Contribution to The Peripheral Vasculopathy and Endothelial Cell Dysfunction by CXCL4 in Systemic Sclerosis

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Research article

Keywords: Systemic sclerosis (SSc), the very early diagnosis of SSc (VEDOSS), vasculopathy, digital ulcer, CXCL4, Fli-1

Posted Date: July 13th, 2020

DOI: https://doi.org/10.21203/rs.3.rs-41064/v1

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Version of Record: A version of this preprint was published at Journal of Dermatological Science on July 1st, 2021. See the published version at https://doi.org/10.1016/j.jdermsci.2021.07.006.
Abstract

**Objective** CXCL4, a chemokine with antiangiogenic property, is reported to be involved in systemic sclerosis (SSc) related pulmonary arterial hypertension (PAH). We investigated the contribution of CXCL4 to SSc development by focusing on the correlation of circulatory CXCL4 levels with their peripheral vasculopathy, as well as the effect of CXCL4 on endothelial cell dysfunction and angiogenesis disturbance in SSc and the potential signaling.

**Methods** We measured the serum CXCL4 levels in 58 patients with SSc, 10 patients with the very early diagnosis of SSc (VEDOSS), and 80 healthy controls. Then, CXCL4 levels were correlated with their clinical features, especially the peripheral vasculopathy. These observations were further validated in an additional cohort including 50 SSc patients, 12 VEDOSS patients, and 80 healthy controls. Moreover, we studied the anti-angiogenesis effects and the underlying signaling of CXCL4 in human umbilical vein endothelial cells (HUVECs) *in vitro*.

**Results** Circulating levels of the CXCL4 were 103.62% higher in patients with SSc and 201.51 % higher in patients with VEDOSS than matched HCs, and these observations were confirmed in two independent cohorts. CXCL4 levels were closely associated with digital ulcers (DU) and nailfold video capillaroscopy (NVC) abnormalities in SSc. The proliferation, migration, and tube formation of HUVECs were significantly inhibited by recombinant human CXCL4 or SSc derived serum, which reversed by CXCL4 neutralizing antibody, but not CXCR3 inhibitor. CXCL4 downregulated the transcription factor Friend leukaemia integration factor-1 (Fli-1) via c-Abl signaling. Furthermore, CXCL4 blocked the transforming growth factor (TGF) -β or platelet-derived growth factor (PDGF) induced cell proliferation of HUVECs.

**Conclusions** CXCL4 may contribute to peripheral vasculopathy in SSc by downregulating Fli-1 via c-Abl signaling in endothelial cells and interfering angiogenesis.

Introduction

Systemic sclerosis (SSc) is an autoimmune disease with skin and multiple viscera involvement, which characterized by a complex interplay of vasculopathy, immune system activation, and persistent tissue fibrosis[1]. Progressive and marked reduction in capillaries is a hallmark finding in early stage SSc with avascularity increasing with disease progression. Patients with SSc may develop a spectrum of vascular disease, mainly including Reynaud’s phenomenon (RP), digital ulcer (DU), pulmonary arterial hypertension (PAH), and scleroderma renal crisis (SRC). In SSc patients, the universal prevalence of RP demonstrated that increased vascular dysfunction exists in the process of SSc[2]. As the first onset symptom, RP leads to continuous digital ischemia, which may develop into digital ulcers (DU) or severe digital ischemia with gangrene in some extreme cases[3]. These early events induce increasing vascular tone, decreasing capillary blood flow, and chronic tissue hypoxia, ultimately leading to the extracellular matrix (ECM)[4] accumulation and tissue fibrosis. Therefore, it is necessary to identify biomarkers that could predict the very early SSc, before inevitable organ injury and fibrosis occurred.
The alteration of both pro-angiogenic mediators and inhibitors of angiogenesis have been recently implicated in vascular dysfunction in SSc[5]. In addition, impairment of angiogenic signal transduction pathways in endothelial cells contributed to disturbing angiogenesis in SSc[6]. Previous studies have focused on CXCL4 and its anti-angiogenic effects in the pathogenesis of atherosclerosis[7], and cancer[8]. Proteome-wide analysis revealed CXCL4 as a biomarker in SSc, and subsequently closely correlated with interstitial lung disease (ILD) and PAH[9]. CXCL4, unlike other chemokines that bind to their specific receptors, exerts its effects through its high affinity for proteoglycans and other negatively charged molecules. It was also reported that CXCL4 can directly bind to the CXCR3B chemokine receptor isoform or lipoprotein-related protein-1(LRP1)[11]. In patients with SSc, CXCL4 binds DNA into specific immune complexes that magnify plasmacytoid dendritic cell (pDC) -hyperactivation and produce inflammatory cytokines, which independent of CXCR3[12]. These data suggest the involvement of CXCL4 in SSc pathology.

Therefore, the aims of this study were to investigate whether the levels of CXCL4 could be altered in the very early diagnosis of SSc (VEDOSS), which manifesting typical vasculopathy but not fibrosis yet. Then, the association between the levels of CXCL4 and the peripheral vasculopathy in SSc evaluated clinically or by nailfold videocapillaroscopy (NVC) was also studied. Furthermore, we explored the potential signaling mechanism mediated by CXCL4, which could be functionally contribute to the defective angiogenic process characteristic of this disease spectrum.

**Method**

**Patients, controls and serum samples.**

In the discovery cohort, serum samples from 58 patients with systemic sclerosis (SSc)[13], 10 patients with VEDOSS[14] were collected between Jun 2016 to Jun 2018 at Huashan Hospital, Fudan University, Shanghai, China. Here, VEDOSS patients presented with RP, puffy fingers, and positivity of antinuclear antibodies, together with SSc specific antibodies and/or pathognomonic microvascular alteration at capillaroscopy, meanwhile not fulfilling the 2013 American College of Rheumatology (ACR)/ European League Against Rheumatism (EULAR) classification criteria for SSc (i.e. total score < 9)[15]. The modified Rodnan skin score (mRSS) of all included patients were assessed by the same rheumatologist (ML) at all time points during the study period. The disease duration of SSc patients was calculated from the first non-RP symptom. Detailed definitions of the cutaneous subsets, clinical features, organ involvements, nailfold capillaroscopy patterns, and immunological features have been shown in Table 1. Eighty healthy controls (HCs) matched for sex and age were also included as a comparison group. The presence of primary RP was an exclusion criterion for healthy controls. An age-matched and sex-matched replication cohort consisting of 50 SSc patients, 12 VEDOSS patients, and 80 controls, was assessed between Jul 2018 to Jun 2019. The demographic and clinical characteristics of patients included in the replication cohort are summarized in Table 2.
<table>
<thead>
<tr>
<th>Characteristics</th>
<th>SSc, n = 58</th>
<th>VEDOSS, n = 10</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years, mean ± SD (range)</td>
<td>58.00 ± 13.23, 2–72</td>
<td>55.70 ± 6.80, 46–65</td>
</tr>
<tr>
<td>Sex</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Male</td>
<td>14 (24.14)</td>
<td>1 (10)</td>
</tr>
<tr>
<td>Female</td>
<td>44 (75.86)</td>
<td>9 (90)</td>
</tr>
<tr>
<td>Disease subset</td>
<td></td>
<td></td>
</tr>
<tr>
<td>dcSSc</td>
<td>24 (41.38)</td>
<td>-</td>
</tr>
<tr>
<td>lcSSc</td>
<td>21 (36.21)</td>
<td>-</td>
</tr>
<tr>
<td>Overlap</td>
<td>10 (17.24)</td>
<td>-</td>
</tr>
<tr>
<td>SSc sine Scleroderma</td>
<td>3 (5.17)</td>
<td>-</td>
</tr>
<tr>
<td>Disease duration&lt;sup&gt;a&lt;/sup&gt;, years, mean ± SD (range)</td>
<td>3.61 ± 3.77, 0.30–18.00</td>
<td>1.51 ± 1.77, 0–5</td>
</tr>
<tr>
<td>Autoantibody positivity</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ANA</td>
<td>55 (94.83)</td>
<td>10 (100)</td>
</tr>
<tr>
<td>ACA</td>
<td>11 (18.97)</td>
<td>8 (80)</td>
</tr>
<tr>
<td>ATA</td>
<td>28 (48.28)</td>
<td>1 (10)</td>
</tr>
<tr>
<td>Interstitial lung disease (ILD)</td>
<td>38 (65.52)</td>
<td>2 (20)</td>
</tr>
<tr>
<td>Digital ulcers</td>
<td>18 (31.03)</td>
<td>0</td>
</tr>
<tr>
<td>mRSS&lt;sup&gt;b&lt;/sup&gt;, mean ± SD (range)</td>
<td>10.19 ± 10.64, 0–38</td>
<td>0.10 ± 0.32, 0–1</td>
</tr>
</tbody>
</table>

Except where indicated otherwise, values are the number (%) of subjects. <sup>a</sup>Disease duration was calculated since the first non-Raynaud's symptom of SSc; <sup>b</sup>modifed Rodnan skin thickness score. ACA, anticentromere antibodies; ANA, antinuclear antibodies; ATA, anti-topoisomerase I antibodies; dcSSc, diffuse cutaneous SSc; lcSSc, limited cutaneous SSc; SSc, systemic sclerosis; VEDOSS, very early diagnosis of SSc.
Table 2
Demographic and clinical characteristics of the patients with SSc and VEDOSS in the replication cohort.

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>SSc, n = 50</th>
<th>VEDOSS, n = 12</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, years, mean ± SD (range)</td>
<td>54.18 ± 13.26, 25–78</td>
<td>57.58 ± 7.19, 47–68</td>
</tr>
<tr>
<td>Sex</td>
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<td></td>
</tr>
<tr>
<td>Male</td>
<td>13 (26)</td>
<td>3 (25)</td>
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<tr>
<td>Female</td>
<td>37 (74)</td>
<td>9 (75)</td>
</tr>
<tr>
<td>Disease subset</td>
<td></td>
<td></td>
</tr>
<tr>
<td>dcSSc</td>
<td>28 (56)</td>
<td>-</td>
</tr>
<tr>
<td>IcSSc</td>
<td>18 (36)</td>
<td>-</td>
</tr>
<tr>
<td>Overlap</td>
<td>3 (6)</td>
<td>-</td>
</tr>
<tr>
<td>SSc sine Scleroderma</td>
<td>1 (2)</td>
<td>-</td>
</tr>
<tr>
<td>Disease duration&lt;sup&gt;a&lt;/sup&gt;, years, mean ± SD (range)</td>
<td>3.88 ± 5.31, 0–28</td>
<td>1.29 ± 1.39, 0–4</td>
</tr>
<tr>
<td>Autoantibody positivity</td>
<td></td>
<td></td>
</tr>
<tr>
<td>ANA</td>
<td>48 (96.00)</td>
<td>12 (100)</td>
</tr>
<tr>
<td>ACA</td>
<td>6 (12)</td>
<td>8 (66.67)</td>
</tr>
<tr>
<td>ATA</td>
<td>21 (42)</td>
<td>2 (16.67)</td>
</tr>
<tr>
<td>Interstitial lung disease (ILD)</td>
<td>29 (58)</td>
<td>1 (8.3)</td>
</tr>
<tr>
<td>Digital ulcers</td>
<td>23 (46)</td>
<td>0</td>
</tr>
<tr>
<td>mRSS&lt;sup&gt;b&lt;/sup&gt;, mean ± SD (range)</td>
<td>10.48 ± 8.34, 0–31</td>
<td>0.10 ± 0.32, 0–1</td>
</tr>
</tbody>
</table>

Except where indicated otherwise, values are the number (%) of subjects. <sup>a</sup>Disease duration was calculated since the first non-Raynaud’s symptom of SSc; <sup>b</sup>modified Rodnan skin thickness score. ACA, anticentromere antibodies; ANA, antinuclear antibodies; ATA, anti-topoisomerase I antibodies; dcSSc, diffuse cutaneous SSc; IcSSc, limited cutaneous SSc; SSc, systemic sclerosis; VEDOSS, very early diagnosis of SSc.
Table 3
RT-qPCR and primers.

<table>
<thead>
<tr>
<th>Primer</th>
<th>Sequence</th>
</tr>
</thead>
<tbody>
<tr>
<td>ET-1 –F</td>
<td>AGAGTGTGTCTACTTCTGCCA</td>
</tr>
<tr>
<td>ET-1 –R</td>
<td>CTTCCAAGTCATACGGGAAACA</td>
</tr>
<tr>
<td>Fli-1 –F</td>
<td>CCAACGAGAGGAGGATCATCG</td>
</tr>
<tr>
<td>Fli-1 –R</td>
<td>TTCCGTGTGGTAGGGGTGT</td>
</tr>
<tr>
<td>IL-6 -F</td>
<td>ACTCACCCTCTTCAAAGCAAATG</td>
</tr>
<tr>
<td>IL-6 -R</td>
<td>CCATCTTTGGAAAGGTTCAGGTT</td>
</tr>
<tr>
<td>S100A8 -F</td>
<td>TTTCCATGCGCTCTACAG</td>
</tr>
<tr>
<td>S100A8 -R</td>
<td>ACGCCCATCTTTATCACC</td>
</tr>
<tr>
<td>MMP7 -F</td>
<td>CATGAGTGAGCTACAGTGGGA</td>
</tr>
<tr>
<td>MMP7 -R</td>
<td>CTATGACGCGGGAGTTTAACAT</td>
</tr>
<tr>
<td>TNF-α -F</td>
<td>CCTCTCTCTAATCMGCCTCTG</td>
</tr>
<tr>
<td>TNF-α -R</td>
<td>GAGGACCTGGGGATAGATGAG</td>
</tr>
<tr>
<td>AT1R -F</td>
<td>ATTTAGCACTGGCTGACTTATGC</td>
</tr>
<tr>
<td>AT1R -R</td>
<td>CAGCGGTATTCCATAGCTGTG</td>
</tr>
<tr>
<td>ETAR -F</td>
<td>TCGGGTTCTATTTCTGTATGCC</td>
</tr>
<tr>
<td>ETAR -R</td>
<td>TGTTTTTGCACTTTCTCGACG</td>
</tr>
<tr>
<td>GAPDH –F</td>
<td>GGAGCGAGATCCCTCCAAAAT</td>
</tr>
<tr>
<td>GAPDH –R</td>
<td>GGCTGTGCTCATACTTCTCATG</td>
</tr>
</tbody>
</table>

Serum samples were obtained excluding from the participants who were given glucocorticoid, nonsteroidal anti-inflammatory drugs (NSAIDs), or systemic immunosuppressants to avoid confounding effects of these medications on clinical parameters and CXCL4 expression. Informed consent was obtained from each patient. This study was approved by the Ethics Committee of Huashan Hospital, Fudan University. Serum was collected, centrifuged, aliquoted, and stored at −80 °C. The serum samples had undergone only 1 freeze/thaw cycles before protein measurements. All samples were measured in duplicates, in accordance with the approved guidelines and regulations.

**Statistical analysis**

GraphPad Prism 5 was used to conduct statistical analysis. Normally distributed data are presented as mean ± SD, unless otherwise indicated. Inter-group differences were assessed for significance using an
The circulating levels of the CXCL4 are raised in the patients with SSc and the patients with a very early diagnosis of SSc (VEDOSS).

We first examined serum CXCL4 in SSc and VEDOSS patients in the identification cohort, and evaluated the possible correlation with specific clinical features of the disease. A total of 58 patients with SSc, 10 patients not fulfilling the 2013 ACR/EULAR classification criteria for SSc (total score < 9) and identified as VEDOSS (presented with Raynaud’s phenomenon, puffy figures, and positivity of antinuclear antibodies, together with SSc specific antibodies and/or pathognomonic microvascular alteration at capillaroscopy), and 80 healthy controls (HC) were recruited and stratified as described in Methods. In this identification cohort, serum CXCL4 levels were 60.59% higher in patients with SSc and 132.95% higher in patients with VEDOSS than matched HC (HC: 1751 ± 917.1; SSc: 2812 ± 1445; VEDOSS: 4079 ± 1978 pg/ml; p < 0.0001 compared with HC respectively; Fig. 1A). Notably, significantly higher serum CXCL4 levels were detected in the patients with VEDOSS than the patients with SSc (p = 0.0089, Fig. 1A). These results were subsequently validated in the replication cohort, where circulating CXCL4 resulted significantly increased in SSc (n=50) and in VEDOSS (n=12) serum compared with control serum (n=80) (HC: 1288 ± 839.2; SSc: 3423 ± 1617; VEDOSS: 5003 ± 1814 pg/ml; p < 0.0001 compared with HC respectively; Fig. 1B). Similarly, further elevation of CXCL4 was detected in the patients with VEDOSS compared to the patients with SSc (p = 0.0004, Fig. 1B).

We performed the pooled analysis by combining patients and controls from the identification and replication cohorts. Circulating CXCL4 levels were 103.62% higher in patients with SSc and 201.51% higher in the patients with VEDOSS than in healthy controls, also significantly higher in SSc patients relative to VEDOSS patients (HC: 1520 ± 906.4; SSc: 3095 ± 1551; VEDOSS: 4583 ± 1903 pg/ml; p < 0.0001 compared with each other; Fig. 1C).

Moreover, we found that CXCL4 levels gradually increased per group in the following order: patients with SSc sine scleroderma (ssSSc) (n=4), overlapping syndrome of SSc (n=13), limited cutaneous SSc (lcSSc) (n=39), diffuse cutaneous SSc (dcSSc) (n=52), and those with very early systemic sclerosis (VEDOSS) (n=22) (Fig. 1D). Additionally, the level of CXCL4 was remarkably increased in VEDOSS patients rather than those in the subsets of ssSSc, overlapping, lcSSc, or dcSSc (Fig. 1D).
Increased serum CXCL4 levels positively correlate to the much severer skin fibrosis and peripheral vasculopathy in SSc patients

We next assessed the association between CXCL4 levels and the clinical phenotype in the combined cohort, and the patients with SSc were stratified as described in Methods. The levels of CXCL4 in SSc were positively correlated with their mRSS score (Fig. 2A), demonstrating that the increased CXCL4 level may implicate much severer skin fibrosis. Notably, significantly higher serum CXCL4 levels were detected in the SSc patients with digital ulcer (DU) (n=41) than those without DU (n=67) (3686±1769 vs 2734±1285 pg/ml, p = 0.0016; Fig. 2B), and the levels of CXCL4 in SSc were positively correlated with the number of digital ulcers (Fig. 2C), indicating the close association of CXCL4 with peripheral microvascular involvement severity in SSc.

We also investigated whether CXCL4 could serve as a biomarker, which performed on the combined group using ROC analysis to select a threshold (Fig. 2D). Patients who had a higher baseline level of CXCL4 (over 2797 pg/ml) had a significantly increased prevalence of newly-onset of digital ulcer in 6 months (34.78% vs 65.22%, odds ratio 6.133, P = 0.0335; Fig. 2E), with unbiased background treatments (data not shown).

Next, we stratified SSc patients according to their nailfold videocapillaroscopy (NVC) pattern and compared the CXCL4 levels between subsets, we also evaluated the possible correlation of CXCL4 levels with the mean number of nailfold capillary in the patients with SSc (n = 58). SSc patients with early NVC pattern demonstrated elevated serum CXCL4 levels rather than those with active or late NVC pattern (p=0.0159 and p=0.0063 for each comparison) (Fig. 2F). The levels of CXCL4 were negatively correlated with the mean number of nailfold capillary in patients with SSc (Fig. 2G).

SSc derived CXCL4 disturbed angiogenesis

It is widely reported that the stimulation with SSc sera disturbed angiogenic performance of HUVECs[16-18], and, as we showed above, circulatory CXCL4 levels were raised in the patients with SSc and correlated with their peripheral vasculopathy. Thereby, we treated HUVECs using recombinant human CXCL4 or SSc sera, with or without CXCL4 neutralized antibody, in order to testify the contribution of SSc sera derived CXCL4 to the angiogenesis of endothelial cell line HUVECs, including viability, migration and tube formation.

Firstly, the addition of CXCL4 or the SSc sera to the medium of HUVECs inhibited endothelial cell proliferation determined using CCK-8 assay, which could be significantly ameliorated by antibody-mediated neutralization (***P < 0.001, Fig. 3A and 3B).

Furthermore, the stimulation with CXCL4 significantly decreased the ability of the tube formation and migration of HUVECs, which were dramatically improved by treating with anti-CXCL4 antibody (*P < 0.05, ***P < 0.001, Fig. 3C and 3E). Similarly, both of tube formation and migration was significantly impaired after challenging with 10% SSc sera, and this inhibitory effects on HUVECs were significantly reversed by
pretreatment with an anti-CXCL4 antibody (Fig. 3D and 3F). Therefore, these data revealed the anti-angiogenic effects of SSc derived CXCL4.

As CXCR3 mediated Ca\(^{2+}\) mobilization and chemotaxis in response to C-X-C chemokines, including CXCL4, we also confirmed the expression of CXCR3 in HUVECs (Supplemental Fig. 1A), we thereby wondered if CXCL4 inhibited endothelial cell proliferation via its receptor CXCR3. However, the addition of AMG487, a specific antagonist of CXCR3, did not show to reverse the inhibition of cell viability, tube formation and migration induced by CXCL4 or SSc sera (P > 0.05 against the CXCL4 group or SSc sera group; Supplemental Fig. 1B-G). Therefore, the data showed that CXCL4 exerted its anti-angiogenic effect on HUVECs not through CXCR3.

**CXCL4 regulated ET-1, c-Abl/Fli-1 pathway in HUVECs**

As ET-1 increased and Fli-1 deficiency participate in SSc peripheral vasculopathy[19-21], we examine whether CXCL4 exerted its anti-angiogenic effect through these mediators. We found that CXCL4 induced the upregulation of ET-1 and downregulation of Fli-1 in a dose-dependent manner at mRNA and protein levels in HUVECs, and these changes were reversed by the pre-treatment of anti-CXCL4 antibody (*P < 0.05, **P < 0.01, ***P < 0.001; Fig. 4 A-D), but not CXCR3 pharmacological inhibition (P > 0.05; Supplemental Fig.1H and 1I).

Since the activation of c-Abl pathway is a negative regulating Fli-1 deficiency[22, 23], we initially looked at the effect of CXCL4 on the c-Abl signaling. As shown in Fig.5A, CXCL4 increased the expression of c-Abl in a dose-dependent manner, and CXCL4 neutralizing antibody inhibited the overexpression of c-Abl induced by CXCL4 (P < 0.001 for each comparison). Next, we showed the time course of c-Abl induction by CXCL4 treatment in HUVECs (P < 0.001 for each comparison; Fig. 5B). We also investigated that c-Abl inhibition by ponatinib[24] normalized the reduced levels of Fli-1(*P < 0.05, **P < 0.01, ***P < 0.001; Fig. 5D and 5E). Collectively, these data suggest that CXCL4 regulates Fli-1 via c-Abl pathway in SSc vasculopathy.

**CXCL4 blocked the pro-angiogenic effect of TGF-β and PDGF in HUVECs**

Numerous pro-angiogenic mediators like transforming growth factor (TGF)-β and platelet-derived growth factor (PDGF) activated in SSc[25, 26]. Since CXCL4 could bind to growth factors directly to exert its effect, we examine whether TGF-β and PDGF are involved in the progressive vasculopathy of CXCL4 induced. As shown in Fig.6 A and 6B, TGF-β and PDGF significantly induced the proliferation of HUVECs as previously reported[27-30] and the inhibitors against TGF-β or PDGF blocked their pro-angiogenic effect respectively (P < 0.0001 for each comparison). Interestingly, the addition of CXCL4 reduced the cell proliferation of HUVECs induced by TGF-β and PDGF, which were completely reversed by the pre-treatment of the CXCL4 neutralizing antibody. These data suggested that CXCL4 exerted its anti-angiogenic effect by antagonizing TGF-β and PDGF signaling.
CXCL4 was recently recognized to induce macrophage into a specific phenotype as "M4", which typically express interleukin (IL) -6, S100A8, matrix metalloproteinase (MMP) -7 and tumor necrosis factor (TNF) -α, activate endothelial cells in atherosclerosis[7]. We herein studied whether CXCL4 exerted its anti-angiogenic effect by shifting the phenotype of macrophage to “M4”. Similar to the previous study[11], we induced THP-1 into “M4” macrophage after CXCL4 stimulation (*P < 0.05, **P < 0.01, ***P < 0.001; Supplemental Fig. 2). The Addition of the conditional medium (CM) from “M4” induced by CXCL4 to the cultures of HUVECs did not change the expressions of ET-1 or Fli-1 (P > 0.05; Supplement Fig. 3). Moreover, when co-culturing with this “M4” macrophage (CXCL4-induced), the HUVECs did not show the changed expressions of these genes in HUVECs (P > 0.05; Supplement Fig. 4). Hence, these results suggested that CXCL4 contributed to anti-angiogenesis independent of triggering macrophage to the “M4” phenotype.

Discussion

In this article, we provide the evidence that CXCL4 plays an important role in peripheral vasculopathy of SSc. First, we showed here that serum CXCL4 levels are significantly increased in patients with SSc and VEDOSS patients. Higher circulating CXCL4 levels associated with the microvascular abnormalities in SSc. Second, CXCL4 was found to exerts its anti-angiogenesis function in SSc, including inhibiting endothelial cells proliferation, migration, and tube formation with c-Abl/Fli-1 pathway involvement. Third, TGF-β and PDGF signaling may participate in the anti-angiogenesis effects of CXCL4.

CXCL4, as one of the recognized anti-angiogenesis chemokines, affects anti-angiogenesis via various ways. CXCL4/PF-4 inhibits endothelial cell proliferation[31] and migration[32], and thus displays anti-tumoral activity by inhibiting tumor growth[33] and suppressing the formation of metastase[34]. In SSc patients, the serum level of CXCL4 is positively correlated with PAH[35], suggesting that CXCL4 may be involved in SSc vasculopathy. Similar to previous studies, we found that exogenous CXCL4 can effectively inhibit angiogenesis on HUVECs, including cell viability, migration, and tube formation. Notably, we demonstrated that the ability of serum from SSc patients to inhibit angiogenesis was largely dependented on CXCL4 in HUVECs, as the addition of anti-CXCL4 almost completely reverted anti-angiogenesis following serum stimulation.

As the mechanism of CXCL4 in SSc vascular lesions is not yet clear, we selected two related factors of vascular mediators, i.e. ET-1 and Fli-1, and further explored the usability of anti-CXCL4 as a possible therapeutical target in SSc vasculopathy. In vitro, we found that CXCL4 and SSc patient serum could increase the expression of ET-1 and decrease Fli-1, and anti-CXCL4 could significantly reverse these effects. In this respect, the downregulation of Fli-1 is concentrated, since the elective endothelial cell deletion of Fli-1 in mice models leads to a disorganized dermal vascular network with greatly compromised vessel integrity and markedly increased vessel permeability[36]. Fli-1 deficiency is also related with the vasculature dysfunction in SSc[37]. Importantly, in HDMECs gene silencing of Fli-1 modulates the expression of various genes, including VE-Cadherin, PECAM1, MMP9, cathepsin B, and cathepsin V, towards an SSc EC phenotype[38–40]. These findings have explained the existence of high
levels of pro-angiogenic factors in SSc patients, as well as the presence of anti-angiogenic factors like CXCL4. The imbalance of pro- and antiangiogenic factors might explain the pathogenetic mechanisms of SSc vasculopathy.

Since c-Abl pathway negatively regulates the expression of Fli-1\[22\], we then put forward to explore the mechanism of Fli-1 deficiency via c-Abl after CXCL4 treatment. We found that the c-Abl pathway was activated under CXCL4 treatment. Furthermore, inhibited c-Abl kinase activity could reverse the decreased expression of Fli-1 in HUVECs. These findings are also supported by a recent study, which shows that blocking c-Abl kinase activity or decreasing the expression of c-Abl upregulates Fli-1 on SSc fibroblasts\[41\]. In fact, c-Abl kinase inhibition like imatinib mesylate is enrolled in dcSSc clinical trials. 1 year of treatment with imatinib significantly improved forced vital capacity (FVC) and mRSS in patients with dcSSc\[42\]. However, imatinib is poorly tolerated and thus limits its application in SSC\[43\]. In this respect, it is critical to find other small-molecules as targeted drugs for SSC. Thus, our study provides direct evidence that CXCL4 participates in SSC pathogenesis, and suggests targeting this cytokine as a potential therapeutic strategy for SSc.

A previous study demonstrated that CXCL4 could exert its effect through multiple ways, including binding to its specific receptors or other growth factors\[44\]. Our data suggest that although CXCR3 is expressed on HUVECs, blocking CXCR3 did not affect the anti-angiogenic effects of CXCL4. Furthermore, a quantity of evidence suggested that growth factors TGF-β and PDGF play important roles in vasculopathy of SSc\[25, 26\]. Thus, we hypothesis that CXCL4 may directly bind to these growth factors and then exerts its effects in SSc vasculopathy. As shown in Fig. 6, TGF-β and PDGF can effectively promote endothelial cell proliferation as previously reported\[25, 26\]. However, the angiogenic effects of TGF- β and PDGF were significantly reversed after CXCL4 pretreatment. Taken together, our data suggested that CXCL4 may directly binding to TGF- β and PDGF in SSc.

CXCL4 not only exerts its anti-angiogenesis ability in SSc, but also activates immune cells, including dendritic cells\[45\], T cell\[46\], neutrophil\[47\], mast cell\[48\], and macrophage\[49\] according to many studies, thus widely participate in the pathogenesis of various diseases. For example, CXCL4 induces monocyte differentiation resulting in a macrophage phenotype called ‘M4’, which especially expresses MMP7 and S100A8. ‘M4’ is distinct from M1 and M2 macrophages, presenting proinflammatory and cytotoxicity, reduce LDL-uptake in atherosclerosis\[7, 50, 51\]. In SSc, CXCL4 can drive fibroblast activation indirectly via PDGF-BB production by macrophage. However, in vascular endothelial cells, it is still unclear whether CXCL4 can regulate angiogenesis by inducing macrophages. In this study, although M4 macrophages have been induced at 7 μg/ml of CXCL4 as previously reported\[51\], neither the ‘M4’ macrophage-endothelial cell co-culture system nor the endothelial cells cultured with ‘M4’ macrophage CM have an effect on the related vascular mediators of SSc. Concerning the fact that we still need further experiments to confirm that ‘M4’ macrophages do exist in SSc patients, we cautiously conclude that CXCL4’s involvement in SSc vasculopathy may not depend on the activation of macrophages.
In our study, 20 ng/ml CXCL4 may inhibit the endothelial cell proliferation, but 7 µg/ml CXCL4 was needed in M4 macrophage polarization. This discrepancy may be attributable to the different sensitivity of these two different cells to CXCL4. A higher concentration is required to exert the effect when CXCL4 mediates rapid processes such as macrophage activation, while this concentration became lower when CXCL4 is involved in multistep differentiation processes such as angiogenesis [52]. However, the mechanism of these phenomena still remains to be further investigated.

The possible pathobiological role of CXCL4 in fibrosis of SSc was highly proved in the previous studies [12, 53, 54]. For example, serum CXCL4 levels were significantly increased in SSc and served as a marker of lung fibrosis [10]. Consistent with these works, we observed mRSS scores positively correlated with CXCL4 levels, suggesting CXCL4 may indicate much severer skin fibrosis in SSc.

The association of circulating CXCL4 levels with peripheral vasculopathy in patients with VEDOSS also deserves discussion. In fact, we strongly put forward that CXCL4 could serve as a biomarker in VEDOSS. Firstly, we found that serum CXCL4 levels were significantly increased in patients with VEDOSS compared with all subsets of SSc, which is similar to other researches [15, 55, 56]. Secondly, our data suggested that the abnormal NVC pattern and DUs were positively correlated with CXCL4 levels. Finally, CXCL4 could predict new DU in an expected cohort for 6 months. Given that the abnormal NVC pattern and DUs are the most prominent clinical symptoms reflecting pathologically activated angiogenesis in VEDOSS, it suggested that CXCL4 may be a helpful biomarker in VEDOSS patients.

Beyond the abovementioned achievements, there is still a limitation in this study. Although we revealed that CXCL4 can regulate the c-Abl pathway to reduce the expression of Fli-1, more experiments are required to further validate our findings in vivo and in vitro. In fact, c-Abl activation sequentially decreased the nuclear localization of PKC-δ and Fli-1 phosphorylation at threonine 312, and eventually leading to Fli-1 deficiency. Considering the commercial factors and intellectual property issues in obtaining Fli-1 phosphorylated protein, it is not available for us to further explore the downstream activation of the c-Abl pathway. However, Akamata et al. demonstrated that activation of the c-Abl/ /PKC-δ/pFli-1 pathway reduced the level of FLI-1 protein in fibroblasts, epithelial cells, endothelial cells, and in SSc animal models [36, 41, 57]. An alternative research perspective, which only explores the activation of the c-Abl pathway, could be adopted in investigating Fli-1 deficiency in SSc.

**Conclusion**

We documented a series of data confirming a possible contribution of CXCL4 to the development of vasculopathy in SSc. CXCL4 may serve as a biomarker in VEDOSS patients. Moreover, CXCL4 is likely to be involved in endothelial cell dysfunction, leading to the development of vasculopathy in SSc. The involvement of the Fli-1 pathway further supports the notion that CXCL4 is a critical anti-angiogenetic factor of SSc.

**List Of Abbreviations**
SSc: Systemic sclerosis
PAH: Pulmonary arterial hypertension
VEDOSS: Very early diagnosis of systemic sclerosis
HUVECs: Human umbilical vein endothelial cells
DU: Digital ulcers
NVC: Nailfold video capillaroscopy
Fli-1: Friend leukaemia integration factor-1
TGF: Transforming growth factor
PDGF: Platelet-derived growth factor
RP: Reynaud’s phenomenon
SRC: Scleroderma renal crisis
ECM: Extracellular matrix
ILD: Interstitial lung disease
LRP1: Lipoprotein-related protein-1
pDC: Plasmacytoid dendritic cell
ACR: American College of Rheumatology
EULAR: European League Against Rheumatism
mRSS: Modified Rodnan skin score
HCs: Healthy controls
NSAIDs: Nonsteroidal anti-inflammatory drugs
ROC: Receiver-operating-characteristic
ssSSc: Sine scleroderma
IcSSc: Limited cutaneous systemic sclerosis
dcSSc: Diffuse cutaneous systemic sclerosis
ET: Endothelin
IL: Interleukin
MMP: Matrix metalloproteinase
TNF: Tumor necrosis factor
ETAR: Endothelin A receptor
AT1R: Angiotensin type 1 receptor
FVC: forced vital capacity
ELISA: Enzyme-linked immunosorbent assay
CCK8: Cell counting kit-8
RT-qPCR: Real-time polymerase chain reaction
CM: Conditional medium

**Declarations**

**Ethics declarations**

Informed consent was obtained from each patient. This study was approved by the Ethics Committee of Huashan Hospital, Fudan University.

**Consent for publication**

Not applicable.

**Availability of data and material**

Our supporting data are available.

**Competing interests**

The data presented in this manuscript are original and have not been published or submitted elsewhere. All listed authors have approved the manuscript and agreed with the submission. The authors declare that they have no conflict of interest.

**Funding**

This work was supported by the National Natural Science Foundation of China (81501319).
Author’s Contributions

ZJ, CC and LW carried out the in vitro studies. XZ, NK and YX were responsible for the pathological assessment. WW and HH performed the statistical analysis and interpreted the data. ZJ and ML wrote the paper. HZ and ML conceived of the study, and participated in its design and coordination and helped to draft the manuscript. All authors have read and approved the final manuscript.

Acknowledgements

Not applicable.

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References


Figures
Figure 1

CXCL4 sera levels detected by ELISA assay. (A-C) Serum CXCL4 levels in health controls (HC), patients with systemic sclerosis (SSc) and patients not fulfilling the 2013 American College of Rheumatology (ACR)/European League Against Rheumatism (EULAR) classification criteria for SSc and enrolled in the very early diagnosis of SSc (VEDOSS) project. Results from the identification (A), replication (B), and combined (C) cohorts are shown. (D) Serum CXCL4 levels in total HC and patients with SSc sine
scleroderma (ssSSc) (n=4), overlapping syndrome of SSc (n=13), limited cutaneous systemic sclerosis (lcSSc) (n=39), diffuse cutaneous systemic sclerosis (dcSSc) (n=52), and those with very early systemic sclerosis (VEDOSS) (n=22). (**p < 0.001)

Figure 2

Correlations between CXCL4 sera levels and skin fibrosis and peripheral vasculopathy in SSc. CXCL4 sera levels detected by enzyme-linked immunosorbbent assay (ELISA) assay. (A) Correlation of serum CXCL4
levels with the modified Rodnan Skin Score (mRSS) in the combined cohort. (B) Serum CXCL4 levels in SSc patients with digital ulcers (DU) and SSc patients without DU. (C) Correlation of DU number and CXCL4 levels. (D) Receiver operating characteristic (ROC) curve for DU onset for prediction of DU. (E) Correlation of patients with a high baseline level of CXCL4 (> 2797 ng/ml) and DU onset. (F, G) The nailfold videocapillaroscopy (NVC) pattern (F) in SSc patients and the correlation of CXCL4 levels and the mean number of nailfold capillary (G) (n=58). (***p < 0.001)
The anti-angiogenesis effects of CXCL4 on HUVECs. HUVECs were stimulated with indicated concentration of CXCL4 with or without an anti-CXCL4 antibody (3 µg/ml). HUVECs were pretreated with SSc sera or health sera with or without an anti-CXCL4 antibody (3 µg/ml). (A, B) Cell viability was evaluated by Cell counting kit-8 (CCK8) assay after 24 hours. (C, D) Tube formation photographed and the number of tubes per well were evaluated at 4 hours. (E, F) Cell scratch test was photographed and wound closure area were assessed at 4 hours. The edges of the cell (red dotted line) are shown in the images. (*P < 0.05, **P < 0.01, ***P < 0.001)
Figure 4

The gene expression level of endothelin (ET)-1 and friend leukaemia integration factor (Fli)-1 after CXCL4 treatment. The anti-angiogenesis effects of CXCL4 on HUVECs. HUVECs were stimulated with indicate concentration of CXCL4 with or without an anti-CXCL4 antibody (3 µg/ml). HUVECs were pretreated with SSc sera or health sera with or without an anti-CXCL4 antibody (3 µg/ml). (A-D) ET-1 and Fli-1 gene expression levels measured by real-time polymerase chain reaction (RT-qPCR) (A, C) and western blot (B, D). (*P < 0.05, **P < 0.01, ***P < 0.001)
Figure 5

The activation of c-Abl pathway by CXCL4. (A) HUVECs were exposed to CXCL4 in different concentrations, as indicated. Western blot analysis and quantification of protein levels of c-Abl. (B) HUVECs were stimulated with CXCL4 at indicate time. Western blot analysis and quantification of protein levels of c-Abl. (C-D) HUVECs were stimulated with 20ng/ml CXCL4 with or without c-Abl inhibitor ponatinib (10nM). (C) Western blot analysis the protein levels of c-Abl. (D) Western blot analysis the protein levels of Fli-1. (*P < 0.05, **P < 0.01, ***P < 0.001)

Figure 6

The involvement of transforming growth factor (TGF)-β and platelet-derived growth factor (PDGF) signaling in the anti-angiogenetic effects of CXCL4. (A, B) Cell viability was evaluated by CCK8 assay after 24 hours. (A) HUVECs were treated with TGF-β (10ng/ml) for 24 hours, and some cultures were pretreated with SB525334 (1µM), a selective inhibitor of TGF-βR1, or with CXCL4 (20ng/ml) with or without an anti-CXCL4 antibody (3 µg/ml). (B) HUVECs were treated with PDGF (10ng/ml) for 24 hours, and some cultures were pretreated with blocking PDGFR imatinib (1µM), or with CXCL4 (20ng/ml) with or without an anti-CXCL4 antibody (3 µg/ml). (**P < 0.001)

Supplementary Files

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