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Differential metabolism -associated gene expression of duck pancreatic cells in response to two subtype s of duck hepatitis A virus type 1

Shaohua SHI
Pourlty disease
Zhen CHEN
Poultry disease
Yu HUANG (huangyu_815@163.com)
Cuiqin HUANG
Poultry disease
Rongchang LIU
Poultry disease
Longfei CHENG
Poultry disease
Guanghua FU
Poultry disease
Hongmei CHEN
Poultry disease
Chunhe WAN
Poultry disease
Qiuling FU
Poultry disease

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Abstract

Background: Duck hepatitis A virus type 1 (DHAV-1) causes a highly contagious disease in domestic ducklings, which is traditionally characterized by lesions in the liver and rarely in the pancreas. However, several outbreaks of DHAV-1, which were characterized by yellow coloration and hemorrhage in pancreatic tissues, have occurred in China. Genomic sequencing indicated variation rates of 3.4-6.5% in the genome of the novel DHAV-1 compared with those of DHAV-1. The antigenic relationship between the novel DHAV-1 and DHAV-1 indicated large variation. Therefore, the novel DHAV-1 was named as DHAV-1 subtype (DHAV-1s). However, the mechanism involved in infection of DHAV-1s is still unclear.

Results: In the present study, transcriptome analysis of duck pancreas infected with DHAV-1 and DHAV-1s was carried out. Following deep sequencing with Illumina-Solexa, a total of 53.9 Gb clean data were obtained from the cDNA library of the pancreas and a total of 29,597 unigenes with an average length of 993.43 bp were generated following *de novo* sequence assembly. The expression levels of D-3-phosphoglyceratedehydrogenase, phosphoserine aminotransferase, phosphoserine phosphatase, which are involved in glycine, serine and threonine metabolism pathway, were significantly downregulated in the DHAV-1s-infected group compared with those of the DHAV-1-infected group.

Conclusion: These findings provide information regarding differential expression levels of metabolism-associated genes between the novel DHAV-1s and DHAV-1, indicating that intensive metabolism disorders may contribute to different phenotypes of DHAV-1-infection.

Background

Duck viral hepatitis (DVH) is a highly fatal contagious disease of ducklings, characterized by clinical signs of opisthotonos and by liver hemorrhagic lesions [1]. This causes mortality rates of approximately 95% in ducklings within a week. The causative agents of DVH include duck hepatitis virus type 1 (DHV-1), DHV-2 and DHV-3. Recently, DHV-1 along with two other types was classified as the species *Avihepatovirus* A of the genus *Avihepatovirus* in the family *Picornaviridae* and was renamed as duck hepatitis A virus (DHAV)[2]. DHAV is divided into three serotypes as follows: DHAV-1, which is the classical type and is the most widespread strain, followed by DHAV-2 and DHAV-3.

To the best of our knowledge, DHAV-1 is distributed worldwide and DHAV-2 has been isolated only in Taiwan to date [3, 4]. The presence of DHAV-3 was mainly reported in Korea and China [5-8]. Generally, the lesion in the liver is a typical lesion of DVH caused by DHAV-1. However, Guérin et al. reported that several DHV-infections of young Muscovy ducklings exhibited nervous symptoms and pancreatitis in 2005 [9]. Several outbreaks of DHAV-1, which were characterized by yellow coloration and hemorrhage in pancreatic tissues, have occurred in China in 2011 [10]. Genomic sequencing indicated a variation rate of 3.4-6.5% in the genome of novel DHAV-1 compared with that of DHAV-1 [11]. The antigenic association between the novel DHAV-1 and the classical type DHAV-1 indicated large variation. The novel DHAV-1 was subsequently named as duck hepatitis A virus type 1 subtype (DHAV-1s)[12]. The mechanism involved in

infection of DHAV-1s is still unclear. The host response to the DHAV-1s infection requires experimental validation.

The host-pathogen interaction is a complex and dynamic biological system with its outcome depending on the microbial pathogen properties to establish infection and the ability of the host to control infection [13]. However, several aspects of DHAV-1s-host interactions remain to be clarified. Transcriptomics is the analysis of genome-wide RNA expression and is an available approach to characterize host and pathogen processes in infectious diseases. This method also provides a clear understanding of the host defense mechanisms during the viral infection. To date, the data on the transcriptome profile of the duck pancreatic tissue and its response to viral infection at the transcriptome level are lacking. Moreover, the differences of host responses to DHAV-1 and DHAV-1s have not been reported to date.

In the present study, transcriptome sequencing was applied to determine the gene transcriptome pattern of duck pancreatic tissues infected with DHAV-1 and DHAV-1s. Dramatic differences were observed in the genes involved in the glycine, serine and threonine metabolism pathway, indicating that intensive metabolism disorders may contribute to the different phenotypes of DHAV-1-infection.

Results

Gross lesions of DHAV-1 and DHAV-1s-infected ducklings

The infection experiments were conducted with DHAV-1s GD1206 and DHAV-1 FZ86 strains at the dose of $10^{5.0}$ ELD50 per ducks. The ducks infected with the DHAV-1 FZ86 strain indicated clinical symptoms of depression, lethargy, and opisthotonos. The major lesions that were typically enlarged with petechial and ecchymotic hemorrhages in the liver and kidney were observed during necropsy. No visible lesions were noted in the pancreas. In contrast to these observations, the ducklings infected with the DHAV-1s GD1206 strain demonstrated no typical signs during the course of infection. The major affected organ in the DHAV-1s group was the pancreas, which exhibited lesions of yellowing or hemorrhage. No hemorrhagic change was noted in the liver of the DHAV-1s-infected ducklings. No gross lesions were observed in the mock-infected control group. The pancreatic samples were DHAV-1- or DHAV-1s-positive in the DHAV-1- or DHAV-1s-group as determined by RT-PCR analysis.

RNA-seq and assembly of transcriptome data from the pancreas of the ducklings

Following Illumina-Solexa deep sequencing, a total of 183 M raw reads were obtained in the cDNA library derived from pancreatic tissues. The removal of low-quality reads (i.e., reads containing only adaptors and empty reads) resulted in the identification of clean reads with total residues of 53.9 Gb clean data. Following *de novo* sequence assembly, a total of 29,597 unigenes with an average length of 993.43 bp were generated. BLAST and ORF analyses indicated that only 9,387 targets matched the known genes among the total number of unigenes. The sequencing data from the GD1206- and FZ86-infected groups were submitted to the NCBI database (accession number: SRR7239978, SRR7239979, SRR7239984, SRR7239985, SRR7239988, and SRR7239989).

The heatmaps indicated the top differentially expressed genes (DEGs) and the classification of gene expression profiles in different subtype DHAV-1-infection (Figures 1). In order to obtain a global view of the change in duck gene expression among different experimental groups, three paired comparisons (DHAV-1 vs. control, DHAV-1s vs. control, DHAV-1s vs. DHAV-1) were performed. RNA-seq analysis detected 3,340 and 5,919 genes, which were expressed at significantly different levels in DHAV-1- and DHAV-1s-infected animals, respectively compared with those noted in the control group (P<0.05). DHAV-1 infection contributed to the differential expression of 2,031 genes that were upregulated and 1,309 genes that were downregulated in the pancreatic tissues of the infected animals compared with those noted in the control ducklings. Moreover, the expression levels of 3,308 genes were upregulated and those of 2,611 genes were downregulated in the DHAV-1s-infected animals. A total of 1,913 genes were differentially expressed in DHAV-1s-infected animals compared with the DHAV-1-group. Specifically, the expression levels of 834 genes were downregulated and those of 1,079 genes were upregulated (Figure 2).

Functional analysis of the transcriptome data derived from the pancreatic tissues of different type DHAV-infected animals

We performed GO enrichment analysis by functional annotation clustering of DEGs and annotated DEGs into three groups, including biological processes (BP), cellular components (CC), and molecular function (MF).

The putative functions of the unigenes in the libraries derived from the DHAV-infected ducklings were analyzed using GO. The analysis of the GO categories of the DHAV-1-group indicated that the differentially expressed genes were mapped to 61 categories of BP, CC and MF (Figure 3A). The categories of BP included the genes mainly involved in the positive regulation of interleukin-12 production, myeloid leukocyte activation and T cell proliferation. The majority of the corresponding genes in the CC categories were involved in the external side of plasma membrane, the oligosaccharyltransferase complex and the integral component of the membrane. The majority of the corresponding genes in the MF categories were involved in the non-membrane spanning protein tyrosine kinase activity, antioxidant activity and calcium-dependent phospholipid binding.

The putative functions of the unigenes in the pancreatic libraries of the DHAV-1s infection were analyzed using GO. The analysis of the GO categories indicated that the differentially expressed genes were mapped to 61 categories including BP, CC and MF (Figure 3B). The majority of the corresponding genes in the BP categories were involved in cholesterol efflux, positive regulation of cell cycle and translation. The categories of the CC included genes that were mainly involved in the cytosolic large ribosomal subunit, extracellular space and ribosome. The majority of the genes of the MF categories included genes involved in the structural constituents of the ribosome, cytokine activity and NADH dehydrogenase (ubiquinone) activity.

The putative functions of the unigenes in the pancreatic libraries of DHAV-1s-infected ducklings were compared with those of the DHAV-1-infected ducklings and were analyzed using GO. The analysis of the GO categories indicated mapping of differentially expressed genes to 61 categories of BP, CC and MF

(Figure 3C). The BP category included mainly genes that were involved in the oxidation-reduction process, the negative regulation of apoptotic process and the myeloid leukocyte activation. The CC category included mainly genes that were involved in the extracellular space, protein-extracellular matrix and ribosome. The majority of the corresponding genes in the MF category were involved in the antioxidant activity, G-protein coupled peptide receptor activity and structural constituents of the ribosome.

Pathway analysis of DEGs based on KEGG following different DHAV-1-infection

The KEGG database was used to analyze specific pathways in order to further define DEG function in the duckling pancreatic tissue following different DHAV-1-infections. The top 20 enrichment KEGG pathways were listed in Figure 4 according to their Q-value ($Q< 0.05$) (Table 1).

A total of 6 functional categories were identified that played important roles in the DHAV-1 FZ86- and DHAV-1s GD1206-mediated infections. These categories were mainly classified into immune system categories, including the Toll-like receptor signaling pathway. However, significant KEGG enrichment in the DHAV-1s group was also involved in the metabolism function, including the glycine, serine and threonine metabolism pathway.

Verification of DEG identification by real-time RT-PCR

In order to verify the differential gene expression levels obtained from the transcriptome sequencing data, we analyzed the expression levels of 10 genes that were involved in immune and metabolism-associated functions. These genes were also involved in host immune defense responses and metabolic function noted in the two phenotype DHAV-1 infection groups. The genes examined were as follows: GNMT-like, GCAT, CBS, PHGDH, SERCA, PLCγ, TLR2, TLR4, TLR7 and IFNα. They were differentially expressed compared with the control ones ($P<0.05$), indicating the reliability of the transcriptome sequencing data (Table 2).

Discussion

Duck virus hepatitis is mainly caused by DHAV-1 and is a malignant disease noted in ducklings, which affects severely the duck industry worldwide. Previous studies performed on DHAV have focused only on the pathogenicity and the host response to DHAV-1 infection. The comparative pathogenicity of DHAV-1 and DHAV-1s strains in the ducklings of different species has been well documented [14, 15]. However, the different mechanisms between DHAV-1 and DHAV-1s strains remain unknown.

In the present study, the ducklings that were inoculated with DHAV-1FZ86 indicated massive haemorrhages on the liver surface. The ducklings that were inoculated with the DHAV-1s GD1206 strain exhibited loss of appetite, lying prone, diarrhea and depression. The gross lesion of the yellowed or hemorrhagic pancreatitis was observed in DHAV-1s-infected ducklings. However, no consistent nervous disorders were observed in the present study, which was not in line with the observations performed by Guérinet al. [9]. Cha et al. described that a DHAV-3 strain induced only liver discoloration without

hemorrhagic mottling, lymphocyte infiltration and bile duct hyperplasia as determined by histology of the lesions [16]. These studies revealed the diversity in the pathogenicity of DHAV.

Transcriptome analysis is a promising tool that can provide a comprehensive understanding of the molecular mechanisms involved in specific biological processes and diseases [17]. Previous studies have described the transcriptome profile of DHAV-, Reovirus- and DHBV-infection in ducks [18-21]. In the present study, transcriptome sequencing was employed to explore and compare the gene expression patterns of infection with different subtype DHAV-1 in duck pancreatic tissues, with the aim to compare different molecular events during DHAV-1s infection.

Following analysis of the clean reads with BLAST and ORF, a total of 9,387 matched known genes were obtained. The DEGs were annotated and categorized by GO and KEGG signaling pathway analyses, which demonstrated that the majority of these genes in DHAV-1 and DHAV-1s-infection groups were classified into immune system and metabolism categories. The investigation of the transcriptome of DHAV-1-infected and the control ducklings indicated that DHAV-1 infection caused downregulation of the expression of genes associated with metabolic pathways and inhibition of the metabolism of the host cell. In addition, upregulation of the expression levels of the immune-associated genes was noted that resulted in inhibition of viral replication and progression of viral infection [22]. Similarly, in the present study, upregulation of the expression of immune genes and downregulation of certain metabolism-related genes were observed in the DHAV-1s infection group compared with the corresponding levels noted in the control group. However, significant KEGG enrichment was noted in the DHAV-1s infection group compared with DHAV-1 infection group that was mainly involved the metabolism function, including the glycine, serine and threonine metabolism pathway. The results suggested that the differences in the metabolism function of the DHAV-infection group may contribute to the different phenotypes of DHAV-1.

D-3-phosphoglycerate dehydrogenase (PHGDH), phosphoserine aminotransferase（PSAT）and phosphoserinephosphatase (PSP) are involved in the three-enzyme step-serine biosynthesis [23]. Initially, 3-phosphoglycerate is converted to 3-phosphohydroxypyruvate by the enzyme PGDH. Subsequently, PSAT converts 3-phosphohydroxypyruvate to 3-phosphoserine. Finally, 3-phosphoserine is converted to L-serine by PSP [24]. L-serine is an important precursor involved in various processes, such as synthesis of proteins and phospholipids as well as the synthesis of tetrahydrofolate metabolites and specific amino acids, namely glycine, cysteine and D-serine [25, 26]. Previous studies have shown that L-serine supplementation can attenuate alcoholic fatty liver formation in mice and rats [27]. In the present study, the expression levels of the PHGDH, PSAT and PSP genes were significantly downregulated in the DHAV-1s-infected ducklings compared with those noted in the DHAV-1 group, indicating that serine metabolism disorders were involved in DHAV-1s infection.

L-serine dehydratase (SDH) catalyzes the deamination of L-serine to yield ammonia and pyruvate. This enzyme uses L-threonine as a substrate to yield 2-oxobutanoate, which is a part of the valine, leucine and isoleucine biosynthetic pathway. The downregulation of serine dehydratase levels suggested that the

pathway of serine to pyruvate was impaired and that it further affected the valine, leucine and isoleucine biosynthetic pathway [28-30].

The innate immune system is essential for recognizing pathogen infections and for the establishment of effective host defense. During the process of innate immune response, highly conserved pathogen-associated molecular patterns (PAMP) are detected by pattern recognition receptors (PRR)[31]. Three classes of PRRs have been shown to be involved in the recognition of virus-specific components in innate immune cells, namely Toll-like receptors (TLRs), retinoic acid-inducible gene I (RIG-I)-like receptors (RLRs) and nucleotide oligomerization domain (NOD)-like receptors s (NLRs)[32]. In the present study, almost similar innate immune responses were observed in both subtypes from DHAV-1 infected animals.

TLRs play a critical role in innate immune responses. In recent years, the role of innate immunity and its interaction with adaptive immunity has been mainly explored. In the TLR pathway, TLR2 plays a critical role in the induction of innate and inflammatory responses [33, 34]. Although TLR2 recognizes various bacterial components, recent studies have indicated that TLR2 is triggered by the HCV protein core and NS3 leading to the activation of inflammatory cells [35, 36]. In the present study, the expression levels of the TLR2 gene were 10.63-or 7.62-fold higher than those of the control group ducklings in the DHAV-1s/DHAV-1 infection groups, respectively, indicating that TLR2 may be involved in the host response to DHAV infection. TLR4 and CD14 have recently been shown to be the major lipopolysaccharide (LPS) receptors. Mutations in mouse and human TLR4 were found to be associated with hyporesponsiveness to LPS and confer an increased risk of gram-negative infections [37-39]. In addition to the interaction with LPS, the TLR4/CD14 complex interacts with other viruses and proteins, such as the respiratory syncytial virus and fibrinogen [40-42]. In the present study, the TLR4/CD14 expression levels in the DHAV-1s and DHAV-1 infection groups were highly upregulated compared with those of the non-infected ducklings, indicating that TLR4/CD14 may be involved in the host response to DHAV-1s and DHAV-1 infection. Surprisingly, TLR4 expression was significantly higher in the DHAV-1s and DHAV-1 groups compared with that in the mock group with an increase of 51.63-and 38.59-fold, respectively. TLR7 can detect single stranded RNA (ssRNA) molecules and induce pro-inflammatory factors, such as the type I interferon to stimulate the body's non-specific immune response. Activation of TLR7 initiates downstream signaling cascades via induction of transcription factors, such as IRF7. This induces the production of pro-inflammatory cytokines and chemokines that are involved in various viral infection outcomes, including spontaneous clearance and viral persistence [43]. The potential implication of TLR7 in the innate immune response against hepatitis C virus was postulated [44, 45]. Nazmi et al. suggested that the TLR7 response following Japanese encephalitis viral infection promoted type-1 interferon production and generation of an antiviral state, contributing to the protective effect of systemic infection [46]. In the present study, upregulation of the expression levels of TLR7, IRF7 and IFN-β was noted in the DHAV-1s and DHAV-1 groups, which was consistent with the findings described in previous studies [46, 47].

Conclusions

In the present study，transcriptome analysis of pancreatic tissues derived from DHAV-1 and/or DHAV-1a-infected ducks was reported. The infection of DHAV-1a caused yellowing and hemorrhagic lesions in the pancreatic tissues of the ducklings and was associated with differences in the expression levels of D-3-phosphoglyceratedehydrogenase, phosphoserine aminotransferase, phosphoserine phosphatase, which are involved in the glycine, serine and threonine metabolism pathway. The expression levels of the aforementioned genes were significantly down-regulated in the DHAV-1a-infected group compared with those from DHAV-1-infected group, indicating that intensive metabolism disorder may contribute to the different phenotypes of DHAV-1 infection.

Methods

Animal Experiment

One-day-old male Muscovy ducklings (*Cairina moschata*) were purchased from a commercial hatchery farm. Each duckling had free access to food and water and shared the same environmental conditions at 35℃. The temperature was decreased by 1℃ every day. The light was continuously available during the animal experiments. Serum samples and cloacal swabs were collected from the ducklings prior to viral challenge. The presence of antibody and antigen was determined using indirect enzyme linked immunosorbent assay (ELISA) and reverse transcription polymerase chain reaction (RT-PCR) methods [10], respectively in order to confirm that the animals were DHAV-1 and DHAV-1s negative. Four days later, 60 5-day-old healthy Muscovy ducklings, which were negative for DHAV-1 and DHAV-1s antigens and antibodies, were randomly divided into three groups (n=20 for each group). The ducklings in each group were challenged intramuscularly with $10^{5.0}$ELD50 (50% egg lethal dose) DHAV-1 FZ86 strain and DHAV-1s GD1206 strain in 200 μl phosphate buffer saline (PBS) or mock-infected with PBS. Each group of ducklings was housed in separate isolators (Suzhou Fung's laboratory Animal Equipment Co., Ltd.) with free access to food and water throughout the experimental period. Each group of ducklings shared the same environmental conditions at an experimental temperature of 31℃. The temperature was reduced by 1℃ every day until 25℃. The light was continuously available during the animal experiments. The daily clinical symptoms were monitored.

At 24 h post-inoculation, six ducklings in each group were randomly selected. The ducklings were administered sodium pentobarbital (40 mg/kg bodyweight; Sigma-Aldrich, USA) intravenously and subsequently the animals were humanly sacrificed under narcosis [47]. The pancreatic samples were harvested and pooled into two tubes (3 samples/tube, 0.5 g each sample) for transcriptional analysis. Concomitantly, the pancreatic samples were also detected by RT-PCR in order to confirm DHAV-1- or DHAV-1s infection. The remaining 14 ducklings of each group were observed for three days. At the end of the observation period, the ducklings were administered intravenously sodium pentobarbital (40 mg/kg bodyweight; Sigma-Aldrich, USA) and subsequently humanly sacrificed under narcosis [47]. The gross lesions were observed.

Host gene expression associated with DHAV-1s infection

RNA extraction, library construction, and sequencing

Total RNA was extracted using RNAiso Plus Total RNA extraction reagent (TaKaRa, Shiga, Japan) according to the manufacturer's instructions. RNA degradation and contamination was monitored on 1.0% agarose gels. RNA purity was confirmed using the NanoPhotometer® spectrophotometer (IMPLEN, CA, USA). RNA concentration was measured using the Qubit® RNA Assay Kit in a Qubit®2.0 Fluorometer (Life Technologies, CA, USA). RNA integrity was assessed using the RNA Nano 6000 Assay Kit of the Agilent Bioanalyzer 2100 system (Agilent Technologies, CA, USA) and subsequently employed as a template for the synthesis of double-stranded cDNAs. Sequencing libraries were generated using NEBNext®Ultra™ RNA Library Prep Kit for Illumina® (NEB, CA, USA) following the manufacturer's recommendations. Finally, index codes were added to the attribute sequences of each sample. The clustering of the index-coded samples was performed on a cBot Cluster Generation System using TruSeq PE Cluster Kit v3-cBot-HS (Illumina) according to the manufacturer's instructions. Following cluster generation, the library preparations were sequenced on an Illumina Hiseq 4000 platform and paired-end reads were generated. Raw reads were filtered and assembled using Trinity software, and then the transcript library was obtained. The RNA-seq data from this study were submitted to the NCBI Sequence Read Archive (SRA) (http://www.ncbi.nlm.nih.gov/sra/).

Transcriptome data analysis

Differential expression analysis of the three groups was performed using the DESeq R package (1.10.1). DESeq provided statistical routine methods for determining differential expression in digital gene expression data using a model, which was based on the negative binomial distribution. The resulting P-values were adjusted using the Benjamini and Hochberg's approach for controlling the false discovery rate. The genes with an adjusted P-value of <0.05, identified using DESeq, were assigned as differentially expressed genes.

GO enrichment analysis and KEGG pathway enrichment analysis

Gene Ontology (GO) enrichment analysis of the differentially expressed genes (DEGs) was implemented by the topGO R packages-based Kolmogorov–Smirnov test. The GO terms with the corrected *P* value less than 0.05 were considered significantly enriched in the differential expressed genes.

The Kyoto Encyclopedia of Genes and Genomes (KEGG) is a database resource for understanding high-level functions and utilities of the biological systems, such as the cell, organism and the ecosystem. Specific molecular-level information is collected notably from large-scale molecular datasets generated by genome sequencing and other high-throughput experimental technologies (http://www.genome.jp/kegg/)[48]. The KOBAS software was used to assess the statistical enrichment of differentially expressed genes in the KEGG pathways [49]. The KEGG terms with a corrected P-value (Q-value) less than 0.05 were considered significantly enriched in the differential expressed genes.

Verification by real-time RT-PCR

The expression levels of the differentially expressed genes were also assessed by real-time RT-PCR in order to confirm the sequencing data. The individual primer sequences of the 10 target genes and of the internal reference gene (GAPDH)[50] were listed in Table 3. Following reverse transcription, a SYBR®Fast qPCR Mix (Takara, Beijing) and a Mastercycler ep realplex (eppendorf) were used according to the manufacturers' instructions. The reaction conditions were as follows: 95°C for 1 min, followed by 40 cycles of 95°C for 10 sec and 60°C for 40 sec. A final solubility curve analysis was completed. The relative mRNA levels were compared to those of the mock-infected ducks and the mean values were calculated using the threshold cycle $2^{-\triangle\triangle CT}$ method [50]. The comparisons of the experimental groups were estimated by the Student's *t*-test with a two-tailed analysis in order to determine significant differences. P-values less than 0.05 were considered significant.

Abbreviations

KEGG: Kyoto encyclopedia of genes and genomes; GO: Gene ontology; RT-PCR: Reverse transcription polymerase chain reaction; ELISA: Enzyme linked immunosorbent assay; DEG: Differentially expressed genes; BLAST: Basic local alignment search tool；ORF: Open reading frame.

Declarations

Ethics Approval and Consent to Participate

All animal experiments were approved by the animal welfare committee of Fujian Academy of Agricultural Sciences (No.1306175FAAS) and complied with the guidelines of Fujian laboratory animal welfare and Fujian Administrative Ethics Committee of Laboratory Animals.

Consent for publication

Not applicable.

Availability of data and materials

The raw data has been deposited in NCBI database, with [DHAV-1-infected group accession number： SRR7239978 and SRR7239979], [Control group accession number：SRR7239984 and SRR7239985], and [DHAV-1a-infected group accession number：SRR7239988 and SRR7239989],

[https://www.ncbi.nlm.nih.gov/sra/?term=SRR7239978,

https://www.ncbi.nlm.nih.gov/sra/?term=SRR7239979,

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Competing interests

The authors declared no conflict of interest.

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preparation.

Author's contribution

SS, CZ and HY conceived and designed the experiments; SS and CZ wrote the original draft preparation; SS, CZ, HC, LR, CL and FG performed the experiments; CH, WC and FQ analyzed the data. All authors read and approved the final manuscript.

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Authors' Information

Institute of Animal Husbandry and Veterinary Medicine, Fujian Academy of Agricultural Sciences, Fuzhou, Fujian, China; Fujian Animal Diseases Control Technology Development Center; Fujian Provincial Key Laboratory for Avian Diseases Control and Prevention, Fuzhou, Fujian, China (Shaohua SHI□Zhen CHEN□ Yu HUANG□Longfei CHENG□Guanghua FU□Hongmei CHEN□Chunhe WAN□Qiuling FU□

College of Life Sciences, Longyan University, Longyan, Fujian, China□Cuiqin HUANG□

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Tables

Figures

Table 1A

Pathway	DEG in Pathway	All Gene in Pathway	P-value	Q-value
Influenza A	55	134	1.58E-10	3.48E-08
Cytokine-cytokine receptor interaction	56	144	1.24E-09	2.73E-07
Protein processing in endoplasmic reticulum	66	187	4.65E-09	1.02E-06
Intestinal immune network for IgA production	20	31	1.03E-08	2.27E-06
Phagosome	53	144	3.26E-08	7.18E-06
Cell adhesion molecules (CAMs)	44	111	3.98E-08	8.76E-06
JAK-STAT signaling pathway	44	120	5.58E-07	0.000123
Toll-like receptor signaling pathway	29	83	0.000132203	0.029085

Table 1B

Pathway	DEG in Pathway	All Gene in Pathway	P-value	Q-value
Cytokine-cytokine receptor interaction	74	144	3.43E-08	8.44E-06
Influenza A	67	134	5.84E-07	0.000144
Toll-like receptor signaling pathway	42	83	5.28E-05	0.012986

Table 1C

Pathway	DEG in Pathway	All Gene in Pathway	P-value	Q-value
Glycine, serine and threonine metabolism	17	52	6.74E-08	9.37E-06
Neuroactive ligand-receptor interaction	26	142	8.65E-06	0.001202
ECM-receptor interaction	22	111	1.18E-05	0.001637
Calcium signaling pathway	24	146	0.000118608	0.016487
Focal adhesion	33	243	0.000311296	0.04327

Table 2

Gene	Abbreviation	Transcriptomics fold change (Log_2FC)		Real-time RT-PCR fold change ($2^{-\Delta\Delta CT}$)	
		DHV-1/ control	DHV-1a/ control	DHV-1/ control	DHV-1a/ control
Glycine N-methyltransferase-like	GNMT-like	-2.08	-5.72	0.210	0.01
Glycine C-acetyltransferase	GCAT	-2.11	-4.80	0.20	0.04
Cystathionine beta-synthase	CBS	-1.87	-5.79	0.25	0.03
Phosphoglycerate dehydrogenase	PHGDH	-2.49	-5.24	0.18	0.02
Sarcoplasmic/endoplasmic reticulum calcium ATPase	SERCA	1.05	1.05	2.46	4.48
1-phosphatidylinositol 4,5-bisphosphate phosphodiesterase gamma	PLCγ	1.58	1.37	4.96	2.95
Toll-like receptor 2	TLR2	2.93	3.41	8.69	12.30
Toll-like receptor 4	TLR4	5.27	5.69	38.05	46.85
Toll-like receptor 7	TLR7	5.81	5.36	45.57	30.27
Interferon alpha	IFNα	2.23	2.62	3.89	8.06

Table 3

Primer name	Sequence（5′- 3′）	Size (
GAPDH [49]	F-ATGTTCGTGATGGGTGTGA	176
	R-CTGTCTTCGTGTGTGGCTGT	
GNMT-like	F-CAGCTGGCTCCTGGCGCTGC	183
	R-GGCTCCTCCTTGCGCCGCTC	
GCAT	F-GCTCCGGCGGCGGCTGGAGG	171
	R-GGGTGGCTCGACAGCCCCAGGTA	
CBS	F-GTACCTCCTGTGCCAGCTGTAGCCA	170
	R-GTGGGAGTAGCGTGGAGACTGAAAA	
PHGDH	F-CCAAGACCCCGGGCTCGCAGCCACC	177
	R-CGTGGGGCTCCCGACCAAGAGGCTG	
SERCA	F-ACGGCCTTCGTCGAGCCCTTCGTCA	186
	R-GGGCACGAGGTCCCTGGCCTTGATG	
PLCγ	F-GTTTTCTGTGATCAGAAATACAC	204
	R-TATTTAAAACCAGTTGTCCTAATTT	
TLR2	F-CTCTTCTTCACGAGGCCACTCCAGG	183
	R-CAGAGCGAGTGGTGCAAGTACGAGC	
TLR4	F-TCACCAGCCTACAAGACCTGCAAGA	184
	R-GGAGGAGGTGGACGGAGGCACTGTA	
TLR7	F-CATGCACTCCCCACTGGAAGTCCCT	150
	R-CTTCCTGGCAGCCTGTGTGCTAAAG	
IFNα	F-GCCCAGATGCGGGACTGTCCACCTG	167
	R-ACACGGCTGCACGATGGGAATCTCC	

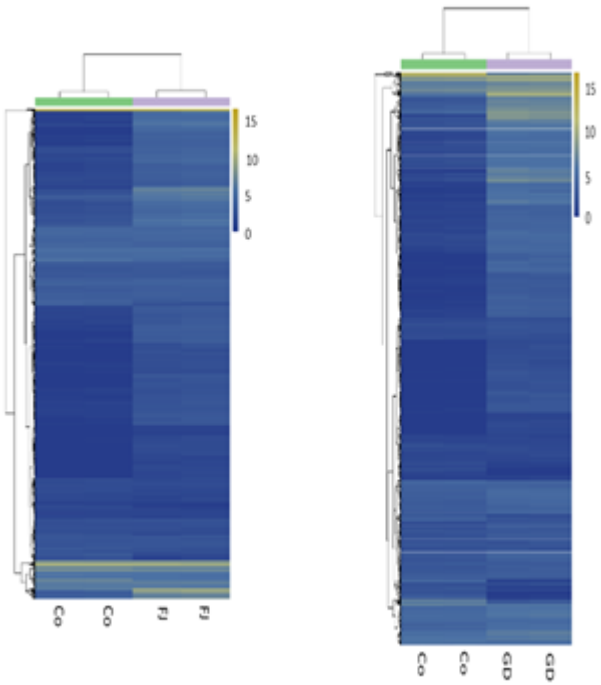

Figure 1

Heat map analysis used to classify gene expression patterns in DHAV-1-infection (A) and DHAV-1a-infection (B). Genes with similar expression patterns were clustered, as shown in the heat maps. Intensity of color indicated gene expression levels that were normalized according to Log2 FPKM value

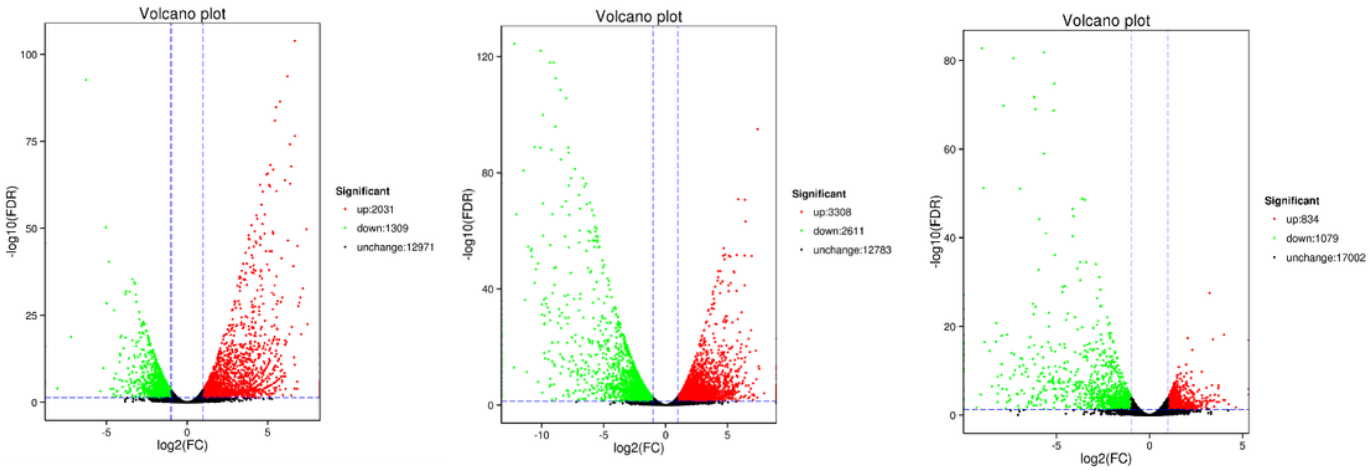

Figure 2

Volcano plots of differentially expressed genes in the pancreas of the ducklings in DHAV-1 vs control (A), DHAV-1a vs control (B) and DHAV-1a vs DHAV-1 (C). Red points represented up-regulated genes, green

points represented down-regulated genes, and blue points represented genes with no significant difference

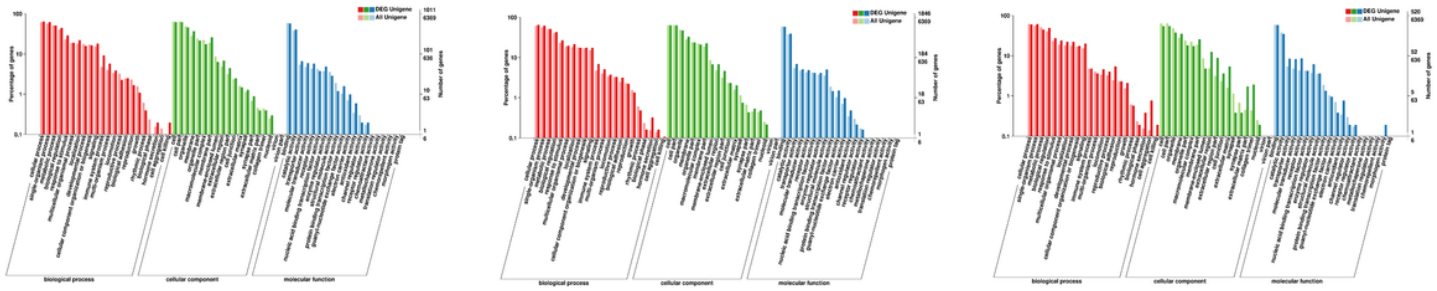

Figure 3

GO classification of differentially expressed genes in the pancreas of the ducklings in DHAV-1 vs control (A), DHAV-1a vs control (B) and DHAV-1a vs DHAV-1 (C). Annotation statistics of differentially expressed genes in the secondary node of GO. The horizontal axis showed secondary nodes of three categories in GO. The left side of vertical axis displayed the percentage of annotated genes versus the total gene number and the right side showed the number of genes

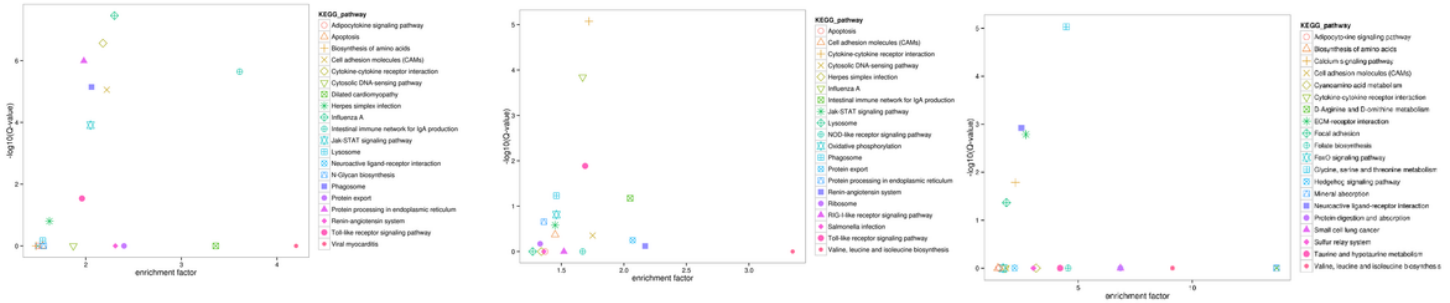

Figure 4

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PCR analysis Figure 1 Heat map analysis used to classify gene expression patterns in DHAV-1-infection (A) and DHAV-1a-infection (B). Genes with similar expression patterns were clustered, as shown in the heat maps. Intensity of color indicated gene expression levels that were normalized according to Log2 FPKM value Figure 2 Volcano plots of differentially expressed genes in the pancreas of the ducklings in DHAV-1 vs control (A), DHAV-1a vs control (B) and DHAV-1a vs DHAV-1 (C). Red points represented up-regulated genes, green points represented down-regulated genes, and blue points represented genes with no significant difference Figure 3 GO classification of differentially expressed genes in the pancreas of the ducklings in DHAV-1 vs control (A), DHAV-1a vs control (B) and DHAV-1a vs DHAV-1 (C). Annotation statistics of differentially expressed genes in the secondary node of GO. The horizontal axis showed secondary nodes of three categories in GO. The left side of vertical axis displayed the percentage of annotated genes versus the total gene number and the right side showed the number of genes Figure 4 Top 20 KEGG pathways enriched in the pancreas of the ducklings in DHAV-1 vs control (A), DHAV-1a vs control (B) and DHAV-1a vs DHAV-1 (C)

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