

Accumulate evidence for IP-10 in diagnosing pulmonary tuberculosis

Xia Qiu

Sichuan University West China Second University Hospital

Tao Xiong

Sichuan University West China Second University Hospital

Xiaojuan Su

Sichuan University West China Second University Hospital

Yi Qu

Sichuan University West China Second University Hospital

Long Ge

Lanzhou University

Yan Yue

Sichuan University West China Second University Hospital

Yan Zeng

Sichuan University West China Second University Hospital

Wenxing Li

Sichuan University West China Second University Hospital

Peng Hu

Sichuan University West China Second University Hospital

Dezhi Mu (✉ mudz@scu.edu.cn)

Research article

Keywords: pulmonary tuberculosis; interferon gamma-induced protein 10; diagnosis; meta-analysis

Posted Date: September 24th, 2019

DOI: <https://doi.org/10.21203/rs.2.11284/v2>

License: © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License. [Read Full License](#)

Version of Record: A version of this preprint was published on October 30th, 2019. See the published version at <https://doi.org/10.1186/s12879-019-4466-5>.

Abstract

Backgrounds: Pulmonary tuberculosis (PTB) is a major health and economic burden. Accurate PTB detection is an important step to eliminating TB globally. Interferon gamma-induced protein 10 (IP-10) has been reported as a potential diagnostic marker for PTB since 2007. In this study, a meta-analysis approach was used to assess diagnostic value of IP-10 for PTB. **Methods:** Web of Science, PubMed, the Cochrane Library, and Embase databases were searched for studies published in English up to February 2019. The pooled sensitivity, specificity, positive likelihood ratio (PLR), negative likelihood ratio (NLR), diagnostic odds ratio (DOR), the area under the curve (AUC) and hierarchical summary receiver operating characteristic (HSROC) curve were estimated by the HSROC model and random effect model. **Results:** Eighteen studies including 2836 total participants met our inclusion criteria. The pooled sensitivity, specificity, PLR, and NLR of IP-10 for PTB detection were 86%, 88%, 7.00, and 0.16, respectively. The pooled DOR was 43.01, indicating a very powerful discriminatory ability of IP-10. The AUC was 0.93 (95% CI: 0.91–0.95), showed the accuracy of IP-10 was good. Meta-regression showed that there was no heterogeneity with respect to TB burden, study design type, age, IP-10 assay method, IP-10 condition and HIV-infection status. **Conclusions:** Our results showed that IP-10 is a promising marker for differentiating PTB from non-TB.

Background

Tuberculosis (TB), a highly contagious disease, is still a major health and economic burden [1]. Globally, approximately 10 million individuals developed TB and more than 1.3 million died of the disease in 2017, according to a WHO report [2]. Pulmonary tuberculosis (PTB), accounting for 75% of all TB cases, contributes substantially to TB mortality, especially with HIV co-infection [3, 4]. Correctly discriminating PTB is an important step to eliminate TB by 2030, a goal established by the WHO [2].

In clinical practice, sputum smear microscopy is ineffective for detecting PTB [5]. Specimen culture for *Mtb* provides the most accurate diagnosis [6]. However, the results of microbiological examination and acid-fast bacillus stains depend on the sputum sample. Immunological tests, such as the tuberculin skin test (TST) and interferon-gamma release assay (IGRA), are auxiliary diagnostic tools for PTB [7]. TST has a low specificity in Bacilli Calmette Guerin (BCG)-vaccinated individuals [7]. In children, IGRAs can yield many indeterminate results [8, 9]. Considering these limitations, additional valid tools are required to improve the diagnosis of PTB.

Interferon gamma-induced protein 10 (IP-10), an IFN-gamma-inducible chemokine, could be expressed at 100-fold higher levels than those of IFN-gamma after TB infection [10, 11]. Age and gender do not affect the level of IP-10 [11, 12]. Since 2007, IP-10 has been reported as a potential parameter for PTB detection [7] [13-29].

Many studies have evaluated the diagnostic potential of IP-10 for PTB, but the results are variable. Therefore, the aim of this study was to synthesize and analyze the diagnostic value of IP-10 for PTB.

Methods

Literature search

This study followed the Preferred Reporting Items for Systematic Reviews and Meta-Analyses Diagnostic Test Accuracy criteria 2018 (PRISMA-DTA 2018) [30]. The Web of Science, PubMed, the Cochrane Library and Embase databases were used to search for relevant English language citations published up to February 2019. Our search terms were “tuberculosis,” “pulmonary tuberculosis,” “Chemokine CXCL10,” and “interferon gamma-induced protein 10.” Comprehensive literature search strategies were used based on the following combination of MeSH terms, title/abstracts and all fields for these databases (Supplementary Table). Additionally, the reference lists of the applicable studies, relevant research letters, and reviews were manually searched to find other potentially relevant studies.

Literature selection

Two investigators independently determined literature eligibility. Studies reporting IP-10 levels for the detection of PTB were included according to the following criteria: (1) reporting on individuals with PTB and non-TB (population); (2) provision of IP-10 in whole blood and plasma as index test; (4) *Mtb* culture as a gold standard, and other reference standard including pathological examination, microscopy and genexpert MTB/RIF test (WHO recommended) [2]; (5) the primary outcomes including diagnostic performance of IP-10 (sensitivity and specificity); (5) randomized controlled trials, prospective and retrospective studies included (study design); (6) more than 10 individuals reported meeting the inclusion criteria. Studies not published in English, other letters (except research letters), conference abstracts, veterinary experiments, reviews and case reports were excluded.

Data extraction

The following data were extracted: the first author, year of publication, country, TB high-burden, study design, age, number of participants (patients with PTB and non-TB subjects), TB site, non-TB status, cut-off for index test (IP-10), diagnostic reference standard, method and condition for the IP-10 assay, HIV-infection status, sensitivity, specificity, true positive (TP), false positive (FP), false negative (FN), and true negative (TN) for IP-10. Two investigators independently extracted data from eligible articles, and disagreements were resolved by discussing and reaching a consensus.

Quality assessment

According to the Cochrane Collaboration, two investigators independently reviewed the methodological quality of eligible articles by Quality Assessment of Diagnostic Accuracy Studies tool-2 (QUADAS-2) [31, 32]. Disagreements were resolved by consensus. Revman (version 5.3) was used to perform the quality assessment.

Data analysis

Excel was used to construct a two-by-two table, including TP, FP, FN, and TN for patients with PTB. Stata (version 14.0) was used to perform the data analysis. The index test had different optimal cut-offs. According to the recommendation of Cochrane Collaboration, the hierarchical summary receiver operating characteristic (HSROC) model by Rutter et al. was utilized when the index test was assessed by applying various thresholds [32, 33]. The HSROC curve was computed with the "metandi" command [34]. Prediction region presented possible point of sensitivity and specificity in the HSROC curve. The summary point showed the pooled sensitivity and specificity under the optimal threshold value. Confidence region reflected the possible summary point.

The main outcomes were the diagnostic performance of IP-10 for detecting PTB by the random effect model, as evaluated by the summary estimates of sensitivity, specificity, positive likelihood ratio (PLR), negative likelihood ratio (NLR), diagnostic odds ratio (DOR), and the area under the curve (AUC). Sensitivity, reflecting the ability of index test to detect patients, calculated by " $\text{Sensitivity} = \text{TP}/(\text{TP} + \text{FN})$ ". Specificity, reflecting the ability of index test to eliminate disease-free, calculated by " $\text{Specificity} = \text{TN}/(\text{FP} + \text{TN})$ ". PLR, a measure of index test for detection potential for disease, could be calculated by the formula " $\text{PLR} = \text{Sensitivity}/(1 - \text{Specificity})$ ". NLR, a measure of index test for detection potential for non-disease, could be calculated by the formula " $\text{NLR} = (1 - \text{Sensitivity})/\text{Specificity}$ ". DOR, a measure for overall accuracy of index test, could be calculated by the formula " $\text{DOR} = (\text{TP}/\text{FN})/(\text{FP}/\text{TN})$ ". AUC, indicated how the index test was accurate, especially exceeded 0.90. 95% confidence interval (CI) was calculated by wilson method and no correction factor applied.

The I^2 value was not suitable for the quantification of heterogeneity in accuracy studies [35]. Thus, to explore potential sources of heterogeneity, we used a meta-regression analysis with the "midas" command. The intercept was zero. Seven subgroups were created: TB high-burden country (yes or no), study design type (cohort or not), age (adults or not), IP-10 method (multiplex cytokines assay or ELISA), IP-10 condition (unstimulated or stimulated), and HIV-infection status (yes/some or no).

The Deeks test was used to assess publication bias using the "midas" command [36]. No publication bias existed when studies evenly distributed on the sides of regression line or P value exceeded 0.05 in Deeks' funnel plot.

Results

Search results

In total, 1349 records were identified from our literature searches (Figure 1). After removing 623 duplicates, we read titles and abstracts and excluded 682 records. An additional 447 records were non-eligible for various reasons (e.g., studies involving leprosy, Crohn's disease, pneumonia, monocyte chemoattractant protein-1, interleukin-12, and interleukin-18), 73 records were animal experiments (mouse, calves, warthogs, etc.), 69 records were reviews, abstracts, and letters, 58 records focused on extra-PTB (pleural TB, TB meningitis, osteoarticular TB, etc.), and 5 records were non-English (Chinese, Russian, Polish, etc.). Then, we reviewed the full texts of 44 articles. Ultimately, 18 articles were included in this study.

Characteristics of included studies

The main characteristics of the 18 articles, comprising 24 trials, are listed in Table 1 and 2 [7] [13-29]. In total, 2836 participants were involved. The year of publication ranged from 2012 to 2018. Nine (50%) studies were from TB high-burden countries (China, South Africa, India, Thailand, and Uganda), and nine (50%) studies were from TB low-burden countries, according to WHO [2]. Study design, TB site, non-TB status, IP-10 cut-off, and reference standards are summarized in Table 1. IP-10 method, condition, HIV-infection status, cut-off values, sensitivity, specificity, TP, FP, FN, and TN of IP-10 for each trial are shown in Table 2.

Quality of included studies

The QUADAS-2 tool reflects the methodological quality of included articles (Supplementary Figure S1). Patient selection bias was unclear for five studies; one study used a case-control design [19] and four studies did not report the time and consecutiveness of patient enrolment [17, 21, 23, 27]. Additionally, 50% of studies had unclear bias in index tests; in particular, we could not determine whether the results were interpreted in blind conditions [7, 18, 20, 23-25, 27-29]. One study had high risk of bias in the reference standard, which was clinical PTB by clinical presentation and radiological confirmation [16]. Flow and timing bias were unclear in three studies, in which patients were lost in the analysis [19-21]. The applicability concerns were generally low.

Summary statistics

A total of 2836 participants, comprising 3633 blood samples were included. The sensitivity for IP-10 was 0.86 (95% CI: 0.80–0.90) and the specificity was 0.88 (95% CI: 0.82–0.92). The pooled PLR was 7.00 (95% CI: 4.76–10.30), and the pooled NLR was 0.16 (95% CI: 0.12–0.23). The pooled DOR was 43.01 (95% CI: 25.80–71.69), indicating that the discriminatory effect of IP-10 was good. The AUC was 0.92 (95% CI: 0.89–0.94), showed the accuracy of IP-10 was good. Figure 2 shows the HSROC curves for IP-10, under the optimal threshold value, the pooled sensitivity and specificity were 0.86 and 0.88, respectively.

Heterogeneity

As shown in Table 3, heterogeneity was assessed by a meta-regression analysis. Heterogeneity was not detected with respect to TB high-burden versus TB low-burden countries ($P = 0.83$), cohort versus other study design types ($P = 0.55$), adults versus children (with or without adults) ($P = 0.59$), multiplex cytokine assay versus ELISA to detect IP-10 ($P = 0.73$), IP-10 stimulation or not ($P = 0.72$), and HIV infection or not ($P = 0.53$).

Publication bias

Deeks' funnel plot showed no statistical significance ($P = 0.20$), indicating no striking publication bias in this study (Supplementary Figure S2).

Discussion

PTB is still a major cause of death worldwide, especially in immunocompromised individuals and children younger than 5 years [37, 38]. The accurate detection and timely treatment of PTB are important components of the "End TB Strategy" globally [39]. Currently, methods for detecting PTB depend on the region, BCG-vaccinated status, HIV status, etc. The search for new markers for the auxiliary diagnosis of PTB is ongoing. Several studies have shown that IP-10 is a promising marker for PTB detection [7] [13-29].

In 2014, Guo et al. published a meta-analysis of studies of IP-10 for diagnosing TB [40]. The diagnostic performance of IP-10 was moderate. In this study, both PTB and extra-PTB individuals were included, and plasma and pleural effusion samples were included. However, the diagnostic standards for PTB and extra-PTB were different. Pleural effusion detection is more traumatic than the use of peripheral venous blood.

Considering these limitations, we performed a meta-analysis to evaluate the overall diagnostic performance of blood IP-10 as a potential biomarker for detecting PTB. We found that IP-10 could be a valuable detection tool (sensitivity: 86%, specificity: 88%). The PLR (7.00) suggested that IP-10 had good detection potential for PTB. The NLR (0.16) indicated that IP-10 distinguished non-TB individuals well. The DOR (43.01) indicated a good overall performance of IP-10 in discriminating between PTB and non-TB.

The TST and IGRA, as immunodiagnostic tests, are recommended for the auxiliary diagnosis of PTB by the WHO [2]. The TST could show cross-reactivity in BCG-vaccinated individuals. However, IP-10 is less influenced by BCG vaccination [7]. Ruhwald et al. reported that IP-10 has a much higher sensitivity (92.5%) when compared to the TST (73.9%), and suggested that IP-10 is an alternative biomarker of TST [41]. The recently developed IGRA can overcome some limitations of TST. However, it lacks power when applied to children and individuals coinfecting with HIV [9, 14]. IP-10 could be produced at a high level in these populations [42, 43]. Vanini et al. showed that the sensitivity is 66.7% for IP-10-based test and 52.4% for the IGRA in HIV-infected individuals [44].

In bivariate analyses, TB-burden country, study design, age, IP-10 detection method, assay conditions, and HIV infection status were not significant sources of heterogeneity. We also found that the diagnostic performance of IP-10 was similar in multiplex cytokine assays and ELISA (sensitivity: 84% vs. 87%, specificity: 89% vs. 87%). These two methods were comparable with respect to reliability and reproducibility [20]. Considering the cost, ELISA is preferred over multiplex cytokine assays. Stimulated and unstimulated IP-10 had similar diagnostic accuracies for PTB, suggesting that IP-10 could be detected in both conditions. IP-10 had a higher diagnostic potential in HIV-infected individuals, consistent with previous findings [45].

Certainly, our meta-analysis had several limitations. First, we enrolled studies which had various cut-offs of IP-10 assays. In most situations, the investigators of included studies might choose the different cut-offs according to their aims. Second, IP-10 assays are usually performed in combination with conventional tests, but we did not address the reliability and incremental benefit of adding IP-10 to other tests. Third, some studies included patients with PTB after treatment while others did not. Furthermore, the severity and extent of PTB might vary. These factors might influence the diagnostic potential of IP-10. Fourth, heterogeneity could not be ignored. Although the TB-burden country, design type, age, IP-10 method, IP-10 condition and HIV-infection status were not significant sources of heterogeneity in this meta-regression analysis ($P > 0.05$), they could also increase the heterogeneity and reduce the generalizability of the overall performance of IP-10. Furthermore, intercurrent diseases (diabetes mellitus and malignancy) in the included studies might influence heterogeneity.

Despite the low probability of publication bias, it was a concern. Based on the linguistic abilities of our team, only studies written in English were included. The true potential of IP-10 for discriminating PTB from non-TB might be lower than we reported.

Conclusion

In conclusion, this meta-analysis shows that IP-10 is a promising and reliable marker for differentiating PTB from non-TB. Updated global TB reports should consider IP-10 as an auxiliary diagnostic method for PTB. Furthermore, large, multi-center, prospective studies are warranted to support our findings.

Abbreviations

PTB = pulmonary tuberculosis; IP-10 = interferon gamma-induced protein 10; PLR = positive likelihood ratio; NLR = negative likelihood ratio; DOR = diagnostic odds ratio; HSROC = hierarchical summary receiver operating characteristic; CI = confidence interval; QUADAS-2 = quality assessment of diagnostic accuracy studies tool-2; TP = true positive; FP = false positive; FN = false negative; TN = true negative.

Declarations

Funding

This work was supported by the National Science Foundation of China (81330016, 81630038, 81771634, 81842011), the National Key R&D Program of China (2017YFA 0104200), the grants from the Ministry of Education of China (IRT0935), the grants from the Science and Technology Bureau of Sichuan Province (2016TD0002), and the grant from the clinical discipline program (Neonatology) from the Ministry of Health of China (1311200003303).

Availability of data and materials

The data generated in this study are included in published article and its supplementary information files.

Authors' contributions

DM had full access to all data in the study and takes responsibility for the integrity of data and accuracy of its analysis. XQ, TX, XS, YQ, LG, YY, YZ, WL, and PH contributed substantially to the study design, data analysis and interpretation, and writing of the manuscript.

Potential conflicts of interest:

All authors: No reported conflicts of interest.

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

References

1. Lawn SD, Zumla AI. Tuberculosis. *Lancet*. 2011; 378(9785):57-72.
2. The World Health Organizations. Global tuberculosis report 2018. Available at: www.aidshub.org/global-tuberculosis-report-2018-who-2018.
3. Denkinger CM, Schumacher SG, Boehme CC, Dendukuri N, Pai M, Steingart KR. Xpert MTB/RIF assay for the diagnosis of extrapulmonary tuberculosis: a systematic review and meta-analysis. *Eur Respir J*. 2014; 44(2):435-46.
4. Ford N, Matteelli A, Shubber Z, Hermans S, Meintjes G, Grinsztejn B, et al. TB as a cause of hospitalization and in-hospital mortality among people living with HIV worldwide: a systematic review and meta-analysis. *J Int AIDS Soc*. 2016;19(1):20714.
5. O'Garra A, Redford PS, McNab FW, Bloom CI, Wilkinson RJ, Berry MP. The immune response in tuberculosis. *Annu Rev Immunol*. 2013;31:475-527.
6. Wallis RS, Pai M, Menzies D, Doherty TM, Walzl G, Perkins MD, et al. Biomarkers and diagnostics for tuberculosis: progress, needs, and translation into practice. *Lancet*. 2010;375(9729):1920-37.
7. Zhao Y, Yang X, Zhang X, Yu Q, Zhao P, Wang J, et al. IP-10 and RANTES as biomarkers for pulmonary tuberculosis diagnosis and monitoring. *Tuberculosis*. 2018;111:45-53.
8. Connell TG, Tebruegge M, Ritz N, Bryant P, Curtis N. The potential danger of a solely interferon-gamma release assay-based approach to testing for latent Mycobacterium tuberculosis infection in children. *Thorax*. 2011;66(3):263-4.

9. Hausteijn T, Ridout DA, Hartley JC, Thaker U, Shingadia D, Klein NJ, et al. The likelihood of an indeterminate test result from a whole-blood interferon-gamma release assay for the diagnosis of *Mycobacterium tuberculosis* infection in children correlates with age and immune status. *Pediatr Infect Dis J*. 2009;28(8):669-73.
10. Azzurri A, Sow OY, Amedei A, Bah B, Diallo S, Peri G, et al. IFN-gamma-inducible protein 10 and pentraxin 3 plasma levels are tools for monitoring inflammation and disease activity in *Mycobacterium tuberculosis* infection. *Microbes and Infection*. 2005;7(1):1-8.
11. Qiu X, Tang Y, Yue Y, Zeng Y, Li W, Qu Y, et al. Accuracy of interferon- γ -induced protein 10 for diagnosing latent tuberculosis infection: a systematic review and meta-analysis. *Clin Microbiol Infect*. 2019;25(6):667-672.
12. Aabye MG, Latorre I, Diaz J, Maldonado J, Mialdea I, Eugen-Olsen J, et al. Dried plasma spots in the diagnosis of tuberculosis: IP-10 release assay on filter paper. *European Respiratory Journal*. 2013;42(2):495-503.
13. Blauenfeldt T, Wagner D, Aabye M, Heyckendorf J, Lange B, Lange C, et al. Thermostability of IFN-gamma and IP-10 release assays for latent infection with *Mycobacterium tuberculosis*: A TBnet study. *Tuberculosis*. 2016;98:7-12.
14. Jacobs R, Malherbe S, Loxton AG, Stanley K, van der Spuy G, Walzl G, et al. Identification of novel host biomarkers in plasma as candidates for the immunodiagnosis of tuberculosis disease and monitoring of tuberculosis treatment response. *Oncotarget*. 2016;7(36):57581-92.
15. Lee K, Chung W, Jung Y, Kim Y, Park J, Sheen S, et al. CXCR3 ligands as clinical markers for pulmonary tuberculosis. *International Journal of Tuberculosis and Lung Disease*. 2015;19(2):191-9.
16. Azab NY, Elmahallawy I, Sharara G, Abdel-Atti E. The utility of the interferon gamma-inducible protein-10 (IP-10) level in bronchoalveolar lavage and blood in the diagnosis of tuberculosis. *Egyptian Journal of Chest Diseases and Tuberculosis*. 2013;62:629-34.
17. Wang X, Jiang J, Cao Z, Yang B, Zhang J, Cheng X. Diagnostic performance of multiplex cytokine and chemokine assay for tuberculosis. *Tuberculosis*. 2012;92(6):513-20.
18. Hong JY, Jung GS, Kim H, Kim YM, Lee HJ, Cho S-N, et al. Efficacy of inducible protein 10 as a biomarker for the diagnosis of tuberculosis. *International Journal of Infectious Diseases*. 2012;16(12):E855-E9.
19. Syed Ahamed Kabeer B, Raman B, Thomas A, Perumal V, Raja A. Role of QuantiFERON-TB gold, interferon gamma inducible protein-10 and tuberculin skin test in active tuberculosis diagnosis. *PLoS ONE*. 2010;5(2):e9051.
20. Luo J, Zhang M, Yan B, Li F, Guan S, Chang K, et al. Diagnostic performance of plasma cytokine biosignature combination and MCP-1 as individual biomarkers for differentiating stages of *Mycobacterium tuberculosis* infection. *The Journal of infection*. 2018;78(4):281-291.
21. La Manna MP, Orlando V, Li Donni P, Sireci G, Di Carlo P, Cascio A, et al. Identification of plasma biomarkers for discrimination between tuberculosis infection/disease and pulmonary non tuberculosis disease. *Plos One*. 2018;13(3):e0192664.
22. Balcells ME, Ruiz-Tagle C, Tiznado C, Garcia P, Naves R. Diagnostic performance of GM-CSF and IL-2 in response to long-term specific-antigen cell stimulation in patients with active and latent tuberculosis infection. *Tuberculosis*. 2018;112:110-9.
23. Nonghanphithak D, Reechaipichitkul W, Namwat W, Naranbhai V, Faksri K. Chemokines additional to IFN-gamma can be used to differentiate among *Mycobacterium tuberculosis* infection possibilities and provide evidence of an early clearance phenotype. *Tuberculosis*. 2017;105:28-34.
24. Biraro IA, Kimuda S, Egesa M, Cose S, Webb EL, Joloba M, et al. The Use of Interferon Gamma Inducible Protein 10 as a Potential Biomarker in the Diagnosis of Latent Tuberculosis Infection in Uganda. *Plos One*. 2016;11(1): e0146098.
25. Wergeland I, Pullar N, Assmus J, Ueland T, Tonby K, Feruglio S, et al. IP-10 differentiates between active and latent tuberculosis irrespective of HIV status and declines during therapy. *Journal of Infection*. 2015;70(4):381-91.
26. Petrone L, Cannas A, Aloï F, Nsubuga M, Sserumkuma J, Nazziwa RA, et al. Blood or Urine IP-10 Cannot Discriminate between Active Tuberculosis and Respiratory Diseases Different from Tuberculosis in Children. *Biomed Research International*. 2015;2015:589471
27. Latorre I, Diaz J, Mialdea I, Serra-Vidal M, Altet N, Prat C, et al. IP-10 is an accurate biomarker for the diagnosis of tuberculosis in children. *J Infect*. 2014;69(6):590-9.
28. Wang S, Diao N, Lu C, Wu J, Gao Y, Chen J, et al. Evaluation of the Diagnostic Potential of IP-10 and IL-2 as Biomarkers for the Diagnosis of Active and Latent Tuberculosis in a BCG-Vaccinated Population. *Plos One*. 2012;7(12):e51338.
29. Jeong YH, Hur Y-G, Lee H, Kim S, Cho J-E, Chang J, et al. Discrimination between Active and Latent Tuberculosis Based on Ratio of Antigen-Specific to Mitogen-Induced IP-10 Production. *Journal of Clinical Microbiology*. 2015;53(2):504-10.
30. McInnes MDF, Moher D, Thombs BD, McGrath TA, Bossuyt PM, Clifford T, et al. Preferred Reporting Items for a Systematic Review and Meta-analysis of Diagnostic Test Accuracy Studies: The PRISMA-DTA Statement. *JAMA*. 2018;319(4):388-96.
31. Whiting PF, Rutjes AW, Westwood ME, Mallett S, Deeks JJ, Reitsma JB, et al. QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. *Ann Intern Med*. 2011;155(8):529-36.
32. Macaskill P, Gatsonis C, Deeks J, Harbord R, Takwoingi Y. Analysing and presenting results. In: Deeks JJ, Bossuyt PM, Gatsonis C, editors. *Cochrane Handbook for Systematic Reviews of Diagnostic Test Accuracy*; 2010.
33. Rutter CM, Gatsonis CA. A hierarchical regression approach to meta-analysis of diagnostic test accuracy evaluations. *Stat Med*. 2001;20(19):2865-84.
34. Harbord RM, Whiting P. metandi: Meta-analysis of diagnostic accuracy using hierarchical logistic regression. *Stata Journal*. 2009;9:211-29.

35. Higgins JPT, Thompson SG. Quantifying heterogeneity in a meta-analysis. *Statistics in Medicine*. 2002;21(11):1539-58.
36. Deeks JJ, Macaskill P, Irwig L. The performance of tests of publication bias and other sample size effects in systematic reviews of diagnostic test accuracy was assessed. *J Clin Epidemiol*. 2005;58(9):882-93.
37. Jenkins HE, Yuen CM, Rodriguez CA, Nathavitharana RR, McLaughlin MM, Donald P, et al. Mortality in children diagnosed with tuberculosis: a systematic review and meta-analysis. *Lancet Infect Dis*. 2017;17(3):285-95.
38. Holm LL, Rose MV, Kimaro G, Bygbjerg IC, Mfinanga SG, Ravn P, et al. A comparison of interferon-gamma and IP-10 for the diagnosis of tuberculosis. *Pediatrics*. 2014;134(6):e1568-75.
39. Uplekar M, Weil D, Lonroth K, Jaramillo E, Lienhardt C, Dias HM, et al. WHO's new end TB strategy. *Lancet*. 2015;385(9979):1799-801.
40. Guo SJ, Jia LQ, Hu QJ, Long HY, Pang CS, Wen FQ. Diagnostic accuracy of interferon gamma-induced protein 10 for tuberculosis: a meta-analysis. *International Journal of Clinical and Experimental Medicine*. 2014;7(1):93-100.
41. Ruhwald M, Bodmer T, Maier C, Jepsen M, Haaland MB, Eugen-Olsen J, et al. Evaluating the potential of IP-10 and MCP-2 as biomarkers for the diagnosis of tuberculosis. *European Respiratory Journal*. 2008;32(6):1607-15.
42. Ruhwald M, Aabye MG, Ravn P. IP-10 release assays in the diagnosis of tuberculosis infection: current status and future directions. *Expert Review of Molecular Diagnostics*. 2012;12(2):175-87.
43. Mandalakas AM, Kirchner HL, Walzl G, Gie RP, Schaaf HS, Cotton MF, et al. Optimizing the detection of recent tuberculosis infection in children in a high tuberculosis-HIV burden setting. *Am J Respir Crit Care Med*. 2015;191(7):820-30.
44. Vanini V, Petruccioli E, Gioia C, Cuzzi G, Orchi N, Rianda A, et al. IP-10 is an additional marker for tuberculosis (TB) detection in HIV-infected persons in a low-TB endemic country. *Journal of Infection*. 2012;65(1):49-59.
45. Aabye MG, Ruhwald M, PrayGod G, Jeremiah K, Faurholt-Jepsen M, Faurholt-Jepsen D, et al. Potential of interferon-gamma-inducible protein 10 in improving tuberculosis diagnosis in HIV-infected patients. *European Respiratory Journal*. 2010;36:1488-90.

Tables

Table 1. The main characteristics of included studies.

Author	Year	Country	TB high-burden country	Study design	Age	Number of participants	TB site	Non-TB status	IP-10 cut-off	Reference standard
Y [7]	2018	China	yes	prospective cohort	children and adult	41	PTB	healthy controls	not report	A, B
Y [7]	2018	China	yes	prospective cohort	children and adult	242	PTB	healthy controls	not report	A, B
Y [7]	2018	China	yes	prospective cohort	children and adult	174	PTB	healthy controls	not report	A, B
feldt T [3]	2016	Denmark	no	prospective cohort	adult	225	PTB	healthy controls	2300 pg/ml	B
R [14]	2016	South Africa	yes	prospective cohort	adult	55	PTB	non-TB controls (other respiratory diseases)	> 746.6 pg/ml	B
Y [15]	2015	Republic of Korea	no	cross-sectional	adult	457	PTB	healthy controls	68.5 pg/ml	A
Y [16]	2013	Egypt	no	prospective cohort	adult	30	PTB	healthy controls	not report	B
X [17]	2012	China	yes	cross-sectional	adult	334	PTB	healthy controls	589 pg/ml	A, B/C
Y [18]	2012	Republic of Korea	no	cross-sectional	adult	78	PTB	healthy controls	2811/119.5pg/ml	A, B
r BSA [9]	2012	India	yes	retrospective case control	adult	277	PTB	healthy controls	not report	A, B
[20]	2018	China	yes	prospective cohort	adult	100	PTB	healthy controls	631.4 pg/ml	A, B
a MPL [1]	2018	Italy	no	retrospective cohort	children and adult	47	PTB	non-TB controls (other non-TB pulmonary infections)	> 6557 pg/ml	A, D
ME [22]	2018	Chile	no	prospective cohort	children and adult	61	PTB	household contacts (QFT -)	> 42.6 pg/ml	A, B
aphithak [23]	2017	Thailand	yes	prospective cohort	adult	87	PTB	healthy controls	6319, 2372, 6964 pg/ml	A, B, D
IA [24]	2015	Uganda	yes	prospective cohort	children and adult	168	PTB	household contacts (TST and QFT -)	4858 pg/ml, 0.47	A
land I [5]	2015	Norway	no	prospective cohort	adult	58	PTB	non-TB controls (HIV-infected)	2532 pg/ml	A, B
EL [26]	2015	Uganda	yes	prospective cohort	children	52	PTB	healthy controls	209.1 pg/ml	A, B
el I [27]	2014	Spain	no	retrospective cohort	children	149	PTB	healthy controls	2400 pg/ml	A
S [28]	2012	China	yes	cross-sectional	children and adult	142	PTB	healthy controls	451.3 pg/ml	A, B
H [29]	2015	Republic of Korea	no	cross-sectional	adult	59	PTB	healthy controls	0.7906	A, C

Abbreviations: TB, tuberculosis; PTB, pulmonary tuberculosis; QFT, QuantiFERON-TB Gold test; TST, tuberculin skin test; HIV: human immunodeficiency virus; IP-10: interferon gamma-induced protein 10; A: MTB positive cultures; B: sputum smear microscopy and/or acid-fast bacilli (AFB) positive; C: pathological examination; D: genexpert MTB/RIF.

Table 2. Baseline data of included studies.

Author	IP-10 method	IP-10 condition	HIV-infection	Sensitivity (%)	Specificity (%)	TP	FP	FN	TN
Zhao Y [7]	multiplex cytokines assay	unstimulated	no	90.9	100	30	0	3	8
Zhao Y [7]	two cytokines assay	unstimulated	no	90.14	86.21	192	4	21	25
Zhao Y [7]	two cytokines assay	unstimulated	no	60.45	100	81	0	53	40
Blauenfeldt T [13]	ELISA	stimulated (TB Ag)	some	82	97	53	5	12	155
Jacobs R [14]	multiplex cytokines assay	unstimulated	some	86	73	19	9	3	24
Lee K [15]	ELISA	unstimulated	no	70.1	88.3	141	30	60	226
Azab NY [16]	ELISA	stimulated (TB Ag)	no	100	60	10	8	0	12
Wang X [17]	multiplex cytokines assay	stimulated (TB Ag)	no	86	85.9	153	22	25	134
Hong JY [18]	ELISA	stimulated (TB Ag)	no	97.8	87.5	45	4	1	28
Hong JY [18]	ELISA	unstimulated	no	87.5	90.5	21	2	3	19
Kabeer BSA [19]	ELISA	stimulated (TB Ag)	no	91.4	48	160	52	15	48
Luo J [20]	multiplex cytokine assay	unstimulated	no	86	57.14	43	21	7	28
Manna MPL [21]	multiplex cytokine assay	stimulated (TB Ag-nil)	no	80.77	85	21	3	5	17
Balcells ME [22]	multiplex cytokine assay	stimulated (TB Ag-nil)	no	44.1	88.9	15	3	19	24
Nonghanphithak D [23]	ELISA	stimulated (TB Ag)	no	79.2	87.2	38	5	10	34
Nonghanphithak D [23]	ELISA	unstimulated	no	95.8	94.9	46	2	2	37
Nonghanphithak D [23]	ELISA	stimulated (TB Ag-nil)	no	58.3	97.4	28	1	20	38
Biraro IA [24]	ELISA	stimulated (TB Ag)	some	83.3	84.9	85	10	17	56
Biraro IA [24]	ELISA	stimulated (TB Ag/mitogen ratio)	some	96.7	78.7	99	14	3	52
Wergeland I [25]	multiplex cytokines assay	unstimulated	yes	100	92.3	6	4	0	48
Petrone L [26]	ELISA	unstimulated	no	79	93.9	15	2	4	31
Latorre I [27]	ELISA	stimulated (TB Ag)	no	66.7	76.7	8	32	4	105
Wang S [28]	ELISA	stimulated (TB Ag)	no	89.4	81.6	59	14	7	62
Jeong YH [29]	multiplex cytokines assay	stimulated (TB Ag/mitogen ratio)	no	93.9	100	31	0	2	26

Abbreviations: IP-10, Interferon gamma-induced protein 10; ELISA: enzyme-linked immuno sorbent assay; TB Ag: M. tuberculosis specific antigens; TB Ag-nil: MTB-specific antigen stimulated minus unstimulated levels; TP: true positive, FP: false positive, FN: false negative, TN: true negative.

Table 3. Heterogeneity Assessment.

Covariate	Studies	Sensitivity (95%)	Specificity (95%)	P Value (Bivariate Model)
TB high-burden country	Yes	14	0.86 (0.81-0.92)	0.83
	No	10	0.85 (0.76-0.93)	
Design type	Cohort	17	0.84 (0.78-0.90)	0.55
	Cross-sectional/case-control	7	0.89 (0.82-0.96)	
Age	Adults	14	0.88 (0.82-0.94)	0.59
	Children and adults/children	10	0.83 (0.74-0.91)	
IP-10 method	Multiplex cytokines assay	10	0.84 (0.75-0.92)	0.73
	ELISA	14	0.87 (0.81-0.93)	
IP-10 condition	Stimulated	14	0.86 (0.80-0.92)	0.72
	Unstimulated	10	0.86 (0.78-0.93)	
HIV-infected	Yes/some	5	0.90 (0.82-0.98)	0.53
	No	19	0.84 (0.79-0.90)	

Additional Files

Supplementary Table

Pubmed, Web of science, Embase and Cochrane library strategies.

Supplementary Figure S1

Methodological quality of the included articles.

Supplementary Figure S2

Deeks' funnel plot the included articles.

Figures

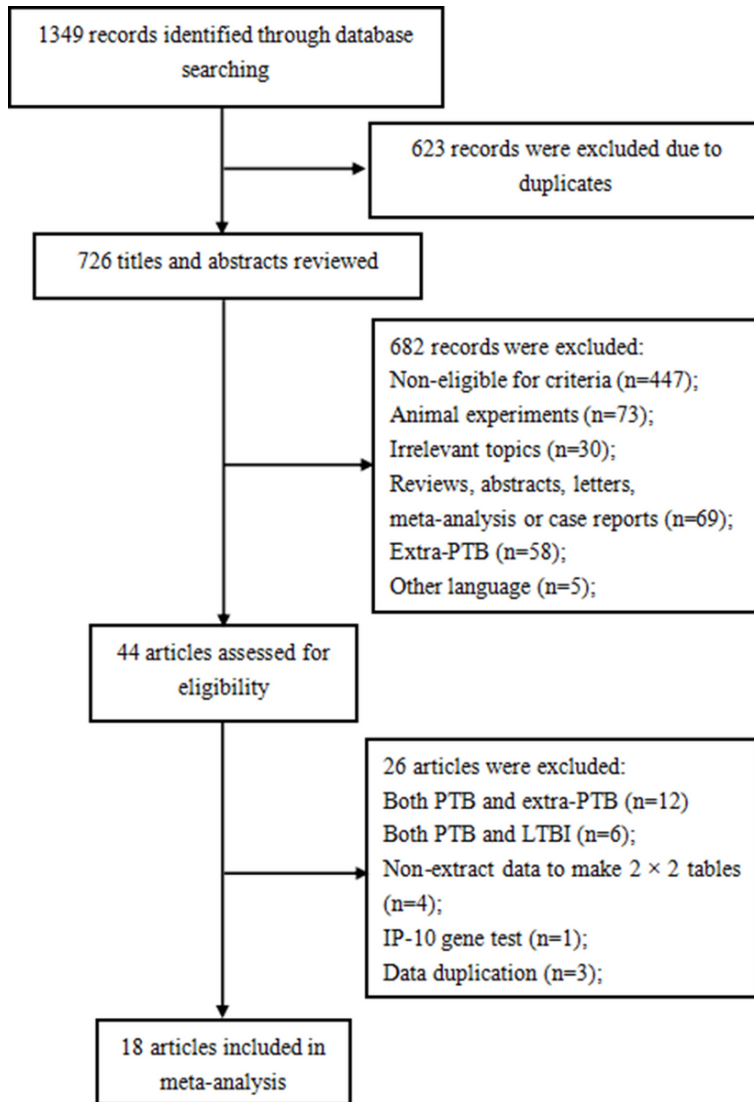


Figure 1

Flow chart of the process of the search strategy for study selection.

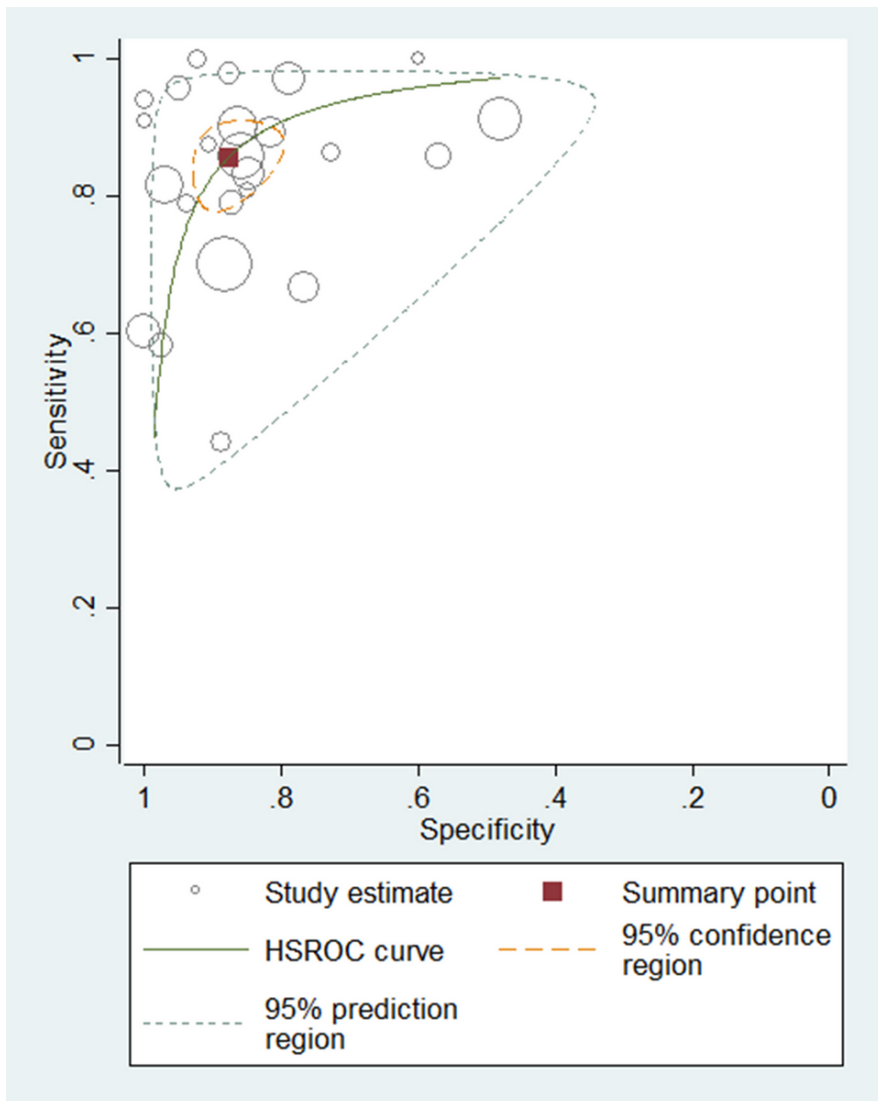


Figure 2

The HSROC curve for assessment of IP-10 for PTB.

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [SupplementaryFigure1.jpg](#)
- [SupplementaryFigure2.jpg](#)
- [Supplementarytable.pdf](#)