2018b; Ma et al. 2018a; Ma et al. 2018b; Su et al. 2016; Su et al. 2018; Sun et al. 2017; Wen et al. 2018), 297 candidate genes associated with cotton fiber quality traits were identified. Twenty genes showed high expression specifically in the developing fibers, some of which are associated with the fiber development. According to the results, the combination of meta-QTL, significant SNP by GWAS, and spatiotemporal expression analysis provides not only stable QTLs used for marker-assisted selection (MAS), but also candidate genes to uncover the molecular mechanisms for cotton fiber development.

Methods

Data collection and organization

From the Web of Science website, *G. hirsutum*, fiber quality traits, GWAS, SNP, QTL, and high density genetic map (HDGM) were searched as keywords, and more than 50 related articles were retrieved. The articles providing QTL intervals and flanking markers were selected for QTL collection. Finally, 884 QTLs with respect to FE, FL, FS, MIC, FU, spinning consistency index (SCI), short fiber (SF), fiber reflectance (FR) and fiber yellowness (FY) traits were identified from the web of science (Ali et al. 2018; Diouf et al. 2018; Huang et al. 2017; Jia et al. 2018; Keerio et al. 2018; Li et al. 2016a; Liu et al. 2018b; Ma et al. 2018a; Tan et al. 2018; Wang et al. 2015; Zhang et al. 2015b; Zou et al. 2018). QTL numbers, traits, population type and size, and number of markers are listed in Table 1. GWAS data, including 8589 SNPs significant loci associated with FL, FS, FU, FE, MIC, MIC, SCI, SF, and FC are listed on Table S3, Table S4 (Chandnani et al. 2018; Fang et al. 2017; Gapare et al. 2017; Handi et al. 2017; Huang et al. 2017; Islam et al. 2016; Li et al. 2018; Li et al. 2017b; Liu et al. 2018b; Ma et al. 2018a; Ma et al. 2018b; Su et al. 2016; Su et al. 2018; Sun et al. 2017; Wen et al. 2018).

Meta-QTL analysis

Since SNPs are developed by genome sequencing, each marker has a fixed and unique location in the genome. By anchoring the SNPs on both sides of the QTLs to the TM-1 genome (Zhang et al. 2015a), the confidence interval of the QTLs can be determined. A stable meta-QTL region was obtained by manual organizing, and the stable meta-QTL intervals were illustrated in the form of Circos plot using Circos software (Krzywinski et al. 2009). The SNP loci significantly correlated with the same trait was compared with the meta-QTL intervals; thereby the most likely location of the candidate genes in the meta-QTL intervals was determined.

Candidate gene identification

8589 significant SNP loci associated to cotton fiber quality traits were collected from 15 GWAS studies and mapped to TM-1 genome (Chandnani et al. 2018; Fang et al. 2017; Gapare et al. 2017; Handi et al. 2017; Huang et al. 2017; Islam et al. 2016; Li et al. 2018; Li et al. 2017b; Liu et al. 2018b; Ma et al. 2018a; Ma et al. 2018b; Su et al. 2016; Su et al. 2018; Sun et al. 2017; Wen et al. 2018) (Table S3, Table S4). Then they were mapped to the 74 meta-QTLs, and the mapped SNPs are shown in Table S5. Lastly, the genes closely linked to the SNPs (SNPs within genes) were selected as candidate genes, which are shown in Table S6.

Gene ontology (GO) analysis and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis

The significant enrichment analysis of gene ontology (GO) terms was carried out using agriGO v2.0 software (p-value < 0.05) (<http://bioinfo.cau.edu.cn/agriGO/>) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis of Genes was performed with KEGG Automatic Annotation Server (KAAS). R package clusterProfiler (<http://bioconductor.org/packages/2.8/bioc/html/clusterProfiler.html>) was used for result visualization.

Gene expression patterns

The tissue expression levels of the candidate genes were obtained from previously reported transcriptome data (Zhang et al. 2015a).

Results

Collection of QTLs and SNPs associated with fiber quality traits

A total of 12 QTL mapping studies for cotton fiber quality traits were used in this study, in which the mapping population size ranged from 107 to 503 lines (Table 1) (Ali et al. 2018; Diouf et al. 2018; Huang et al. 2017; Jia et al. 2018; Keerio et al. 2018; Li et al. 2016a; Liu et al. 2018b; Ma et al. 2018a; Tan et al. 2018; Wang et al. 2015; Zhang et al. 2015b; Zou et al. 2018), and the number of SNP markers ranged from 168 to 19,191 (Table 1). As a result, a total of 884 initial QTLs related to cotton fiber quality traits were collected, which were unevenly distributed on each chromosome, and ranged from 12 to 57 (Fig. 1, Table S1). Chromosome A4 had the lowest number of QTLs and chromosome A10 had the highest number of QTLs. Among them, there were a large number of QTLs related to FL, FS, MIC, FU, and FE,

which were 204, 207, 179, 118, and 108, respectively. However, there were small number of QTLs related to the SCI, SF, FR, and FY, which were 21, 19, 13 and 15, respectively (Table S1).

Table 1 Fiber quality traits QTLs mapped by SNP markers from 12 papers

QTL	Traits	Population type	Population size	Number of markers	Reference
8	FS	RIL	250	168 SNP	[Zou et al. 2018]
37	FL, FS, MIC	RIL	196	106 SSR & 104 SNP	[Zhang et al. 2015]
9	FS	RIL	161	304 SSR & 5571 SNP	[Wang et al. 2015]
104	FL, FS, MIC, FU, FE	RIL	180	12116 SNP	[Tan et al. 2018]
21	FL, FE	BIL	176	15369 SNP	[Ma et al. 2018]
134	FL, FS, MIC	RIL	231	122 SSR& 4729 SNP	[Liu et al. 2018]
30	FL, FS, MIC, FU, FE	ILs	107	3157 SNP	[Keerio et al. 2018]
186	FL, FS, MIC, FU, FE	RIL	137	139 SSR & 6295 SNP	[Jia et al. 2018]
50	FL, FS, MIC, FU, FE, SF	Natural population	503	19 191 SNP	[Huang et al. 2017]
193	FL, FU, MIC, FS, FE, FR, FY, SCI	F _{2:3} population	277	5178 SNP	[Diouf et al. 2018]
48	FL, FS, MIC, FU, FE	RIL	188	2618 SNP	[Li et al. 2016]
59	FL, FS, MIC, FU, FE	RIL	180	6254 SNP	[Ali et al. 2018]

FL, fiber length; FS, fiber strength; MIC, micronaire; FU, fiber uniformity; FE, fiber elongation; SCI, spinning consistency index; SF, short fiber; FR, fiber reflectance; FY, fiber yellowness.

Meta-analysis of QTL for fiber quality traits

A meta-analysis was performed with 884 QTLs related to cotton fiber quality traits, and a total of 74 stable meta-QTLs related to FL, FS, FE, MIC, and FU were obtained, including 19 for FL, 18 for FS, 11 for

FU, 11 for FE, and 15 for MIC, which covered 26 upland cotton chromosomes. There were 33 meta-QTLs in the A sub-genome and 41 in the D sub-genome. The confident intervals (CI) of all meta-QTLs were smaller than their respective initial QTLs, which were ranged from 2.4 to 13.4 Mb, with an average of 8.5 Mb (Fig. 2, Table S2). Among the 74 meta-QTLs, 19 were obtained from multiple QTLs coincident regions of 4 or more studies (Table S2), indicating that these regions had high correlation with cotton fiber quality traits.

Meta-QTLs for FL

A total of 19 meta-QTLs related to FL were obtained, covering 17 chromosomes, including 7 meta-QTLs on the A sub-genome (A01, A05, A08, A09, A10, and A12), and 12 Meta-QTLs on the D sub-genome (D02, D03, D04, D05, D06, D08, D09, D10, D11, D12, and D13) (Fig. 2, Table S2). Three meta-QTLs have been mapped in more than 5 studies, namely meta-QTL-3, meta-QTL-15, and meta-QTL-17, which were located in the 18.8-31.70 Mb region on A05 chromosome, the 54.17-61.86 Mb region on D10 chromosome, and the 15.70-24.32 Mb region on D11 chromosome, respectively (Table S2).

Meta-QTL for FS

A total of 18 meta-QTLs related to FS were obtained, covering 15 chromosomes, including 9 meta-QTLs on the A sub-genome (A01, A02, A03, A05, A07, A09, and A10), and 9 meta-QTLs on the D sub-genome (D04, D05, D06, D07, D08, D10, D11, and D12) (Fig. 2, Table S2). Among them, four meta-QTLs have been identified in the four studies, namely meta-QTL-23, meta-QTL-25, meta-QTL-27, and meta-QTL-30, which were located in 5.81-14.90 Mb on A05, 63.33-73.57 Mb on A07, 93.51-100.20 Mb on A10, and 4.01-15.81 Mb region on D05, respectively (Table S2).

Meta-QTL for FU

A total of 11 meta-QTLs related to FU were obtained, covering 11 chromosomes, including 5 meta-QTLs on the A sub-genome (A02, A06, A09, A10, and A11), and 6 meta-QTLs on the D sub-genome (D01, D02, D03, D05, D11, and D12). Among the meta-QTLs, meta-QTL-45 on D03 and meta-QTL-47 on D11 chromosomes are more reliable for identification in four studies (Fig. 2, Table S2).

Meta-QTLs for FE

A total of 11 meta-QTLs related to FE were obtained, covering 11 chromosomes, including 4 meta-QTLs on the A sub-genome (A05, A10, A11, and A13), and 7 meta-QTLs on the D sub-genome (D01, D04, D07, D11, and D12) (Fig. 2, Table S2). Among these meta-QTLs, meta-QTL-53 on chromosome D01 was identified in four studies.

Meta-QTL for MIC

A total of 15 meta-QTLs related to MIC were obtained, covering 14 chromosomes, including 8 meta-QTLs on the A sub-genome (A05, A06, A08, A09, A10, A11, and A13), and 7 meta-QTLs on the D sub-genome

(D03, D05, D06, D08, D09, D11, and D12) (Fig. 2, Table S2). Two meta-QTLs have been identified in the four studies, namely meta-QTL-61 and meta-QTL-71, which were located in 17.32-26.9 Mb on A05, and 52.98-62.38 Mb on D08, respectively (Table S2).

Candidate genes identification combined and meta-QTL intervals and significant SNPs

8,589 significant SNP loci associated to cotton fiber quality traits were collected from 15 GWAS studies and mapped to the TM-1 genome (Chandnani et al. 2018; Fang *et al.* 2017; Gapare et al. 2017; Handi et al. 2017; Huang et al. 2017; Islam et al. 2016; Li et al. 2018; Li et al. 2017b; Liu et al. 2018b; Ma et al. 2018a; Ma et al. 2018b; Su et al. 2016; Su et al. 2018; Sun et al. 2017; Wen et al. 2018) (Table S3, Table S4, Fig. 2), 4343 of which were mapped in the 74 meta-QTL regions (Table S5). Two hundred and ninety-seven candidate genes were identified closely linked to the 4343 SNPs, including 126 genes for FL, 93 for FS, 40 for FU, 20 for FE, 18 for MIC (Table S6).

GO and KEGG enrichment analysis of candidate genes

To identify common characteristics of these genes in biological functions, gene ontology (GO) analysis was performed with the 297 candidate genes, and 200 of them had ontology annotations, which were classified into the three main GO categories (biological process, molecular function, and cellular component) and 15 GO terms (Fig. 3; Table S7). In the biological process category, protein modification (30, 15%), cellular protein modification (30, 15%), protein metabolism (46, 23%), macromolecule modification (30, 15%), macromolecule metabolism (69, 34.5%), cellular protein metabolism (37, 18.5%), cellular macromolecule metabolism (60, 30%), and proteolysis (11, 5.5%) were the major subcategories (Fig. 3, Table S7). In the cellular component category, 35 (17.5%) genes were enriched in the membrane subcategory (Fig. 3; Table S7). In the molecular function category, protein serine/threonine kinase activity (18, 9%), protein binding (55, 27.5%), peptidase activity (10, 5%), protein tyrosine kinase activity (20, 10%), DNA binding (22, 11%), and transporter activity (16, 8%) were the major subcategories (Fig. 3, Table S7).

To further comprehend the enriched pathways of the candidate genes, KEGG pathway analysis was performed, and 234 annotated genes were assigned to 4 KEGG pathways ($P < 0.05$), including pentose and glucuronate interconversions, acarbose and validamycin biosynthesis, vitamin digestion and absorption, and membrane trafficking (Table 2). Some of the pathways have been reported to be associated with fiber development, such as the pentose and glucuronate interconversions pathway is associated with fiber elongation.

Table 2 KEGG analysis of candidate genes

Accession	Name	Input gene number	<i>p</i> -value
ko00040	Pentose and glucuronate interconversions	2 (1.89%)	0.01
ko00525	Acarbose and validamycin biosynthesis	1 (0.94%)	0.02
ko04977	Vitamin digestion and absorption	1 (0.94%)	0.02
ko04131	Membrane trafficking	10 (9.43%)	0.04

Key candidate genes identified from expression patterns

To better understand the molecular function of the candidate genes, the expression in 10 tissues (root, stem, leaf, petal, anther, stigma, fibers at four developmental stages) obtained from the transcriptome datasets of the upland cotton genetic standard TM-1 were used for spatiotemporal expression analysis (Zhang *et al.* 2015a). Among the 126 candidate genes for FL, nine of which (*Gh_D08G1950*, *Gh_D06G0479*, *Gh_D11G1626*, *Gh_D13G1900*, *Gh_D10G0833*, *Gh_D13G1965*, *Gh_A09G1231*, *Gh_D08G1970*, and *Gh_D04G1574*) showed high expression specifically during the development of cotton fiber. *Gh_D08G1950*, *Gh_D06G0479*, *Gh_D11G1626*, *Gh_D13G1900*, *Gh_D10G0833*, and *Gh_D13G1965* showed high expression specifically at fiber secondary wall biosynthesis stages (20 and 25 DPA); *Gh_A09G1231*, *Gh_D08G1970*, and *Gh_D04G1574* showed high expression specifically at fiber elongation stages (5 and 10 DPA). Among the 93 candidate genes for FS, 5 of which (*Gh_A01G1474*, *Gh_A05G2203*, *Gh_D08G2110*, *Gh_A10G2036*, and *Gh_A07G1801*) showed high expression specifically during the development of cotton fiber. *Gh_A01G1474*, *Gh_A05G2203* and *Gh_D08G2110* showed specifically high expression at fiber secondary wall biosynthesis stages (20 and 25 DPA); *Gh_A10G2036*, and *Gh_A07G1801* showed high expression specifically at fiber elongation stages (5 and 10 DPA). Among the 40 candidate genes for FU, 2 of which (*Gh_A11G2663* and *Gh_D11G2059*) showed high expression specifically during the development of cotton fiber. *Gh_A11G2663* showed high expression specifically at fiber secondary wall biosynthesis stages (20 and 25 DPA); *Gh_D11G2059* showed high expression specifically at fiber elongation stages (5 and 10 DPA). Among the 20 candidate genes for FE, 3 of which (*Gh_A13G0282*, *Gh_A13G0354* and *Gh_A11G1313*) showed high expression specifically at fiber elongation stages (5 and 10 DPA). Among the 20 candidate genes for FE, *Gh_D11G1416* showed high expression specifically at fiber secondary wall biosynthesis stages (20 and 25 DPA) (Table S8).

To better understand the molecular function of the key candidate genes identified from expression patterns, they were annotated in CottonFGD (<https://cottonfgd.org/>). As a result, 4 genes (*Gh_D10G0833*, *Gh_A05G2203*, *Gh_D11G2059*, and *Gh_A13G0354*) have no protein annotation, and others encode a variety of proteins (Table 3). The annotations of the key candidate genes were listed in Table 3.

Table 3 The annotation of key candidate genes

Traits	Gene ID	Annotation
FL	<i>Gh_D08G1950</i>	Probable copper-transporting ATPase HMA5
FL	<i>Gh_D06G0479</i>	Basic endochitinase
FL	<i>Gh_D11G1626</i>	COBRA-like protein 4
FL	<i>Gh_D13G1900</i>	WAT1-related protein At1g09380
FL	<i>Gh_D10G0833</i>	Not available
FL	<i>Gh_D13G1965</i>	Protein WVD2-like 1
FL	<i>Gh_A09G1231</i>	40S ribosomal protein S28
FL	<i>Gh_D08G1970</i>	Probable aquaporin PIP1-2
FL	<i>Gh_D04G1574</i>	Snakin-1
FS	<i>Gh_A01G1474</i>	WAT1-related protein At1g25270
FS	<i>Gh_A05G2203</i>	Not available
FS	<i>Gh_D08G2110</i>	CASP-like protein 5A2
FS	<i>Gh_A10G2036</i>	Rop guanine nucleotide exchange factor 5
FS	<i>Gh_A07G1801</i>	Peptidyl-prolyl cis-trans isomerase FKBP15-1
FU	<i>Gh_A11G2663</i>	Protein WVD2-like 1
FU	<i>Gh_D11G2059</i>	Not available
FE	<i>Gh_A13G0282</i>	Xylulose kinase
FE	<i>Gh_A13G0354</i>	Not available
FE	<i>Gh_A11G1313</i>	EPIDERMAL PATTERNING FACTOR-like protein 9
MIC	<i>Gh_D11G1416</i>	Transcriptional corepressor LEUNIG_HOMOLOG

Discussion

SNP-based meta-QTL analysis for cotton fiber quality traits supports complete and accurate genetic information

In previous studies, genetic maps were constructed with SSR markers and other low throughput molecular markers in cotton genetic studies (Guo et al. 2013; Nie et al. 2016; Sun et al. 2012). The low molecular markers density in genetic research could not only reduce the number and accuracy of QTL identification, but also result in large confidence intervals of the QTL in the genome (Guo et al. 2006b; Liu

et al. 2012; Su et al. 2016). With the development of sequencing technology, the application of SNP markers are used in cotton genetic research, such as high-density genetic maps (HDGM) construction (Ali et al. 2018; Diouf et al. 2018) and genome-wide association (GWAS) study (Cai et al. 2017; Li et al. 2017a; Sun et al. 2018), which results in the identification of a large number of QTL for cotton fiber quality traits. Though the meta-analysis of QTL for cotton fiber quality traits were already reported, they were based on the low throughput molecular markers (Said et al. 2015a; Said et al. 2013), which could result in the loss of genetic information, as well as the reduction of accuracy of meta-QTLs. Previous studies have indicated that a region containing multiple QTLs of same traits can be used as a hotspot with a region size of approximately 20 cM (Said et al. 2013) or a physical distance of approximately 10 Mb in upland cotton (Keerio et al. 2018). Therefore, in order to ensure the credibility and inclusiveness of the meta-QTLs, the genome region of about 10 Mb was used as the confidence interval of the meta-QTLs. In this study, 884 QTLs for cotton fiber quality traits were collected from high-density genetic maps, which were constructed with large numbers of SNP markers. Seventy-four meta-QTLs were identified. The CI of all meta-QTLs was smaller than their respective initial QTLs, which is ranged from 2.4 to 13.4 Mb, with an average of 8.5 Mb. In addition, the application of unique SNP markers makes it easier to map the QTLs from different studies to TM-1 genome of upland cotton.

The meta-QTLs contribute to cotton fiber quality improvement by MAS

MAS needs to be enabled through the identification of robust QTLs, the design of reliable marker systems to select for these QTLs, and the delivery of these QTLs into elite genomic backgrounds to enable their use without associated genetic drag (Cobb et al. 2019). In the study, though 884 QTLs for cotton fiber quality traits were collected for meta-analysis, only 74 meta-QTLs were identified, among which, 19 were obtained from multiple QTL coincident regions of 4 or more studies, including 11 for FL, 4 for FS, 1 for FU, 2 for FE, and 2 for MIC. So the flanking SNP markers of the meta-QTLs identified in the study, especially the 19 meta-QTLs, can be used for MAS to improve the cotton fiber quality.

The combination of meta-QTL intervals and significant SNP provide reliable information to identify candidate genes

Due to the challenging detection of rare variants in GWAS and high false-positive rates in QTL mapping, the combination of association mapping and linkage analysis has been widely used for revealing the genetic architecture of complex quantitative traits (Andersen et al. 2005; Li et al. 2016b; Visscher 2008). Seventeen candidate genes for kernel test weight were identified in maize by the combination of association mapping and linkage analysis (Zhang et al. 2019c). Nineteen candidate genes for plant and ear height were identified in maize by combining association mapping and linkage analysis (Li et al. 2016b). Twenty-five candidate genes for soybean seed protein and oil content were identified by combining association mapping and linkage analysis (Zhang et al. 2019b). In our study, the combination of meta-QTL intervals and significant SNP identified by association mapping was used for candidate gene identification covering the whole cotton genome, which provides more comprehensive and reliable information. As a result, 297 candidate genes associated with cotton fiber quality traits were identified.

Candidate genes are probably involved in the development of cotton fibers

Due to the difficulty of forward genetics research in cotton, transcriptome analysis combined with QTLs has been widely used to identify candidate genes for fiber development (Fang et al. 2014; Shi et al. 2006; Tu et al. 2007; Yoo and Wendel 2014). In our study, transcriptome analysis of ten cotton tissues was used for 297 candidate genes expression pattern analysis, as a result, 20 genes showed high expression specifically in the developing fibers. In addition, the encoded proteins and functions of the 20 genes were annotated, and many were associated with fiber development (Table 4). *Gh_D11G1626* encoded a COBRA-like protein 4, and expressed in the fiber secondary wall biosynthesis stages; the function of COBRA-like protein has been reported in sorghum and rice, which were involved in secondary cell wall cellulose biosynthesis (Dai et al. 2011; Li et al. 2019; Sato et al. 2010), so *Gh_D11G1626* is probably involved in the secondary wall biosynthesis. *Gh_D13G1900* and *Gh_A01G1474* encoded WAT1-related proteins, and showed specifically high expression at the fiber secondary wall biosynthesis stages; the WAT1-related protein was probably related to high cotton fiber yield in cotton (Liu et al. 2018a). *Gh_D13G1965* and *Gh_A11G2663* encoded protein WVD2-like 1, and showed specifically high expression at the fiber secondary wall biosynthesis stages; in Arabidopsis, WVD2 was involved in the cell expansion (Yuen et al. 2003). *Gh_D08G1970* encoded a probable aquaporin PIP1-2, and showed specifically high expression at fiber elongation stages; aquaporin proteins were reported to be involved in the cotton fiber cell elongation and development (Li et al. 2013; Yang and Cui 2009). *Gh_A10G2036* encoded a Rop guanine nucleotide exchange factor 5, and showed specifically high expression at fiber elongation stages; Rho of plants (ROP) was reported participated in the spatial patterning of secondary cell walls and regulated the polarized cell growth (Kost 2008; Oda and Fukuda 2014; Yanagisawa et al. 2018), so *Gh_A10G2036* is probably involved in the cotton fiber elongation. According to the results, the candidate genes identified by the combination of meta-QTL intervals, significant SNPs, and transcriptome data, are reliable, however, there are still lots of work to do to study the function of the key candidate genes.

Table 4 The functional information of the key candidate gene homologous

Gene	Higher expression stages	Homologous function	Reference
<i>Gh_D11G1626</i>	Fiber secondary wall biosynthesis stages	Cellulose biosynthesis	(Dai et al. 2011)
<i>Gh_D13G1900</i> <i>Gh_A01G1474</i>	Fiber secondary wall biosynthesis stages	Fiber development	(Liu et al. 2018a)
<i>Gh_D13G1965</i> <i>Gh_A11G2663</i>	Fiber secondary wall biosynthesis stages	Cell expansion	(Yuen et al. 2003)
<i>Gh_D08G1970</i>	Fiber elongation stages	Fiber cell elongation	(Li et al. 2013)
<i>Gh_A10G2036</i>	Fiber elongation stages	Polarized cell growth	(Oda and Fukuda 2014)

Conclusion

In the study, we identified 74 meta-QTLs and 297 candidate genes associated with cotton fiber quality traits, and 20 of which showed high expression specifically in the developing fibers and thus are assumed to be associated with the fiber development. The study provides not only stable QTLs used for marker-assisted selection (MAS), but also candidate genes to uncover the molecular mechanisms for cotton fiber development.

Declarations

Acknowledgements

We would like to thank the anonymous reviewers for their valuable comments and helpful suggestions which help to improve the manuscript.

Authors' contributions

NX, ZD, XS, and PZ designed and performed the experiments. NX, XS and PZ wrote the main manuscript text and prepared all figures. XS, YF, YQ and WT performed data analysis. NX, PZ, ZD, LZ, and ZL, revised and polished the manuscript. All authors contributed in the interpretation of results and approved the final manuscript.

Funding

This work was supported by the National Natural Science Foundation of China (31760402), Public welfare research projects in the Autonomous Region (KY2019002), and Special Programs for New

Varieties Cultivation of Shihezi University (YZZX201701).

Availability of data and materials

The datasets used and analyzed during the current study are available from the corresponding author on reasonable request.

Ethics approval and consent to participate

Not applicable

Consent for publication

All Authors have provided ethical approval and consent to participate as well as consent for publication.

Competing interests

The authors have declared that no competing interests exist.

Author details

^aKey Laboratory of Oasis Ecology Agricultural of Xinjiang Bingtuan, Agricultural College, Shihezi University, Shihezi 832003, Xinjiang, China. ^bHubei Key Laboratory of Agricultural Bioinformatics, College of Informatics, Huazhong Agricultural University, Wuhan 430070, Hubei, China. ^cNational Key Laboratory of Crop Genetic Improvement, College of Plant Science and Technology, Huazhong Agricultural University, Wuhan 430070, Hubei, China. ^dResearch Institute of economic crops, Xinjiang Academy of Agricultural Sciences Urumqi 830091, China

References

- Ademe MS, He SP, Pan ZE, et al. Association mapping analysis of fiber yield and quality traits in Upland cotton (*Gossypium hirsutum* L.). *Mol Genet Genomics*. 2017; 292: 1267-1280. doi: 10.1007/s00438-017-1346-9.
- Ali I, Teng ZH, Bai YT, et al. A high density SLAF-SNP genetic map and QTL detection for fibre quality traits in *Gossypium hirsutum*. *BMC Genomics* 2018;19(1):879. doi: 10.1186/s12864-018-5294-5.
- Andersen JR, Schrag TA, Melchinger AE, et al. Validation of Dwarf8 polymorphisms associated with flowering time in elite European inbred lines of maize (*Zea mays* L.). *Theor Appl Genet*. 2005;111(2):206-217. doi: 10.1007/s00122-005-1996-6.
- Anderson JA. Marker-assisted selection for Fusarium head blight resistance in wheat. *Int J Food Microbiol*. 2007;119(1-2):51-53. doi: 10.1016/j.ijfoodmicro.

- Avni R, Oren L, Shabtay G, et al. Genome based meta-QTL analysis of grain weight in tetraploid wheat identifies rare alleles of GRF4 associated with larger grains. *Genes (Basel)*. 2018;9(12):636. doi: 10.3390/genes9120636.
- Cai CP, Zhu GZ, Zhang TZ, et al. High-density 80 K SNP array is a powerful tool for genotyping *G. hirsutum* accessions and genome analysis. *BMC Genomics*. 2017;18(1):654. doi: 10.1186/s12864-017-4062-2.
- Chandnani R, Kim C, Guo H, et al. Genetic analysis of gossypium fiber quality traits in reciprocal advanced backcross populations. *Plant Genome*. 2018;11(1):10.3835. doi: 10.3835/plantgenome2017.06.0057.
- Chen L, Bian JM, Shi HL, et al. Genetic analysis for the grain number heterosis of a super-hybrid rice WFYT025 combination using RNA-Seq. *Rice (N Y)*. 2018;11(1):37. doi: 10.1186/s12284-018-0229-y.
- Cobb JN, Biswas PS, Platten JD. Back to the future: revisiting MAS as a tool for modern plant breeding. *Theor Appl Genet*. 2019;132(3):647-667. doi: 10.1007/s00122-018-3266-4.
- Cui ZH, Xia AA, Zhang A, et al. Linkage mapping combined with association analysis reveals QTL and candidate genes for three husk traits in maize. *Theor Appl Genet*. 2018;131(10):2131-2144. doi: 10.1007/s00122-018-3142-2.
- Dai XX, You CJ, Chen GX, et al. OsBC1L4 encodes a COBRA-like protein that affects cellulose synthesis in rice. *Plant Mol Biol*. 2011;75(4-5):333-345. doi: 10.1007/s11103-011-9730-z.
- Diouf L, Magwanga RO, Gong WF, et al. QTL mapping of fiber quality and yield-related traits in an intra-specific upland cotton using genotype by sequencing (GBS). *Int J Mol Sci*. 2018;19(2):441. doi: 10.3390/ijms19020441.
- Fang L, Tian RP, Chen JD, et al. Transcriptomic analysis of fiber strength in upland cotton chromosome introgression lines carrying different *Gossypium barbadense* chromosomal segments. *PLoS One*. 2014;9(4):e94642. doi: 10.1371/journal.pone.0094642.
- Fang L, Wang Q, Hu Y, et al. Genomic analyses in cotton identify signatures of selection and loci associated with fiber quality and yield traits. *Nat Genet*. 2017;49(7):1089-1098. doi: 10.1038/ng.3887.
- Gapare WJ, Conaty W, Zhu QH, et al. Genome-wide association study of yield components and fibre quality traits in a cotton germplasm diversity panel. *Euphytica*, 2017; 213, 66. doi: 10.1007/s10681-017-1855-y.
- Goffinet B, Gerber S. Quantitative trait loci: a meta-analysis. *Genetics*, 2000; 155: 463-473.
- Guo BH, Sleper DA, Lu P, et al. QTLs associated with resistance to soybean cyst nematode in soybean: meta-analysis of QTL locations. *Crop Sci*, 2006a; 46(2): 595-602. doi: 10.1007/s00122-005-0031-2.

- Guo WZ, Ma GJ, Zhu YC, et al. Molecular tagging and mapping of quantitative trait loci for lint percentage and morphological marker genes in upland cotton. *J Integr Plant Biol.* 2006b; 48, 320-326.
- Guo X, Guo YP, Ma J, et al. Mapping heterotic loci for yield and agronomic traits using chromosome segment introgression lines in cotton. *J Integr Plant Biol.* 2013;55(8):759-774. doi: 10.1111/jipb.12054.
- Han LB, Li YB, Wang HY, et al. The dual functions of WLIM1a in cell elongation and secondary wall formation in developing cotton fibers. *Plant Cell.* 2013;25(11):4421-4438. doi: 10.1105/tpc.113.116970.
- Handi SS, Katageri IS, Adiger S, et al. Association mapping for seed cotton yield, yield components and fibre quality traits in upland cotton (*Gossypium hirsutum* L.) genotypes. *Plant Breeding.* 2017; 136(6): 958-968.
- Huang C, Nie XH, Shen C, et al. Population structure and genetic basis of the agronomic traits of upland cotton in China revealed by a genome-wide association study using high-density SNPs. *Plant Biotechnol J.* 2017;15(11):1374-1386. doi: 10.1111/pbi.12722.
- Huang YQ, Wang J, Zhang LD, et al. A cotton annexin protein anxGb6 regulates fiber elongation through its interaction with actin 1. *PLoS One.* 2013;8(6):e66160. doi: 10.1371/journal.pone.0066160.
- Ijaz B, Zhao N, Kong J, et al. Fiber quality improvement in upland cotton (*Gossypium hirsutum* L.): quantitative trait loci mapping and marker assisted selection application. *Front Plant Sci.* 2019;10:1585. doi: 10.3389/fpls.2019.01585.
- Islam MS, Thyssen GN, Jenkins JN, et al. A MAGIC population-based genome-wide association study reveals functional association of GhRBB1_A07 gene with superior fiber quality in cotton. *BMC Genomics.* 2016;17(1):903. doi: 10.1186/s12864-016-3249-2.
- Jia XY, Wang HT, Pang CY, et al. QTL delineation for five fiber quality traits based on an intra-specific *Gossypium hirsutum* L. recombinant inbred line population. *Mol Genet Genomics.* 2018;293(4):831-843. doi: 10.1007/s00438-018-1424-7.
- Keerio AA, Shen C, Nie YC, et al. 2018. QTL mapping for fiber quality and yield traits based on introgression lines derived from *Gossypium hirsutum* x *G. tomentosum*. *Int J Mol Sci.* 2018;19(1):243. doi: 10.3390/ijms19010243.
- Kost B. Spatial control of Rho (Rac-Rop) signaling in tip-growing plant cells. *Trends in Cell Biology.* 2008; 18(3): 119-127. doi: 10.1016/j.tcb.2008.01.003.
- Krzywinski M, Schein JE, Birol I, et al. Circos: An information aesthetic for comparative genomics. *Genome Res.* 2009;19(9):1639-1645. doi: 10.1101/gr.092759.109.
- Li C, Dong YT, Zhao TL, et al. Genome-wide SNP linkage mapping and QTL analysis for fiber quality and yield traits in the upland cotton recombinant inbred lines population. *Front Plant Sci.* 2016a;7:1356. doi:

10.3389/fpls.2016.01356.

Li CQ, Fu YZ, Sun RR, et al. Single-locus and multi-locus genome-wide association studies in the genetic dissection of fiber quality traits in upland cotton (*Gossypium hirsutum* L.). *Front Plant Sci.* 2018;9:1083. doi: 10.3389/fpls.2018.01083.

Li DD, Ruan XM, Zhang J, et al. Cotton plasma membrane intrinsic protein 2s (PIP2s) selectively interact to regulate their water channel activities and are required for fibre development. *New Phytol.* 2013;199(3):695-707. doi: 10.1111/nph.12309.

Li Y, Wang NN, Wang Y, et al. The cotton XLIM protein (GhXLIM6) is required for fiber development via maintaining dynamic F-actin cytoskeleton and modulating cellulose biosynthesis. *Plant J.* 2018 Dec;96(6):1269-1282. doi: 10.1111/tpj.14108.

Li P, Liu YR, Tan WQ, et al. Brittle Culm 1 encodes a COBRA-like protein involved in secondary cell wall cellulose biosynthesis in sorghum. *Plant Cell Physiol.* 2019;60(4):788-801. doi: 10.1093/pcp/pcy246.

Li TG, Ma XF, Li NY, et al. 2017a. Genome-wide association study discovered candidate genes of Verticillium wilt resistance in upland cotton (*Gossypium hirsutum* L.). *Plant Biotechnol J.* 2017a;15(12):1520-1532. doi: 10.1111/pbi.12734.

Li XB, Fan XP, Wang XL, et al. The cotton ACTIN1 Ggene is functionally expressed in fibers and participates in fiber elongation. *Plant Cell.* 2005;17(3):859-875. doi: 10.1105/tpc.104.029629.

Li XH, Wu M, Liu GY, et al. Identification of candidate genes for fiber length quantitative trait loci through RNA-Seq and linkage and physical mapping in cotton. *BMC Genomics.* 2017b;18(1):427. doi: 10.1186/s12864-017-3812-5.

Li XP, Zhou ZJ, Ding JQ, et al. Combined linkage and association mapping reveals QTL and candidate genes for plant and ear height in maize. *Front Plant Sci.* 2016b;7:833. doi: 10.3389/fpls.2016.00833.

Liu C, Zeng LB, Zhu SY, et al. Draft genome analysis provides insights into the fiber yield, crude protein biosynthesis, and vegetative growth of domesticated ramie (*Boehmeria nivea* L. Gaud). *DNA Res.* 2018a;25(2):173-181. doi: 10.1093/dnares/dsx047.

Liu RX, Gong JW, Xiao XH, et al. GWAS analysis and QTL identification of fiber quality traits and yield components in upland cotton using enriched high-density SNP markers. *Front Plant Sci.* 2018b;9:1067. doi: 10.3389/fpls.2018.01067.

Liu RZ, Wang BH, Guo WZ, et al. Quantitative trait loci mapping for yield and its components by using two immortalized populations of a heterotic hybrid in *Gossypium hirsutum* L. *Molecular Breeding*, 2012; 29(2):297-311. doi: 10.1007/s11032-011-9547-0

- Ma JJ, Geng YH, Pei WF, et al. Genetic variation of dynamic fiber elongation and developmental quantitative trait locus mapping of fiber length in upland cotton (*Gossypium hirsutum* L.). *BMC Genomics*. 2018a;19(1):882. doi: 10.1186/s12864-018-5309-2.
- Ma JJ, Liu J, Pei WF, et al. Genome-wide association study of the oil content in upland cotton (*Gossypium hirsutum* L.) and identification of GhPRXR1, a candidate gene for a stable QTLqOC-Dt5-1. *Plant Sci*. 2019;286:89-97. doi: 10.1016/j.plantsci.2019.05.019.
- Ma ZY, He SP, Wang XF, et al. Resequencing a core collection of upland cotton identifies genomic variation and loci influencing fiber quality and yield. *Nat Genet*. 2018b;50(6):803-813. doi: 10.1038/s41588-018-0119-7.
- Mahuku G, Chen JF, Shrestha R, et al. Combined linkage and association mapping identifies a major QTL (qRtsc8-1), conferring tar spot complex resistance in maize. *Theor Appl Genet*. 2016;129(6):1217-1229. doi: 10.1007/s00122-016-2698-y.
- Martinez AK, Soriano JM, Tuberosa R, et al. Yield QTLome distribution correlates with gene density in maize. *Plant Sci*. 2016;242:300-309. doi: 10.1016/j.plantsci.2015.09.022.
- Nie XH, Huang C, You CY, et al. Genome-wide SSR-based association mapping for fiber quality in nationwide upland cotton inbred cultivars in China. *BMC Genomics*. 2016;17:352. doi: 10.1186/s12864-016-2662-x.
- Oda Y, Fukuda H. Emerging roles of small GTPases in secondary cell wall development. *Front Plant Sci*. 2014;5:428. doi: 10.3389/fpls.2014.00428.
- Qin HD, Guo WZ, Zhang YM, et al. QTL mapping of yield and fiber traits based on a four-way cross population in *Gossypium hirsutum* L. *Theor Appl Genet*. 2008;117(6):883-894. doi: 10.1007/s00122-008-0828-x.
- Reyna NS, Sneller C. Evaluation of marker-assisted introgression of yield QTL alleles into adapted soybean. *Crop Sci*. 2001; 41, 1317-1321.
- Ribaut J, Ragot M. Marker-assisted selection to improve drought adaptation in maize: the backcross approach, perspectives, limitations, and alternatives. *J Exp Bot*. 2007;58(2):351-360. doi: 10.1093/jxb/erl214.
- Rodgers J, Zumba J, Fortier C. Measurement comparison of cotton fiber micronaire and its components by portable near infrared spectroscopy instruments. *Textile Research Journal*. 2017; 87, 0040517515622153.
- Said JI, Knapka JA, Song M, et al. Cotton QTLdb: a cotton QTL database for QTL analysis, visualization, and comparison between *Gossypium hirsutum* and *G. hirsutum* × *G. barbadense* populations. *Mol Genet Genomics*. 2015a;290(4):1615-1625. doi: 10.1007/s00438-015-1021-y.

- Said JI, Lin ZX, Zhang XL, et al. A comprehensive meta QTL analysis for fiber quality, yield, yield related and morphological traits, drought tolerance, and disease resistance in tetraploid cotton. *BMC Genomics*. 2013a;14:776. doi: 10.1186/1471-2164-14-776.
- Said JI, Song MZ, Wang HT, et al. A comparative meta-analysis of QTL between intraspecific *Gossypium hirsutum* and interspecific *G.hirsutum* × *G.barbadense* populations. *Mol Genet Genomics*. 2015b;290(3):1003-1025. doi: 10.1007/s00438-014-0963-9.
- Salnikov VV, Grimson MJ, et al. Localization of sucrose synthase and callose in freeze-substituted secondary-wall-stage cotton fibers. *Protoplasma*. 2003;221(3-4):175-184. doi: 10.1007/s00709-002-0079-7.
- Salvi S, Tuberosa R. The crop QTLome comes of age. *Curr Opin Biotechnol*. 2015;32:179-185. doi: 10.1016/j.copbio.2015.01.001.
- Sato K, Ito S, Fujii T, et al. The carbohydrate-binding module (CBM)-like sequence is crucial for rice CWA1/BC1 function in proper assembly of secondary cell wall materials. *Plant Signal Behav*. 2010;5(11):1433-1436. doi: 10.4161/psb.5.11.13342.
- Sebastian SA, Streit LG, Stephens PA, et al. Context-specific marker-assisted selection for improved grain yield in elite soybean populations. *Crop Sci*, 50, 1196-1206. doi: 10.2135/cropsci2009.02.0078
- Shen XL, Guo WZ, Lu QX, et al. Genetic mapping of quantitative trait loci for fiber quality and yield trait by RIL approach in upland cotton. *Euphytica*. 2007; 155(3): 371-380. doi: 10.1186/s12864-018-4890-8.
- Shi YH, Zhu SW, Mao XZ, et al. Transcriptome profiling, molecular biological, and physiological studies reveal a major role for ethylene in cotton fiber cell elongation. *Plant Cell*. 2006;18(3):651-664. doi: 10.1105/tpc.105.040303.
- Shimono H, Abe A, Aoki N, et al. Combining mapping of physiological quantitative trait loci and transcriptome for cold tolerance for counteracting male sterility induced by low temperatures during reproductive stage in rice. *Physiol Plant*. 2016;157(2):175-192. doi: 10.1111/ppl.12410.
- Stewart JMD. 1975. Fiber initiation on the cotton ovule (*Gossypium hirsutum*). *American Journal of Botany* 62: 723– 730.
- Su JJ, Li LB, Pang CY, et al. Two genomic regions associated with fiber quality traits in Chinese upland cotton under apparent breeding selection. *Sci Rep*. 2016;6:38496. doi: 10.1038/srep38496.
- Su JJ, Ma Q, Li M, et al. Multi-locus genome-wide association studies of fiber-quality related traits in Chinese early-maturity upland cotton. *Front Plant Sci*. 2018;9:1169. doi: 10.3389/fpls.2018.01169.
- Sun FD, Zhang JH, Wang SF, et al. QTL mapping for fiber quality traits across multiple generations and environments in upland cotton. *Molecular Breeding*, 2012, 30(1): 569-582. doi: 10.1186/1471-2229-10-

- Sun ZW, Wang XF, Liu ZW, et al. Genome-wide association study discovered genetic variation and candidate genes of fibre quality traits in *Gossypium hirsutum* L. *Plant Biotechnol J*. 2017;15(8):982-996. doi: 10.1111/pbi.12693.
- Sun ZG, Wang XF, Liu ZW, et al. A genome-wide association study uncovers novel genomic regions and candidate genes of yield-related traits in upland cotton. *Theor Appl Genet*. 2018;131(11):2413-2425. doi: 10.1007/s00122-018-3162-y.
- Swamy BPM, Vikram P, Dixit S, et al. Meta-analysis of grain yield QTL identified during agricultural drought in grasses showed consensus. *BMC Genomics*. 2011;12:319. doi: 10.1186/1471-2164-12-319.
- Tan ZY, Zhang ZQ, Sun XJ, et al. Genetic map construction and fiber quality QTL mapping using the cottonSNP80K array in upland cotton. *Front Plant Sci*. 2018a;9:225. doi: 10.3389/fpls.2018.00225.
- Tang WX, He YH, Tu LL, et al. Down-regulating annexin gene GhAnn2 inhibits cotton fiber elongation and decreases Ca²⁺ influx at the cell apex. *Plant Mol Biol*. 2014;85(6):613-625. doi: 10.1007/s11103-014-0208-7.
- Tu LL, Zhang XL, Liang SG, et al. Genes expression analyses of sea-island cotton (*Gossypium barbadense* L.) during fiber development. *Plant Cell Rep*. 2007;26(8):1309-1320. doi: 10.1007/s00299-007-0337-4.
- Vasconcellos RCC, Oraguzie OB, Soler A, et al. Meta-QTL for resistance to white mold in common bean. *PLoS One*. 2017;12(2):e0171685. doi: 10.1371/journal.pone.0171685.
- Visscher PM. Sizing up human height variation. *Nat Genet*. 2008;40(5):489-490. doi: 10.1038/ng0508-489.
- Walford S, Wu YR, Llewellyn DJ, et al. GhMYB25-like: a key factor in early cotton fibre development. *Plant J*. 2011;65(5):785-797. doi: 10.1111/j.1365-313X.2010.04464.x.
- Walford S, Wu YR, Llewellyn DJ, et al. Epidermal cell differentiation in cotton mediated by the homeodomain leucine zipper gene, GhHD-1. *Plant J*. 2012;71(3):464-478. doi: 10.1111/j.1365-313X.2012.05003.x.
- Wang HY, Wang J, Gao P, et al. Down-regulation of GhADF1 gene expression affects cotton fibre properties. *Plant Biotechnol J*. 2009;7(1):13-23. doi: 10.1111/j.1467-7652.2008.00367.x.
- Wang HT, Huang C, Zhao WX, et al. Identification of QTL for fiber quality and yield traits using two immortalized backcross populations in upland cotton. *PLoS One*. 2016;11(12):e0166970. doi: 10.1371/journal.pone.0166970.

- Wang HT, Zhang RT, Shen C, et al. Transcriptome and QTL analyses reveal candidate genes for fiber quality in Upland cotton. *Crop Journal*. 2020; 8 (1): 98-106.
- Wang J, Wang HY, Zhao PM, et al. Overexpression of a profilin (GhPFN2) promotes the progression of developmental phases in cotton fibers. *Plant Cell Physiol*. 2010;51(8):1276-1290. doi: 10.1093/pcp/pcq086.
- Wang L, Cook A, Patrick JW, et al. Silencing the vacuolar invertase gene GhVIN1 blocks cotton fiber initiation from the ovule epidermis, probably by suppressing a cohort of regulatory genes via sugar signaling. *Plant J*. 2014;78(4):686-696. doi: 10.1111/tpj.12512.
- Wang YK, Ning ZY, Hu Y, et al. Molecular mapping of restriction-site associated DNA markers in allotetraploid upland cotton. *PLoS One*. 2015;10(4):e0124781. doi: 10.1371/journal.pone.0124781.
- Wen TW, Wu M, Shen C, et al. Linkage and association mapping reveals the genetic basis of brown fibre (*Gossypium hirsutum*). *Plant Biotechnol J*. 2018;16(9):1654-1666. doi: 10.1111/pbi.12902.
- Wu XM, Wang B, Xie FG, et al. QTL mapping and transcriptome analysis identify candidate genes regulating pericarp thickness in sweet corn. *BMC Plant Biol*. 2020;20(1):117. doi: 10.1186/s12870-020-2295-8.
- Xiao WM, Yang QY, Huang M, et al. Improvement of rice blast resistance by developing monogenic lines, two-gene pyramids and three-gene pyramid through MAS. *Rice (N Y)*. 2019;12(1):78. doi: 10.1186/s12284-019-0336-4
- Yanagisawa M, Alonso JM, Szymanski DB. Microtubule-dependent confinement of a cell signaling and actin polymerization control module regulates polarized cell growth. *Curr Biol*. 2018;28(15):2459-2466.e4. doi: 10.1016/j.cub.2018.05.076.
- Yang SS, Cui LR. The action of aquaporins in cell elongation, salt stress and photosynthesis. *Chinese Journal of Biotechnology*. 2009;25(3):321-327.
- Yang X, Wang Y, Zhang G, et al. Detection and validation of one stable fiber strength QTL on c9 in tetraploid cotton. *Mol Genet Genomics*. 2016;291(4):1625-1638. doi: 10.1007/s00438-016-1206-z.
- Yoo M, Wendel JF. Comparative evolutionary and developmental dynamics of the cotton (*Gossypium hirsutum*) fiber transcriptome. *PLoS Genet*. 2014;10(1):e1004073. doi: 10.1371/journal.pgen.1004073.
- Yu JW, Yu SX, Gore MA, et al. Identification of quantitative trait loci across interspecific F2, F2:3 and testcross populations for agronomic and fiber traits in tetraploid cotton. *Euphytica*. 2013; 191, 375-389. doi: 10.1007/s10681-013-0875-5
- Yuen CYL, Pearlman R, Silosuh L, et al. WVD2 and WDL1 modulate helical organ growth and anisotropic cell expansion in arabidopsis. *Plant Physiol*. 2003;131(2):493-506. doi: 10.1104/pp.015966.

Zhang C, Li LB, Liu QB, et al. Identification of loci and candidate genes responsible for fiber length in upland cotton (*Gossypium hirsutum* L.) via association mapping and linkage analyses. *Front Plant Sci.* 2019a;10:53. doi: 10.3389/fpls.2019.00053.

Zhang JF, Fang H, Zhou HP, et al. Genetics, breeding, and marker-assisted selection for Verticillium Wilt resistance in cotton. *Crop Sci.* 2014; 54: 1289-1303.

Zhang TZ, Hu Y, Jiang WK, et al. Sequencing of allotetraploid cotton (*Gossypium hirsutum* L. acc. TM-1) provides a resource for fiber improvement. *Nat Biotechnol.* 2015b;33(5):531-537. doi: 10.1038/nbt.3207.

Zhang TF, Wu TT, Wang LW, et al.. A Combined linkage and GWAS analysis identifies QTLs linked to soybean seed protein and oil content. *Int J Mol Sci.* 2019b; 20 (23): 5915. doi: 10.3390/ijms20235915.

Zhang XX, Guan ZR, Wang L, et al. Combined GWAS and QTL analysis for dissecting the genetic architecture of kernel test weight in maize. *Mol Genet Genomics.* 2019c;295(2):409-420. doi: 10.1007/s00438-019-01631-2.

Zhang Z, Li JW, Muhammad J, et al. High resolution consensus mapping of quantitative trait loci for fiber strength, length and micronaire on Chromosome 25 of the upland cotton (*Gossypium hirsutum* L.). *PLoS One.* 2015b;10(8):e0135430. doi: 10.1371/journal.pone.0135430.

Zou XY, Gong JW, Duan L, et al. High-density genetic map construction and QTL mapping for fiber strength on Chr24 across multiple environments in a CCRI70 recombinant inbred lines population. *Euphytica.* 2014; 214 (6), 102. doi: 10.1007/s10681-018-2177-4