

# Anaerobic Treatment of Oily Wastewater Using A Biofilm-Electrode Reactor: A Kinetic Study And Energy Consumption

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## Research Article

**Keywords:** Bioelectrochemical, Anaerobic, Oily Wastewater, Kinetic, Stover-Kincannon model

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21 modified Stover-Kincannon model. The present study findings indicated that BER is a promising  
22 method for the treatment of oil-contaminated wastewaters.

23

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25

## 26 **1. Introduction**

27 Recently, oil-contaminated wastewater is considered one of the environmental problems. The large  
28 quantity of these wastewaters is generated by various sources due to the rapid growth of  
29 industrialization and urbanization over the past decades. Oils can enter the environment at different  
30 stages of production, transportation, refining, and use (Zhu et al., 2018). The main sources of oil-  
31 contaminated wastewater include food industries, oil refineries, petrochemical companies, and  
32 metal, textiles, and leather producing plants, as well as restaurants, kitchens, and vehicles  
33 (Mirshafiee et al., 2018). Discharge of oily effluent to the environment may cause irreparable  
34 damages. Their very low solubility in water and very poor degradability cause detrimental effects  
35 on the environment (Gurd et al., 2020). Layers of oil and grease can threaten aquatic life by  
36 reducing oxygen and light penetration (Sharma et al., 2020). Oil, at higher levels, harms aquatic  
37 life, creates obnoxious odors and unpleasant sights, reduces tourism activities, and causes  
38 economic damage (Brillas et al., 2020). Various physical, chemical, and biological methods are  
39 introduced for the removal of oil from wastewater, of which biological systems are significantly  
40 used due to their advantages, of which more compatibility with the environment is noteworthy.  
41 Moreover, no environmentally harmful chemicals are used in biological systems; therefore, their  
42 effluent and sludge are less hazardous to the receiving waters than the chemical systems. These

43 features make bioelectrochemical systems a cost-effective and environmental-friendly approach.  
44 Biodegradation of organic matter is triggered by supporting microbial growth and creating  
45 optimum environmental conditions to turn pollutants into carbon dioxide and other gases,  
46 inorganic matter, water, safe and stable substances, and biomass. Among biological methods,  
47 biofilm reactors have advantageous properties. It is well understood that biofilm reactors are  
48 suitable for the treatment of effluents containing poorly biodegradable compounds and  
49 decomposable organic matter. Biofilm, formed by the fixation on active microorganisms, increases  
50 biomass and improves organic and hydraulic loading (Karadag et al.,2015). Fixed biofilm is  
51 effective in reducing toxic compounds and the treatment of wastewater containing biodegradable  
52 matter (Zhang et al., 2015). Other advantages of biofilm systems are longer shelf life, more diverse  
53 microbial species, and increased stability(Tan et al., 2018). Numerous researchers tried to treat  
54 oily effluent in anaerobic systems (Chan, et al., 2010; Wang et al., 2016). The anaerobic process,  
55 widely used for industrial wastewater treatment, has advantages such as the generation of less  
56 sludge, lower costs, less energy consumption, no need for aeration, low nutritional requirements,  
57 high organic load tolerance, high shock loading resistance, and production of methane gas. The  
58 improvement and repair of this process are faster if it is used in industries closed for a period in  
59 the year with no utilization. It also has disadvantages as the quality of wastewater is not high and  
60 does not meet the effluent disposal standards (Kong, et al., 2019). Therefore, an anaerobic system  
61 needs further measures to be improved and upgraded, for example, increasing the activity of  
62 bacteria by electrostimulation (Adibzadeh, et al., 2016). Bioelectrochemical systems (BESs) or  
63 biofilm electrode reactors (BERs), as a relatively new technology, are appropriate and promising  
64 methods with great potentials for wastewater treatment and are considered clean technology. In  
65 this technology, microorganisms are used as an electrode-attached biofilm. In this method,

66 oxidation-reduction (redox) reactions are catalyzed by the interaction between the electrode and  
67 the biofilm. In recent years, wastewater treatment by BESs is critically considered by  
68 researchers(Cao, et al., 2018). Evidence suggests that induced current can stimulate metabolism  
69 and enhance biochemical function in bacteria. The applied current increases the rate of ion  
70 migration and reactions on the surface of the electrodes. Lower voltages are used in these systems,  
71 which can overcome some problems such as corrosion of the anode and high energy consumption  
72 observed in the electrochemical method. The induced current must be adequate, otherwise inverse  
73 results are obtained, and the activity of microorganisms is restrained (Liu, et al., 2015). Due to the  
74 increasing generation of oil-contaminated wastewater and the benefits of applying the anaerobic  
75 method, and that this method needs upgrading and improvement of efficiency, the present study  
76 aimed at investigating increased efficiency of anaerobic treatment of oil-contaminated wastewater  
77 by electrostimulation and BESs. The main objectives of the present study were: (1) adaptation of  
78 bacteria to BES under anaerobic conditions and determination of the effect of changes in the  
79 applied electrical current on biomass performance by removal efficiency, (2) investigation of the  
80 effect of operational parameters such as initial pollutant concentration, reaction time, and  
81 supporting electrolyte, (3) improvement of the anaerobic process by BESs and comparison of the  
82 removal efficiency of BES with a conventional method in the absence of applied current, (4)  
83 estimation of energy consumed, and (5) kinetic study of the substrate removal process.

## 84 **2. Materials and methods**

### 85 *2.1. Experimental setup*

86 A cylindrical Plexiglas reactor, with an effective volume of 2.25 L, was used in the present study.  
87 It contained stainless steel electrodes fixed by a holder. It was covered to provide anaerobic

88 conditions. The anode electrode was steel mesh, and its cavities were so small to facilitate the  
89 loading of biomass. A magnetic stirrer (Alfa, HS-860, Iran) was used throughout the experiments  
90 for gently mixing and homogenizing the reactor contents. Direct current was supplied by a power  
91 supply (ATTEN APS3005S-3D, China). The electrodes were washed with HCl, rubbed with a  
92 sponge, and rinsed with distilled water in order to be prepared.

### 93 *2.2. Wastewater preparation*

94 Synthetic oil emulsions were prepared by adding edible oil and sodium dodecyl sulfate, as an  
95 emulsifier, to distilled water and mixing for one hour at 900 rpm (Mirshafiee et al., 2018).

### 96 *2.3. Experimental procedure*

97 The current experimental, laboratory-scale study was performed as a batch system in a BER under  
98 anaerobic conditions. The seed sludge was taken from a wastewater treatment plant (Tehran, Iran),  
99 washed three times with tap water to remove impurities, and used as a microbial inoculant. All  
100 experiments were performed at ambient temperature ( $25\pm 2^{\circ}\text{C}$ ) and neutral pH range. Nutrients,  
101 added to enrich the bacteria, were composed of  $\text{NH}_4\text{Cl}$  0.50 g/L,  $\text{KH}_2\text{PO}_4$  0.25 g/L,  $\text{K}_2\text{HPO}_4$  0.25  
102 g/L,  $\text{MgCl}_2$  0.30 g/L,  $\text{CoCl}_2$  25 mg/L,  $\text{ZnCl}_2$  11.50 mg/L,  $\text{CuCl}_2$  10.50 mg/L,  $\text{CaCl}_2$  5 mg/L,  $\text{MnCl}_2$   
103 15 mg/L,  $\text{NiSO}_4$  16 mg/L, and  $\text{FeCl}_3$  25 mg/L.

104 All chemicals used in the experiments had an analytical degree. After the formation of biofilm and  
105 acclimatization of biomass, the effects of different operating parameters, such as changes in the  
106 applied current (5-30 mA), initial concentration of COD (1500 - 5000 mg/L), reaction time (1-3  
107 days) and supporting electrolyte (50-200 mg/L), were evaluated. In addition, the bacterial  
108 community was investigated under optimal conditions, and the amount of energy consumed was  
109 calculated.

110 *2.4. Analysis*

111 COD concentration was measured by the closed reflux method, described in the standard method.  
112 The samples were analyzed using a spectrophotometer (Rayleigh, Vis-7220 / UV-9200). The  
113 following equation was used to calculate the removal efficiency (Re%):

114 
$$\text{Removal Efficiency} = \frac{C_0 - C_t}{C_0} \times 100 \quad (1)$$

115 where (C<sub>0</sub>) is the initial concentration (mg.L<sup>-1</sup>) and (C<sub>t</sub>) the concentration at any time (mg.L<sup>-1</sup>). A  
116 portable pH meter (Oaklon, Malaysia) was used to measure the pH. A scanning electron  
117 microscopy (SEM, Seron technology, AIS-2100) was used to investigate the biofilm morphology.

118 *2.5. Kinetic modeling*

119 Kinetic models can be useful in the design and modeling of bioreactors, predict their performance,  
120 determine the relationship between variables, and optimize the design. In the present study, three  
121 common models of first-order, Grau model, and modified Stover–Kincannon were evaluated to  
122 investigate the kinetics of the biological reactions.

123 *2.5.1. First-order kinetic model*

124 Assuming that the first-order kinetic reactor is predominant, the changes in substrate concentration  
125 in a completely-mixed system were expressed as follows:

126 
$$-\frac{ds}{dt} = \frac{Q S_0}{V} - \frac{Q S_e}{V} - K_1 S_e \quad (2)$$

127 In a biological reactor, under steady-state conditions, changes in the removal of substrate  
128 concentrations (-ds/dt) are negligible, so Equation 2 can be modified to Equation 3:

129 
$$\frac{S_0 - S_e}{\text{HRT}} = K_1 S_e \quad (3)$$

130 where  $S_0$  and  $S_e$  are the substrate concentrations of influent and effluent (mg/L),  $Q$  the flow rate  
 131 of wastewater (L/d),  $V$  effective reactor volume (L), HRT hydraulic retention time (day), and  $K_1$   
 132 the first-order substrate removal rate constant (1/d).

133 The  $K_1$  can be obtained by plotting  $[(S_0 - S_e)/\text{HRT}]$  versus  $S_e$ . It can also be calculated based on the  
 134 slope of the line.

135 *2.5.2. Grau model*

136 The Grau model represents the second-order kinetics, which can be expressed by the following  
 137 equation:

138 
$$- \frac{ds}{dt} = K_2 \cdot X \left( \frac{S_e}{S_0} \right)^2 \quad (4)$$

139 If Equation 4 is integrated and then linearized, Equation 5 is obtained as follows:

140 
$$\frac{S_0 \text{ HRT}}{S_0 - S_e} = \text{HRT} - \frac{S_0}{K_2 X} \quad (5)$$

141 By holding the second part of the right side of Equation 5 as a constant value of  $a$ , Equation 6 is  
 142 obtained as follows:

143 
$$\frac{S_0 \text{ HRT}}{S_0 - S_e} = a + b\text{HRT} \quad (6)$$

144  $(S_0 - S_e)/S_0$  expresses the substrate removal efficiency and is symbolized as  $E$ . Therefore, the last  
 145 equation is as follows:



146 
$$\frac{\text{HRT}}{E} = a + b\text{HRT} \quad (7)$$

147 where a and b are second-order kinetic constants, a  $S_0/K_2X$ , and b dimensionless.

148 The kinetic parameters of a and b can be determined by the intercept and slope of the plotline  $S_0$   
 149  $\text{HRT}/(S_0-S_e)$  versus HRT, respectively(Pahlavanzadeh et al., 2018).

150 *2.5.3. Modified Stover-Kincannon model*

151 Generally, the modified Stover-Kincannon model is widely used to evaluate the kinetic  
 152 parameters of biofilm reactors. This model is expressed as follows:

153 
$$\frac{dS}{dt} = \frac{Q(S_0 - S_e)}{V} = \frac{U_{\max} \left( \frac{QS_0}{V} \right)}{K_B + \left( \frac{QS_0}{V} \right)} \quad (8)$$

154 where  $K_B$  and  $U_{\max}$  represent saturation constant (mg/L.day) and maximum substrate removal rate  
 155 (mg/L.day), respectively. The linear form of this equation is expressed as Equation 9:

156 
$$\left( \frac{dS}{dt} \right)^{-1} = \frac{V}{Q(S_0 - S_e)} = \frac{K_B}{U_{\max}} \cdot \left( \frac{V}{QS_0} \right) + \frac{1}{U_{\max}} \quad (9)$$

157 Plotting  $V/[Q(S_0 - S_e)]$  versus  $(V/QS_0)$  creates a straight line that gives the intercept and slope of  
 158 the line as  $1/U_{\max}$  and  $K_B/U_{\max}$ , respectively.

159 **3. Results and discussion**

160 *3.1. Start-up and biomass adaptation process*

161 Adaptation and acclimation is an important stage in the biological process. It is more sensitive in  
 162 the BESs since, in addition to pollutants and wastewater, the biomass should also be adapted to

163 electric current. Oil-contaminated compounds usually contain substances that are not easily  
164 degraded by bacteria in nature, so the bacteria should be acclimatized to the environment. The  
165 gradual increase in pollutant concentration is a method that can be used for better adaptation. This  
166 strategy prevents severe shocks, and by gradually adding oil-contaminated wastewater, the bacteria  
167 are given the chance to adapt to the environment. Figure 1 shows the biomass adaptation results.  
168 COD concentration at this stage was 1500 mg/L, and HRT was considered three constant days.  
169 Glucose was first used to acclimatize the bacteria to oil-contaminated wastewater as it is a palatable  
170 organic matter for them. During the experiment, glucose levels were reduced and the oil-  
171 contaminated wastewater was gradually added. On the other hand, since the electric current causes  
172 bacterial lysis, a very low current was used in the adaptation stage in order to acclimatize them  
173 over time. The induction current was 1 mA at this stage. The reactor was managed until reaching  
174 stable conditions. The removal efficiency decreased to some extent after each increase in the  
175 concentration of oil-contaminated wastewater, indicating that the system was shocked. The  
176 bacteria then adapted to new conditions, and the system improved over time by continuing the  
177 operation. The operation proceeded slowly until the entire influent was just the oil-contaminated  
178 wastewater. The adaptation period lasted 105 days, and at the end of the start-up period, the  
179 influent substrate was only the oil-contaminated wastewater.

### 180 *3.2. Effect of applied current*

181 Figure 2 shows the effect of the intensity of different applied currents on the bioelectrochemical  
182 removal rate. In the present study, the current intensities of 5, 10, 15, 20, 25, and 30 mA were  
183 investigated. Hydraulic retention time and initial COD concentration were considered constant.  
184 Evaluation of the effect of changes in the applied current on the removal efficiency using BES  
185 indicated that by applying a current intensity of 5 mA, the removal efficiency was 72.3%, and the

186 applied current intensity increased after reaching stable conditions. With increasing current  
187 intensity above 15 mA, the removal efficiency decreased, and the decreasing trend even continued  
188 by increasing the current intensity. In other words, a very high increase in current intensity reduced  
189 efficiency. As shown in Figure 2b, maximum efficiency was obtained at 15 mA that was 83.4%.  
190 Therefore, it was selected as the optimum current and used in the experiment. The obtained results  
191 show that the optimum current has a beneficial effect on bacteria and their enzymatic activity.  
192 When an electric field is applied to the microbial system, the permeability of the cytoplasmic  
193 membrane increases, and as a result, nutrients better pass through the cell membrane. Increasing  
194 the electrical field can also affect the activation of species carrying electrons. These species  
195 directly affect enzymes and promote bacterial metabolism. The activity and metabolism of most  
196 microorganisms increase by electrostimulation or biostimulation (Zhang et al., 2014).  
197 Electrostimulation of cells improves DNA and protein synthesis, cell membrane permeability, and  
198 growth and can promote the biological removal of COD (Wei et al., 2011). Likewise, the direct  
199 oxidation can affect the removal of contaminants, in which the organic matter is adsorbed on the  
200 surface of the anode and is removed (García-García et al., 2015). Over-optimum current intensities  
201 have adverse effects on bacteria. High current intensity can cause direct oxidation in cell structure  
202 and death. In addition, it makes alterations in the permeability of the cell wall so that the molecules  
203 diffuse to the outside (Adibzadeh, et al., 2016). High electric current can cause water electrolysis  
204 and the generation of  $H_2O_2$  and radicals, such as  $\bullet OH$  and  $\bullet O_2$ , which are harmful and have a  
205 detrimental effect on microbial metabolism, and ultimately inhibit microbial growth. Over-  
206 optimum currents can cause irreversible permeability of cell membranes and subsequently lead to  
207 leakage of essential cytoplasmic contents and lower cell respiration (Wei, et al. 2011). In the  
208 indirect oxidation process, strong oxidants, such as hypochlorite/ chlorine, ozone, and hydrogen

209 peroxide, are produced based on electrochemical reactions. All these oxidants are produced in situ  
210 and used immediately; they can also be harmful to the biofilm in higher concentrations (García-  
211 García et al., 2015). Zhang et al., (2011) studied BER, in which microorganisms specifically attach  
212 to the electrode as a biofilm and play a pivotal role in biodegradation. They reported that applying  
213 over-optimum currents causes separation and fall-off microorganisms from the electrode (Zhang  
214 et al., 2011).

### 215 *3.3. Effect of the initial concentration of COD*

216 Change in influent concentration is one of the factors affecting biological treatment processes.  
217 After determining the optimum current intensity, the efficacy of the bioreactor in the removal of  
218 influent was evaluated at different concentrations. Figure 3 shows the removal efficacy of BER at  
219 different COD concentrations. According to Figure 3, after increasing the COD concentration, the  
220 system efficacy decreased due to the shock caused by the applied concentration, and a few days  
221 later, the efficiency gradually increased and approached the previous state. When the system  
222 stabilized, the influent concentration was suddenly increased. When the reactor faced an organic  
223 shock loading, the biomass tried gradually be adapted to it to regenerate the system; however, at  
224 higher concentrations, the improvement remained incomplete as, under stable conditions and high  
225 concentrations, the removal efficiency decreased. The average removal efficiency of oil-  
226 contaminated wastewater by bioelectrochemical method at different initial concentrations is shown  
227 in Figure 3b. The maximum COD removal efficiency was obtained at 1500 mg/L. With further  
228 increase in the influent concentration, shock loadings imposed on the bioreactor increased so that  
229 the average removal efficiency decreased with concentration increase. The results showed that  
230 increased concentrations exceeded the maximum biodegradation capacity of microorganisms and  
231 played a limiting role in biomass. Therefore, to prevent biomass loss, continue the experiment, and

232 investigate the effect of retention time on process efficiency, the influent concentration returned  
233 to the baseline state (1500 mg/L). Different concentrations can directly affect the biomass activity  
234 and ohmic resistance of BES. The reason for a decrease in removal efficiency with an increase in  
235 influent concentration may be due to the point that higher oil-contaminated wastewater as substrate  
236 concentrations can act as a limiting factor on the biofilm (Mudliar et al., 2008). High  
237 concentrations create a critical state in microorganisms. With an increase in initial concentration,  
238 the time required to achieve the latest removal efficiency also increases. Therefore, with an  
239 increase in concentration, bacteria need more time for degradation. Wen et al. (2013), in a study  
240 on BES, reported that at the reaction time of 24 hours, under the same conditions, the removal  
241 efficiency of the system decreased from 82.3% to 50.3% with increasing the initial concentration  
242 of pollutants from 0.19 to 0.78 mM (Wena et al. 2013). Xuena et al., (2009) concluded that in  
243 BESs, increasing the concentration of influent can inhibit the growth of bacteria on biofilm (Xuena  
244 et al., 2009).

#### 245 *3.4. Effect of hydrolic retention time*

246 The reaction time is one of the major parameters in BESs. After examining different concentrations  
247 and applying organic shock loading, the COD concentration was set again at 1500 mg/L, and when  
248 the system stabilized, the effect of hydrolic retention time (HRT) changes on the optimal current  
249 intensity was investigated. At this stage, HRT was reduced step by step while the concentration  
250 was constant. As shown in Figure 4, HRT was studied at different reaction times of 1, 1.5, 2, 2.5,  
251 and 3 days. Changes in reaction time increased organic loads. As shown in Figure 4a, the efficiency  
252 decreased with decreasing reaction time from 3 to 2.5 days. However, the bioreactor efficiency  
253 improved somewhat over time, although it did not reach the initial state. As soon as the system  
254 stabilized, the retention time was reduced again from 2.5 to 2 days, in which an immediate decrease

255 was observed in efficiency, but improved over time, although it did not reach the initial state. The  
256 greater reduction in retention time, the greater reduction in efficiency. Figure 4b shows the average  
257 removal efficiency of COD by BES after reaching stable conditions at different time points. It can  
258 be concluded that time reduction adversely affects COD removal efficiency. The maximum  
259 efficiency was 84.2% in the best conditions in terms of reaction time. Organic loading increases  
260 with a reduction in HRT. Therefore, it can be concluded that the removal efficiency increases at  
261 higher HRTs, which is proportional to the lower load on the system. Investigation of residuals in  
262 the system showed that the residual COD increased with decreasing reaction time in BES. The  
263 reason for decreased efficiency versus reduced time is mainly related to reduced contact time  
264 between the substrate and biomass, which does not provide sufficient time for conveying the  
265 materials of liquid mass to biomass. As a result, the COD concentration increases in the effluent,  
266 and the removal efficiency decreases. In other words, a shorter contact time between biomass and  
267 oil-contaminated wastewater reduces biodegradation. In addition, a decrease in efficiency due to  
268 a reduction in retention time results from the fact that, despite a constant concentration of the  
269 substrate, microorganisms encounter an increase in organic load (Dareioti et al., 2014).  
270 Mohanakrishna et al., (2018) conducted a study on the treatment of oil-contaminated wastewater  
271 in an oil refinery by BES. They utilized a reactor with an effective volume of 1.13 L and two  
272 5×5×1-cm electrodes. Four different HRTs were considered in the study, and they concluded that  
273 the conditions for electrostimulation are more suitable at higher HRTs than shorter times in BESs.  
274 That is, the removal efficiency is higher at higher HRTs. They also found that the oil and grease  
275 removal efficiencies were significantly higher than that of COD, which could be due to the  
276 degradation of oil complex molecules into simpler ones still occurring as total COD (  
277 Mohanakrishna et al., 2018). Zhuang et al. (2014), in a study on biological treatment of wastewater,

278 concluded that as the retention time decreases, the efficiency of denitrification decreases, and the  
279 reason can be attributed to insufficient contact time between biomass and substrate, as well as the  
280 separation of biofilm from media and its wash-out (Zhuang et al., 2014). Guo et al. (2017), in a  
281 study on the effect of HRT on denitrification performance, indicated that the critical point of HRT  
282 effect on microbial process performance is the provision of contact time between biomass and  
283 substrate for microbial reaction. With an increase in retention time (within a particular range), the  
284 removal efficiency also increases. This increase in time leads to sufficient contact time between  
285 bacteria and wastewater. Sufficient contact time between the microbial population and substrate is  
286 not provided at shorter retention times for complete degradation, which leads to a reduction in  
287 removal efficiency. In addition, HRT can affect the secretion of extracellular polymeric substances  
288 (EPS) and the activity and accumulation of microorganisms. These materials also have a protective  
289 effect; they hold the biofilm-forming bacteria together and protect them against toxins and sudden  
290 increase in concentration to prevent them from wash-out (Guo, et al., 2017). HRT is a key  
291 parameter in BERs. Increased retention time persuades bacteria to excessively consume and  
292 degrade organic matter. It can also affect the type of microbial population. The longer retention  
293 time helps bacteria to more acclimatize to biodegradation-resistant and toxic substances. They can  
294 repair their enzyme system to acclimatize to these compounds or biodegrade them. Large bacterial  
295 populations may also develop enzyme systems suitable for degrading such organic matters.  
296 Therefore, researchers believe that higher HRTs are beneficial for the removal of poorly  
297 degradable or biodegradation-resistant compounds. In a bioreactor that refines poorly-degradable  
298 materials, a longer HRT can help to achieve high biodegradation efficiencies. Increased HRT gives  
299 more chance to substances to better contact microorganisms that increases the biodegradation rate.  
300 In lower HRTs, organic loading increases, leading to incomplete biodegradation so that the

301 degradation process remains incomplete. In addition to influencing the efficiency of the process,  
302 HRT affects reactor volume and manufacturing costs. Therefore, determining the optimum time  
303 for an acceptable and satisfactory efficiency is one of the main stages in the bioreactor design,  
304 considering the minimum bioreactor volume required (Shi et al., 2017). To prevent the loss of  
305 system biomass, resulting from shock loading, and improve the reactor performance, HRT  
306 increased at the initial stage (three days).

### 307 *3.5. Effect of supporting electrolyte*

308 Considering a relatively high solubility, availability, low cost, and less toxicity, NaCl was used in  
309 the present study as a supporting electrolyte. The effect of NaCl at different concentrations of 50,  
310 100, 150, and 200 mg/L was investigated, and the results are shown in Figure 5. According to  
311 Figure 5, by increasing NaCl concentration from 50 to 100 mg/L, the efficiency slightly increased  
312 and the trend continued by increasing to 150 mg/L. When the system stabilized, the NaCl  
313 concentration was increased to 200 mg/L, along which the efficiency decreased as the bioreactor  
314 was shocked. The removal efficiency somewhat improved by continuing the system exploitation  
315 in the next days but did not return to the initial state even after the system stabilization. According  
316 to the results, a threshold could be considered for the system biomass. As shown in Figure 5b, the  
317 highest removal efficiency was 86.7% at 150 mg/L. When concentration increases, the system  
318 bacteria are shocked, and the efficiency decreases.

319 Solution resistance (R) can be calculated using Ohm's law as Equation 10:

$$320 \quad \eta_{\text{ohm}} = IR \quad (10)$$

321 where  $\eta_{\text{ohm}}$  is ohmic overpotential (V), I current intensity (A), and R the local resistance of the  
322 electrochemical cell (ohm).



323 Also, the resistance of the solution can be calculated using Equation 11:

324 
$$R = \rho \frac{d}{A} \quad (11)$$

325 Accordingly, Equation 10 would be:

326 
$$\eta_{ohm} = \rho \frac{d}{A} I \quad (12)$$

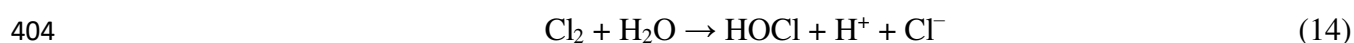
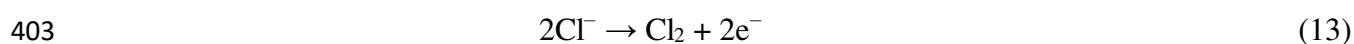
327 where  $\rho$  is the electrical resistivity ( $\Omega \text{ m}$ ) and the electrical conductivity ( $\sigma$ ) is its inverse,  $d$  the  
328 distance between the anode and the cathode (m), and  $A$  the surface area between the anode and the  
329 cathode ( $\text{m}^2$ ). One of the features of electrochemical and bioelectrochemical systems is that with  
330 an increase in the NaCl concentration and increase in the ionic conductivity of the solution, the  
331 ohmic resistance of the electrochemical cell reduces significantly, leading to improved efficiency.  
332 Considering equations 10 and 12, an increase in supporting electrolyte reduces energy  
333 consumption. Therefore, the presence of NaCl in the solution is economically advantageous over  
334 the electrochemical process and reduces costs. Increased NaCl concentration in the solutions of  
335 electrochemical and bioelectrochemical systems facilitates the electric current and reduces energy  
336 consumption. Likewise, many researchers suggested the addition of NaCl to increase electrical  
337 conductivity in such systems (Alam et al., 2016). Kokabian et al., (2013) stated that the effect of  
338 NaCl on the performance of anaerobic treatment systems depends on the nature of the microbial  
339 population. The study results showed that with increasing salinity, the removal efficiency of  
340 anaerobic bacteria increased initially but reduced with a further increase (exceeding the  
341 recommended threshold) due to its adverse effects on the system (Kokabian et al., 2013). Aslan et  
342 al. (2012), in a study entitled "The Influence of Salinity on Partial Nitrification in a Submerged  
343 Biofilter", concluded that with adding NaCl, the removal efficacy increased initially but reduced

344 with further increase. Different studies reported the stimulation of various bacterial species in low  
345 salinity. Although the low salinity of influent stimulates bacteria and increases their activity, higher  
346 concentrations act inversely. The effect of shock loading is evident at higher salt concentrations.  
347 High salinity causes plasmolysis and decreased bacterial activity. The susceptibility of  
348 microorganisms to salinity- e.g., bacterial tolerance to NaCl- is not the same, and some bacteria are  
349 less sensitive than others. Even laboratory conditions, such as pH, temperature, solid retention time  
350 (SRT), HRT, and suspended or attached growth system, can affect bacterial strains' susceptibility  
351 to salinity (Aslan et al., 2012). Lefebvre et al., (2012) concluded that salinity is generally useful  
352 for power production in the microbial fuel cell (MFC) process because an increase in ionic  
353 conductivity improves proton transfer and reduces the internal resistance of the system. However,  
354 excess salinity has inverse consequences on the physiology of the anaerobic microbial consortia  
355 (Lefebvre et al., 2012). Electrolyte and electrode ohmic losses are the two major types of ohmic  
356 resistance in BESs. The first one represents the voltage drop due to the movement of ions through  
357 weak electrolytes (usually low-conductivity wastewater), and the second refers to the movement  
358 of electrons through electrodes and wires connected to them. Many real wastewaters encounter  
359 significant ohmic losses since their conductivity is low. Therefore, in the treatment of wastewaters  
360 with ohmic resistance, approaches to overcome this dilemma should be considered. To overcome  
361 the ohmic resistance, first, it is recommended to reduce the distance between the electrodes as  
362 much as possible; however, it is impossible in most cases. Second, increase the conductivity or  
363 ionic strength by adding the required amounts of NaCl to the solution (Rozendal et al., 2008). Gui  
364 et al. (2017), in a study on the effect of NaCl on the denitrification process, indicated different  
365 effects of salinity across various concentrations and its inhibitory and toxic role in concentrations  
366 exceeding threshold limits. The toxicity of NaCl in low salinity is related to the hindrance of the

367 enzymatic activity of denitrifiers; in high salinity, however, in addition to the mentioned reason, it  
368 is also related to bacterial death (Gui et al., 2017). Ahmadi et al. (2017), in a study on the treatment  
369 of petrochemical wastewater by a salt-tolerant bacterial consortium, investigated the effect of  
370 salinity and determined the threshold under an HRT of three days and the initial COD  
371 concentration of 1240 mg/L. The results showed that salinity above the threshold caused a sharp  
372 and sudden decrease in COD removal efficiency, which is related to the loss and death of biomass  
373 due to the harmful effects of high salinity on the enzymatic activity of bacteria and their  
374 plasmolysis. At this time, the turbidity and TSS of the effluent increase significantly due to  
375 bacterial death and minimal biomass deposition (Ahmadi et al., 2017). The survival of the  
376 microbial population usually depends on adequate osmotic pressure in the environment. The  
377 concentration of the solution is correlated with the osmotic pressure so that if the mineral salt  
378 concentration is high in a solution, the osmotic pressure increases. Under isotonic conditions (e g,  
379 0.85 wt.% NaCl), microbial metabolism and growth are optimum. Since water molecules penetrate  
380 microorganisms, they may swell and even burst in pure or low salinity waters (e g, 0.01 wt.%  
381 NaCl). But bacterial plasmolysis occurs in environments with very high salinity (e g, 2 wt.% of  
382 NaCl) because water molecules diffuse to the outside, hindering microbial growth and even  
383 causing death. At this time, the concentration of suspended solids increases in the effluent due to  
384 microbial death. If salinity increases slowly in the system, the bacteria can reduce its adverse  
385 effects and acclimatize to the environment as much as possible. The reason may be that bacteria  
386 can regulate osmotic pressure by the efflux pump mechanism (e g, contractile vacuole) or synthesis  
387 of compatible salts when salinity increases. The higher salinity leads to more energy consumption  
388 by bacteria to maintain osmotic pressure. As a result, the energy accessible to microbial synthesis  
389 and function reduces. Microorganisms need organic matter for growth, the process performed

390 with the contribution of enzymes. Microbial enzymes are sensitive to toxins as they are typically  
391 proteins. If wastewaters contain some toxic substances, the enzymes are inactivated, and as a result,  
392 the removal efficiency of wastewater treatment reduces. The right amount of mineral salts  
393 improves microbial metabolism, but exerts toxic effects in excessive amounts, reduces the activity  
394 of enzymes, and destructs them. However, the bacteria probably can trigger a new enzyme system  
395 in the saline environment that helps to acclimatize to the new environment (He et al.,2017). Indirect  
396 oxidation at the anode is also one of the reactions happening during electrochemical and  
397 bioelectrochemical processes in the presence of chlorine ions. In indirect oxidation, strong  
398 oxidants, such as hypochlorite/chlorine, hydrogen peroxide, and ozone are generated by  
399 electrochemical reactions. All these oxidants in high concentrators are harmful to the biofilm.  
400 Indirect reactions of chlorine can be triggered by NaCl decomposition, based on the following  
401 equations, and its overproduction can cause microbial death.

402 These reactions occur when chlorine ions are the structural component of the salts used.



406 In addition to  $\text{ClO}^-$ , side reactions generate  $\text{ClO}_2^-$ ,  $\text{ClO}_3^-$ , and  $\text{ClO}_4^-$  through anodic oxidation,  
407 which can be hazardous to bacteria in high concentrations (Bassyouni et al., 2017).

### 408 *3-6- Evaluation of bacterial community*

409 Today, various methods are employed to identify biofilms, one of which is microscopic  
410 examination. In this method, magnified images are taken from the specimen. The SEM was utilized

411 in the present study to investigate the bacterial community of the biofilm. According to Figure 6,  
412 the predominant bacteria in the bioreactor were rod-shaped (bacilli) and cocci. The biofilm  
413 includes a bacterial consortium and is composed of a dense bacterial population. In terms of  
414 structure, the biofilm has channels allowing the transfer of substrates to the inner part for bacterial  
415 accessibility. The results of the present study were consistent with those of Luo et al. (2005). It was  
416 reported that at the optimum current intensity, the biofilm-holding bacteria were rod-shaped and  
417 cocci, which their natural shape might be changed at different current intensities (Luo et al., 2005).  
418 Figure 6 illustrates the presence of EPS secretions between bacteria. A biofilm is a community of  
419 bacteria attached to a surface and covered by EPSs. Huang et al., (2013) concluded in a study that  
420 an increase in electric current can trigger EPS efflux, which ultimately leads to the better formation  
421 of biofilm in BES. However, the microbial activity varies across electric currents. The biofilm  
422 formation in a BES is of great importance for the removal of pollutants or the production of electric  
423 current. They found that at optimum current intensities, the biofilm formation is promoted and  
424 improves. However, when the current intensity exceeds the optimum level affects the biofilm  
425 adversely and diminishes its formation. They also concluded that the biofilm in BESs nurtured  
426 with organic matter is better formed near the anode than the cathode. They reported that EPSs are  
427 essential for biofilm formation. Their structural composition depends on the growing conditions  
428 of the biofilm. EPSs are composed of polysaccharides, proteins, nucleic acids, and DNAs;  
429 however, they are mostly constructed from proteins and carbohydrates. Studies on the composition  
430 of EPSs led to a variety of results. Some studies concluded that polysaccharides predominate in  
431 the EPS layer of biofilms, while others reported proteins as the main constituent. These  
432 contradictory findings indicate that the composition of biofilms depends on growth conditions. In  
433 the study by Huang, with excessing the optimum electric current, the EPS also increased and

434 decreased with a further increase, which disrupted the formation of biofilms and inhibited  
435 microbial growth (Huang et al., 2013). Some cavities and channels seen in Figure 6 are related to  
436 the inlet of nutrients and outlet of substances generated by bacteria; they allow substrates to  
437 transfer through the inner part of the biofilm (Muda et al., 2010). The results of Gram staining are  
438 shown in Figure 6b. According to Figure 6b, the effluent was treated with a consortium, including  
439 Gram-positive and -negative bacteria. Similarly, Poh et al. (2010), in a study on the treatment of  
440 palm oil mill effluent by an anaerobic method, reported that most bacteria detected in the  
441 consortium were rod-shaped and cocci, and Gram staining proved that both Gram-positive and -  
442 negative species were involved in the process (Poh et al., 2010).

### 443 *3-7- The effect of electric current*

444 An experiment was performed to evaluate the effect of induction and electrostimulation on the  
445 removal efficiency of COD. It was performed under the same conditions. Accordingly, BES was  
446 compared with a similar bioreactor in the absence of DC (Figure 7). The removal efficiency at the  
447 biological state without DC was 65.9%, but it was 86.7% at the BES state. Based on the obtained  
448 results, the electric current could positively affect BES, leading to higher removal efficiency than  
449 other reactors since it stimulates bacterial growth. In the electrostimulation or biostimulation  
450 process, enzymatic activity, cellular biopolymers synthesis, membrane transfers, and reproduction  
451 are influenced by the process and can improve the removal efficiency. Cardenas-Robles et al.  
452 (2013), in a study on azo dye degradation by BES, concluded that the application of  
453 electrostimulation increases the dye removal efficiency. The results also showed that  
454 electrostimulation, in addition to an increase in efficiency, reduces the time required for the  
455 complete removal of pollutants. This reduction in time also decreases the reactor volume, which  
456 leads to the lower design and implementation costs (Cardenas-Robles et al., 2013).

457 Zhang et al. (2011), in a study on the degradation of 2,4-dichlorophenol by BES in an anaerobic  
458 process, reported good results and high efficiency. They compared the removal efficiency of BES  
459 and biological (in the absence of electric current) and electrochemistry methods and showed that  
460 the bioelectrode process had higher efficiency than the other two methods, and the removal  
461 efficiency of pollutants was lower in pure biological and pure electrochemical processes. The  
462 removal efficiency of the investigated processes was 100%, 42%, and 61%, respectively. This  
463 improvement and increase in removal efficiency were due to the electrostimulation of bacteria  
464 (Zhang et al., 2011).

### 465 *3.8. Kinetic study*

#### 466 *3.8.1. First-order kinetic model*

467 The steady-state data of each stage were used to determine the kinetic coefficients. As shown in  
468 Figure 10a and Table 1, the first-order kinetic constant ( $K_1$ ) was 0.485 (1/d). Also, the correlation  
469 coefficient ( $R^2$ ) was 0.85.

#### 470 *3.8.2. Second-order kinetic model (Grau model)*

471 The  $R^2$  for the second-order kinetic model was 0.961, indicating that the COD removal process  
472 could also follow the second-order model (Figure 8). The kinetic constants of a and b were 1.338  
473 and  $0.694 \text{ day}^{-1}$ , respectively. Besides, increasing a or b parameters had a direct and adverse effect  
474 on efficiency, the removal efficiency increased in the system.

#### 475 *3.8.3. Modified Stover-Kincannon model*

476 In the present study,  $U_{\max}$  was 1.106 and  $K_B$  0.767 g/ L.d. Both  $K_B$  and  $U_{\max}$  play a pivotal role in  
477 determining the volume of a bioreactor. The results showed that among the three proposed models,

478 the R<sup>2</sup> of the modified Stover-Kincannon model was higher (0.975), indicating that the model was  
479 more fitted with bioreactor performance and the COD removal reaction was more consistent with  
480 it. Therefore, the modified Stover-Kincannon model can be used for the accurate prediction of the  
481 removal of biodegradable organic matter.

### 482 3-9- Energy consumption

483 Cost is one of the most important factors in choosing water and wastewater treatment methods.  
484 The consumed energy plays a particular role in the costs of electrochemical processes. Therefore,  
485 energy consumption was calculated using the following equation:

$$486 \quad E \left( \frac{\text{Kwh}}{\text{m}^3} \right) = \frac{U \times I \times t}{V} \quad (16)$$

487 where U is the applied voltage (V), I the current intensity (A), t the reaction time (h), and V the  
488 volume of the effluent (L). According to the applied optimum current intensity (15 mA) and  
489 reaction time (three days), the amount of energy consumed by BES was 1.9145 kWh/m<sup>3</sup>. The  
490 electrochemical process was used to obtain the same removal efficiency, which according to the  
491 applied current intensity (1 A) and reaction time (2 hours), the energy consumed by the  
492 electrochemical process was 45.33 kWh/m<sup>3</sup>. The comparison of the above results indicated that  
493 the cost of energy consumed by BES was much lower than that of the electrochemical system. In  
494 addition, the cost of electrodes was another factor affecting the cost, which considering higher  
495 current intensity and voltage in the electrochemical method, corrosion and consumption rate of  
496 electrodes was higher in this method than BES. Jinyou Shen et al., (2012) reported similar results  
497 in a study on BES. They concluded that the cost of energy consumed by the BES was much lower  
498 than that of the electrochemical one. Excessive energy consumption, as an important drawback,  
499 limits the use of the electrochemical system on a large scale. BESs can be used to reduce energy



500 consumption in the electrochemical method, which significantly reduces energy consumption in  
501 the system compared to other ones (Shen et al., 2012). Wen et al., (2013) investigated the  
502 degradation of 4-chlorophenol by BES in an anaerobic process. They estimated that the energy  
503 consumed by BES was 5 to 30 times lower than that of the electrochemical one. They concluded  
504 that this technology would be a method of choice for the removal of many pollutants due to its  
505 higher efficiency and lower energy consumption (Wena et al., 2013).

#### 506 **4. Conclusions**

507 In the present study, BERs were used to treat oil-contaminated wastewater. The results showed  
508 that this process could be very advantageous for the treatment of such wastewaters. The study also  
509 confirmed that BERs have higher efficiencies than pure electrochemical ones under the same  
510 conditions, indicating that electrostimulation under optimal conditions can increase removal  
511 efficiency. However, if the current intensity exceeds optimal levels, the biofilm biomass is lost,  
512 and thus the removal efficiency decreases. The results showed that the removal of biodegradable  
513 organic matter could accurately be predicted by the modified Stover–Kincannon model. Based on  
514 the findings, the cost of energy consumed by BERs was much lower than that of the  
515 electrochemical one. The study also indicated that BERs are a promising method with good  
516 prospects for the treatment of oil-contaminated wastewaters.

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518 paper; A. Rezaee designed experiments and supervised the research.

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520 **Data availability:** All data generated or analyzed during this study are included in this published  
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522 **Compliance with ethical standards:** Competing interests The authors declare that they have no  
523 conflict of interest.

524 **Ethics approval and consent to participate:** This research did not involve any human  
525 participants and/or animals.

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637

638

### Figure and Table captions

639 **Fig. 1.** Adaptation and acclimation period of biomass (pH 7.0 and induced current = 1 mA).

640 **Fig. 2.** (a) Effect applied current on removal efficiency and (b) COD removal efficiencies under  
641 different applied currents (COD = 1500 mg/L; pH 7.0; reaction time = 3 day).

642 **Fig. 3.** (a) The effect of changes in the initial concentration of COD on the removal efficiency by  
643 the bioelectrochemical process; (b) the removal efficiency of COD at different initial  
644 concentrations (applied current intensity of 15 mA at ambient temperature).

645 **Fig. 4.** (a) The effect of HRT on removal efficiency of the bioelectrochemical process; (b) COD  
646 removal efficiency at different reaction times (COD = 1500 mg/L and applied current intensity =  
647 15 mA).

648 **Fig. 5.** (a) Effect of different concentrations of NaCl on removal efficiency; (b) COD removal  
649 efficiency at different NaCl concentrations (COD = 1500 mg / L and applied current intensity =  
650 15 mA).

651 **Fig. 6.** a) Scanning electron microscopy photographs of bacteria in the biofilm, b) image of Gram-  
652 stained bacteria.

653 **Fig. 7.** Comparison of COD removal efficiency with and without electric field.

654 **Fig. 8.** Kinetic plots of the COD removal through the BER process: (a) the first-order model; (b)  
655 the second-order (Grau) model; (c) the modified Stover-Kincannon model.

656 **Table 1.** Summary of Kinetic Parameters Based on the First-order, Second-order, and Stover-  
657 Kincannon Models.

658



# Figures

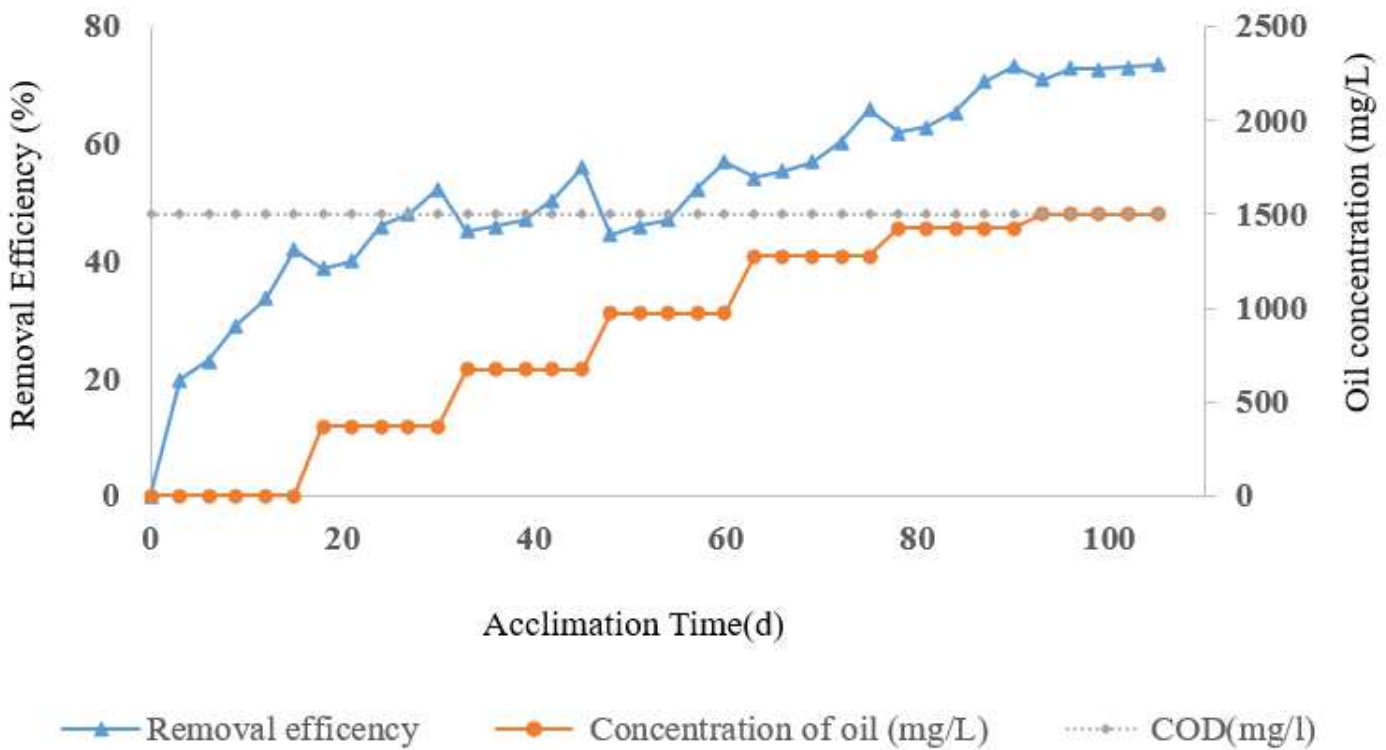


Figure 1

Adaptation and acclimation period of biomass (pH 7.0 and induced current = 1 mA).

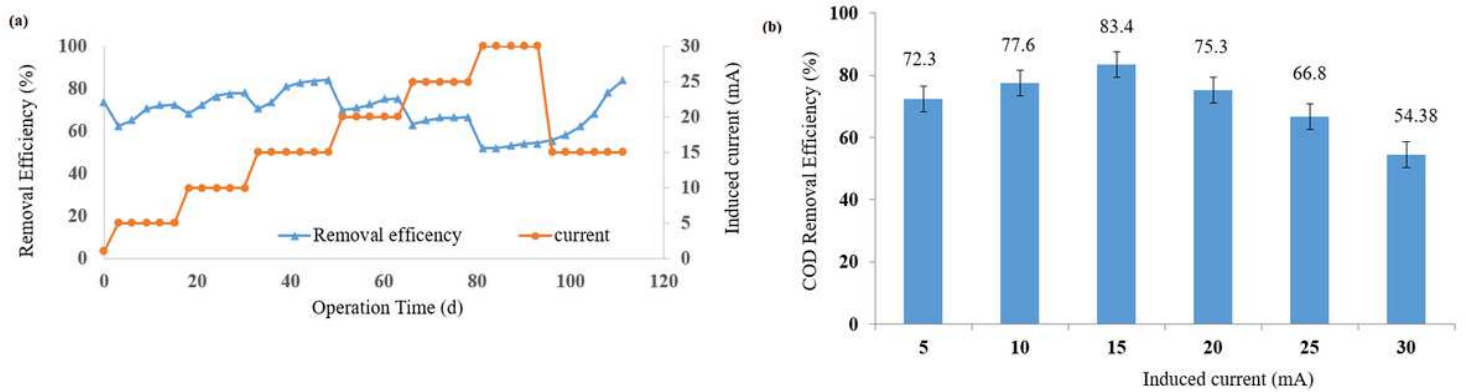
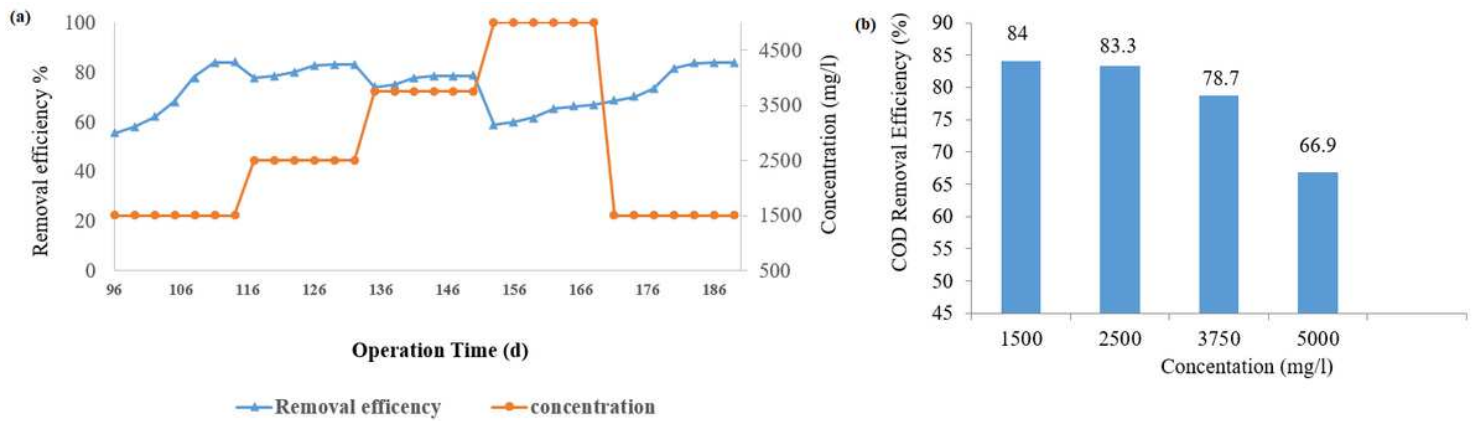


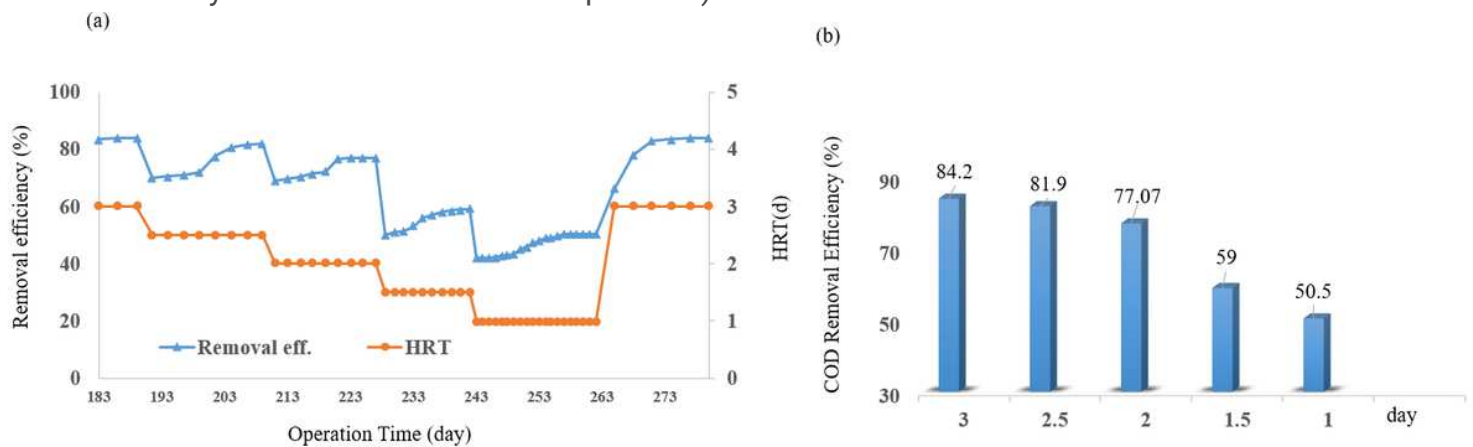
Figure 2

(a) Effect applied current on removal efficiency and (b) COD removal efficiencies under different applied currents (COD = 1500 mg/L; pH 7.0; reaction time = 3 day).



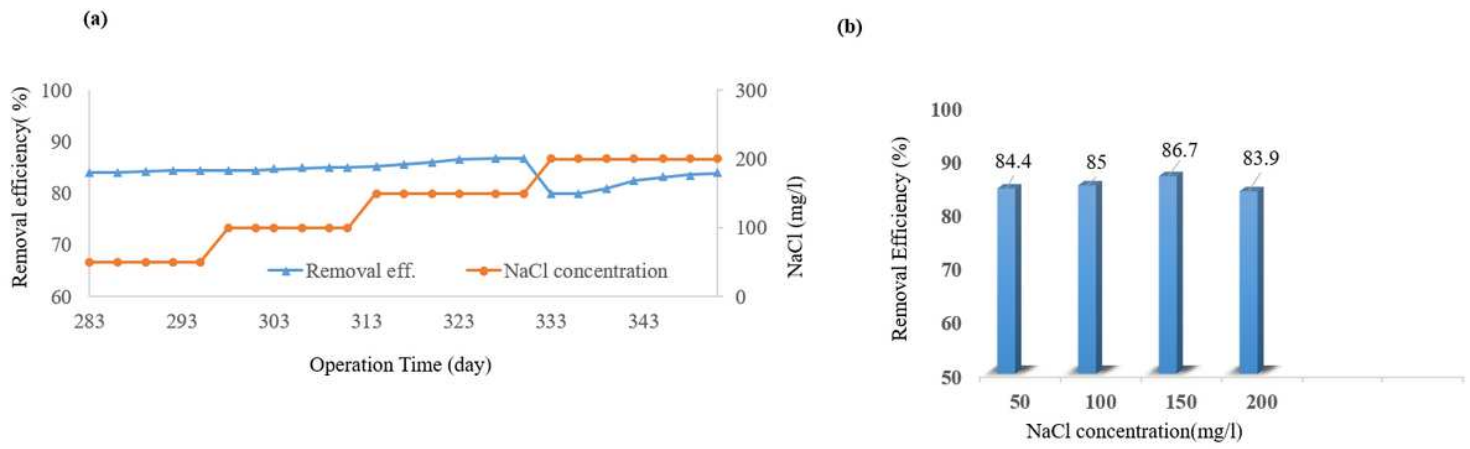
**Figure 3**

(a) The effect of changes in the initial concentration of COD on the removal efficiency by the bioelectrochemical process; (b) the removal efficiency of COD at different initial concentrations (applied current intensity of 15 mA at ambient temperature).



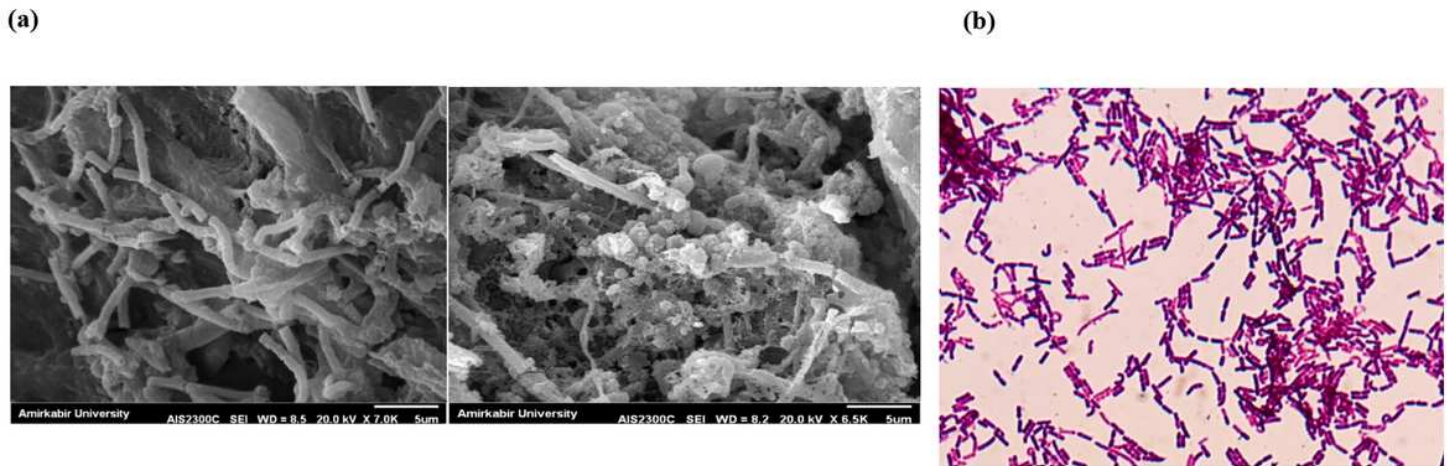
**Figure 4**

(a) The effect of HRT on removal efficiency of the bioelectrochemical process; (b) COD removal efficiency at different reaction times (COD = 1500 mg/L and applied current intensity = 15 mA).



**Figure 5**

(a) Effect of different concentrations of NaCl on removal efficiency; (b) COD removal efficiency at different NaCl concentrations (COD = 1500 mg / L and applied current intensity = 15 mA).



**Figure 6**

a) Scanning electron microscopy photographs of bacteria in the biofilm, b) image of Gramstained bacteria.

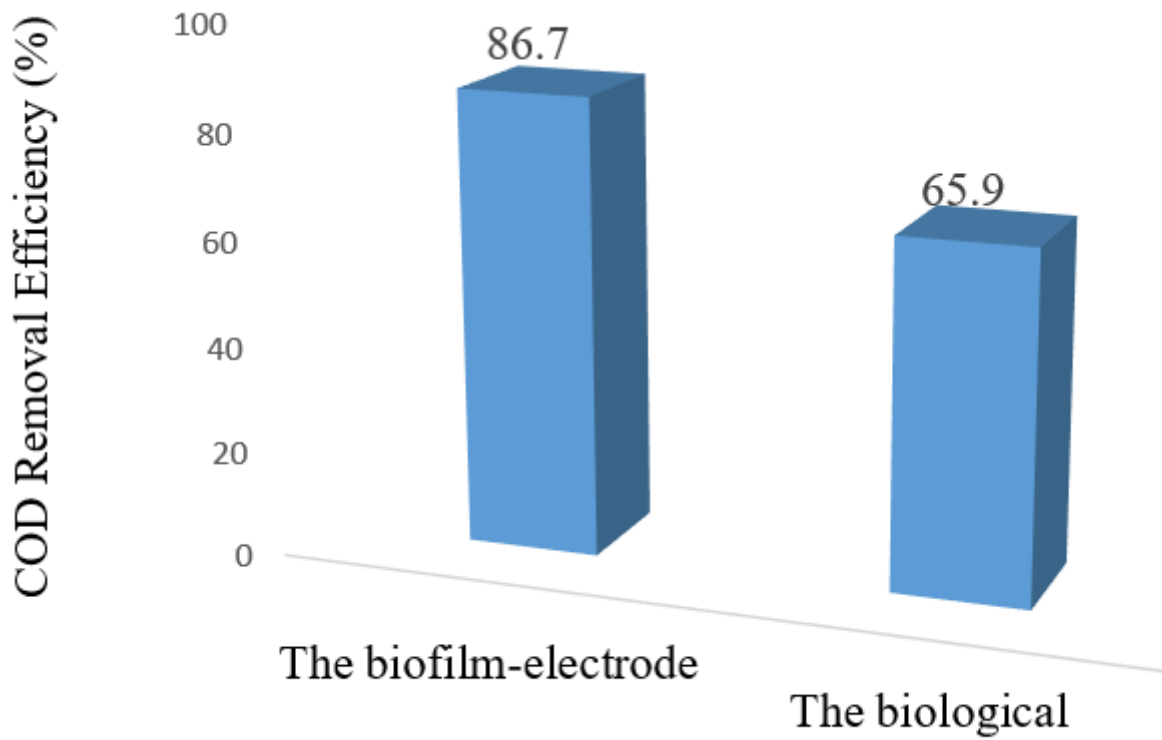


Figure 7

Comparison of COD removal efficiency with and without electric field.

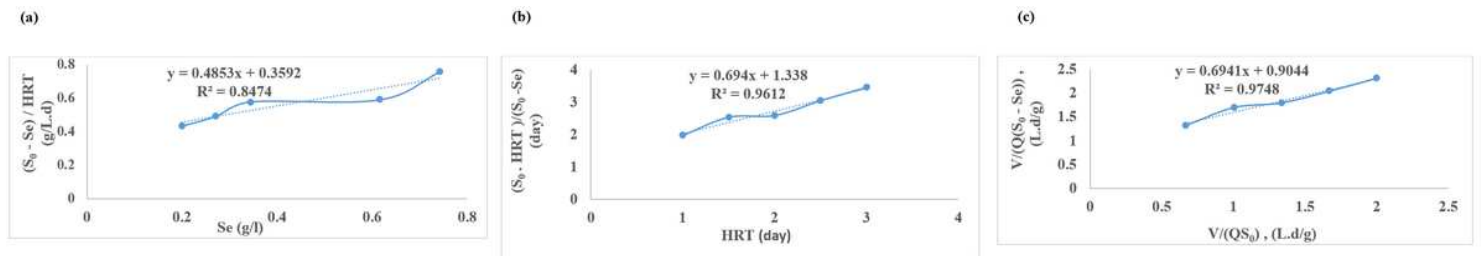


Figure 8

Kinetic plots of the COD removal through the BER process: (a) the first-order model; (b) the second-order (Grau) model; (c) the modified Stover-Kincannon model.