#' Simulation of SNP array intensities

#'

#' @description \code{SNParraySim} simulates X and Y array intensities, based

#' on a matrix of SNP dosages simulated by PedigreeSim. Individuals of

#' different ploidy can be mixed in the same dosage matrix (e.g. diploid and

#' tetraploid parents of a triploid F1 population). Array intensity (of X or Y)

#' is modelled as the summation of two sources of fluorescence intensity:

#' signal and background (here also called background offset).

#'

#' @param dsg a matrix of allele dosages with markers in rows and individuals

#' in column.

#' @param ploidy a vector indicating the ploidy of each individual.

#' @param avint numeric. It is an overall average intensity

#' @param sd\_mrk a number between 0 and 1. It is a coefficient of variation (CV)

#' to define the average Mf of each marker. It will affect the variation of

#' R (R=X+Y) between markers. The default value is 0.05 (5%).

#' @param sd\_ind a numeric vector of two elements, defining the parameters shape1

#' and shape2 of a beta distribution, used to sample a coefficient of variation

#' (CV) per marker. This CV will define the sd of Mf values within each marker,

#' affecting in the end the spread of R within markers.

#' @param ale\_b a numeric vector of two elements, defining the parameters shape1

#' and shape2 of a beta distribution, used to sample the yunb ratios (allelic bias).

#' @param od numeric, between 0 and 1. It is a coefficient of variation

#' defining the sd for the error term in X and Y simulation, which add extra variation

#' to model.

#' @param bc the parameter shape2 of rbeta (rbeta(n=1000,shape1=2,shape2=10)),

#' used to sample the Roff values (Roff = Roffset/Rsignal).

#' @param bc\_XY a numeric vector of two elements, defining the parameters shape1

#' and shape2 of a beta distribution, used to sample Yoff, that is the Y

#' proportion of the background offset intensity.

#' @param seed\_number a numeric. Define the seed.

#'

#' @return a list containing X (matrix of X intensity), Y (matrix of Y

#' intensity) and mpar (listing values of yunb, Roff and Yoff per marker)

#' @export

#'

SNParraySim <- function(dsg,

 ploidy,

 avint = 1000, #overal average intensity

 sd\_mrk = 0.05, #affect spread of mean intensities (per marker)

 sd\_ind = c(20,100), #affect spread of intensities within markers

 ale\_b = c(2,2), #affect XY unbalance (allelic bias) for signal intensity (yunb)

 od = 0.02, #affect cluster width (overdispersion)

 bc = 10, #affect amount of background offset (Roff)

 bc\_XY = c(2,2), #affect XY unbalancement for offset

 seed\_number = NULL) #set a seed for random sampling

{

 #### step 1 - convert dosages in expected XY ratios

 thr <- sapply(1:ncol(dsg), function(x) {

 round(dsg[,x]/ploidy[x],3)

 })

 #### step 2 - define Mf, a multiplication factor of individual intensity

 ## Average Mf per marker

 # \*sd\_mrk\* is a coefficient of variation (CV), defining the sd of Mf values

 if(is.numeric(seed\_number)) set.seed(seed\_number)

 Mf <- rnorm(nrow(dsg), avint, avint\*sd\_mrk)

 Mf[Mf<0] <- 0.1 #to prevent negative Mf values

 ## Individual variation of Mf (Mf variation within markers)

 # Here, we want to define the variation of Mf within each marker, based on a normal distribution.

 # Therefore, we need to sample a Coefficient of Variation (CV) for each marker. Since CV values

 # ranges from 0 to 1, they are sampled from a beta distribution. A beta distribution is defined by

 # the two parameters shape1 and shape2, here called sd\_ind [1] and sd\_ind [2]. To increase variation,

 # that is to make clusters "longer", you need to increase sd\_ind [1].

 # See the example distributions below:

 # hist(rbeta(10000,20,100), xlim = c(0,1))

 # hist(rbeta(10000,40,100), xlim = c(0,1))

 # hist(rbeta(10000,60,100), xlim = c(0,1))

 # hist(rbeta(10000,80,100), xlim = c(0,1))

 # The CV is used to calculate the Standard Deviation (SD) of each marker Mf distribution.

 if(is.numeric(seed\_number)) set.seed(seed\_number)

 SD <- sapply(Mf, function(x) x \* rbeta(1, sd\_ind[1], sd\_ind[2]))

 ## full matrix of individual Mf values (with x markers and n individuals)

 if(is.numeric(seed\_number)) set.seed(seed\_number)

 Mfi <- t(sapply(1:nrow(dsg), function(x){

 sapply(1:ncol(dsg), function(n){

 rnorm(1,Mf[x],SD[x])

 })

 }))

 Mfi[Mfi<0] <- 0.1 #to prevent negative Mf values

 #### step 3 - sample allelic bias

 # yunb is a number between 0 and 1, representing the "fluorescence quantum yield" of Y (Qy),

 # relative to Qx. yunb = Qy/(Qx+Qy)

 if(is.numeric(seed\_number)) set.seed(seed\_number)

 yunb <- rbeta(length(Mf),ale\_b[1],ale\_b[2])

 #### step 4 - simulate signal intensity of X and Y

 # simulate Y

 if(is.numeric(seed\_number)) set.seed(seed\_number)

 Y <- t(sapply(1:nrow(thr), function(i){ #per marker

 sapply(1:ncol(thr), function(j){ #per individual

 Mfi[i,j] \* thr[i,j] \* yunb[i] + rnorm(1, 0, Mf[i] \* od) #add overdispersion

 })

 }))

 colnames(Y) <- colnames(dsg);rownames(Y) <- rownames(dsg)

 # simulate X

 if(is.numeric(seed\_number)) set.seed(seed\_number)

 X <- t(sapply(1:nrow(thr), function(i){ #per marker

 sapply(1:ncol(thr), function(j){ #per individual

 Mfi[i,j] \* (1-thr[i,j]) \* (1-yunb[i]) + rnorm(1, 0, Mf[i] \* od) #add overdispersion

 })

 }))

 colnames(X) <- colnames(dsg); rownames(X) <- rownames(dsg)

 #### step 5 - background offset

 # Roff is a number between 0 and 1, so that Roff = Roffset/Rsignal

 if(is.numeric(seed\_number)) set.seed(seed\_number)

 Roff <- rbeta(length(Mf),2,bc)

 # Yoff is a number between 0 and 1, so that Yoff = Yoffset/Roffset

 if(is.numeric(seed\_number)) set.seed(seed\_number)

 Yoff <- rbeta(length(Mf),bc\_XY[1],bc\_XY[2])

 ## add the background offset

 Y <- t(sapply(1:nrow(Y), function(i) { #per marker

 Y[i,] + mean(Y[i,]+X[i,]) \* Roff[i] \* Yoff[i]

 })) # where mean(Y[i,]+X[i,]) is the mean R signal per marker

 X <- t(sapply(1:nrow(X), function(i) {

 X[i,] + mean(Y[i,]+X[i,]) \* Roff[i] \* (1-Yoff[i])

 }))

 #### step 6 - add balanced background (to prevent negative intensities)

 yneg <- apply(Y,1,min); xneg <- apply(X,1,min)

 mneg <- apply(cbind(yneg,xneg),1,min) #minimum intensity per marker

 mneg[mneg>0] <- 0

 Y <- t(sapply(1:nrow(Y), function(i) { #per marker

 Y[i,] - mneg[i] + avint\*0.1

 }))

 X <- t(sapply(1:nrow(X), function(i) {

 X[i,] - mneg[i] + avint\*0.1

 }))

 #### step 7 - output

 mpar <- data.frame(MarkerName=rownames(dsg),

 yunb=round(yunb,2),

 Roff=round(Roff,2),

 Yoff=round(Yoff,2))

 output <- list(X=X, Y=Y, mpar=mpar)

 return(output)

}