Green synthesis of iron oxide nanoparticles using *Ceratonia siliqua* L. aqueous extract: Optimization, characterization, stabilization and evaluation of its antibacterial activity against gram-positive and gram-negative bacteria

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Research Article

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Abstract

This study focused on the optimization of reaction parameters for a simple and reliable method for the green synthesis of iron oxide nanoparticles (gIONPs). The potential of aqueous extract of the Ceratonia siliqua L. (carob pod) as a reducing, capping, and stabilizing agent for gIONPs synthesis, and the antibacterial activity of as-synthesized gIONPs was investigated. The synthesis parameters were determined as the concentration of iron ion, the concentration of extract, pH of extract, temperature, stirring rate, and reaction time. The significance of the studied factors in controlling the particle size distribution of nanoparticles was quantitatively evaluated via analysis of variance (ANOVA). Iron oxide nanoparticles were produced at the optimum parameters with an average zeta potential of $+ 40.75 \pm 1.22$ mV, hydrodynamic size of $78 \pm 22$ nm, and a polydispersity value of $0.42 \pm 0.06$, respectively. gIONPs showed the highest inhibition zones of 24 mm and 21 mm in 250 mg/mL concentration against Gram-negative Escherichia Coli and Gram-positive Staphylococcus aureus, respectively. Antibacterial activity was observed when compared with the standard antibiotics. The findings of the study revealed that the highly stable and antibacterial iron oxide nanoparticles can be synthesized through the adjustment of the reaction parameters by eliminating chemicals.

1. Introduction

Nanoparticles have been prepared through a diverse range of synthesis approaches over the last decades and different fundamental principles of synthesis procedures have been investigated to obtain nanoparticles of desired sizes, shapes, and functionalities. Their unique size-dependent thermal, electrical, chemical, and optical properties have enabled their use in fields as medicine and chemical analysis [1].

The green synthesis approach is a promising synthesis procedure in the research and development of materials science and technology due to the biosynthetic pathway of nanoparticles preparation, potentially eliminating the usage of chemicals and making the nanoparticles more biocompatible. Some basic principles of “green synthesis” can be explained by prevention or minimization of waste, reduction of pollutant derivatives, and the use of safer solvent as well as renewable feedstock [2]. The biomaterials extracted from several parts of the plant are mixed with metal precursor solutions at different reaction conditions for nanoparticle synthesis. The biomaterials play several roles such as reducing, capping, and stabilizing agents in the nanoparticle synthesis process. Nanoparticles are synthesized within a few minutes or hours depending upon the type and concentration of biochemicals arising from plant sources. The plant extracts have been proven to possess high efficiency as stabilizing and reducing agents for the synthesis of nanoparticles but detailed investigation on the role of reaction parameters in synthesis is still needed to overcome existing problems in ‘green’ synthesis [3].

Capping agents significantly modify the properties of colloidal suspensions of nanoparticles. The efficacy of colloidal NPs application is linked to high suspension stability. Small changes to the solution (e.g., background salt concentration, temperature, pH), or changes to the nanoparticles (e.g., surface
coating or concentration) can substantially affect stability. There are several parameters that affect the green synthesis of nanoparticles, including pH of the solution, temperature, the concentration of the plant material used, reaction time, and above all the protocols that are used for the synthesis process. The change in the type and characteristic of the synthesized nanoparticles is especially affected by the type of plant material used in the synthesis process [4]. The morphology and surface property of nanoparticles are important parameters for their applications. The better antibacterial action may be attributed to their smaller size and surface charge to come in contact with the microbial cells [5]. An ideal synthesis method should be able to allow reliable adjustment of particle distribution, size, and composition [6].

There are different phases of iron oxides as hematite ($\alpha$-Fe$_2$O$_3$), maghemite ($\gamma$-Fe$_2$O$_3$), goethite ($\alpha$-FeOOH), and magnetite (Fe$_3$O$_4$). The size scale affects the optical, electrical, and biocompatibility properties of these materials [7] [8]. The functional groups of coating materials alter the surface charge, which influences the biological behaviors of iron oxide nanoparticles [9]. The dispersity, size, and surface chemistry of the iron oxide materials are crucial for environmental applications in the aqueous phase due to the porous structure with particle-surface interaction [10]. Iron oxide nanoparticles act as promising antibacterial agents that have high surface areas with crystalline morphologies at a high number of edges and corners and generation oxidative stress by reactive oxygen species [11].

In this study, we report the green synthesis of iron oxide nanoparticles using biochemicals extracted from Ceratonia siliqua. L. (carob pod). Carob pod is rich in sugars as sucrose (437.3 mg/g dry weight), glucose (395.8 mg/g dry weight), and fructose (42.3 mg/g dry weight). The other content is as total phenolics group (13.51 mg gallic acid equivalents [GAE]/g dry weight), proanthocyanidins (0.36 mg GAE/g dry weight), gallotannins (0.41 catechin equivalents [CE]/g dry weight), and flavanols (3.21 mg CE/g dry weight protein). Gallic acid (3.27 mg/g dry weight) is the most abundant phenolic material. Aspartic acid (18.25 mg/g dry weight protein) is the predominant amino acid in the protein fraction. Major minerals are vitamin K, calcium, potassium, and magnesium [12].

The effects of reaction parameters on the synthesis of iron oxide nanoparticles were investigated by the Taguchi method in this study. The Taguchi method is used for evaluating the results of matrix experiments to determine the best levels of experimental parameters. It makes it possible to provide an acceptable formulation using minimum raw materials and time [13]. The signal-to-noise (S/N) ratio and the analysis of variance (ANOVA) were employed to analyze the experimental parameters. The parameters including the concentration of plant extract, the concentration of iron ion, reaction temperature, pH of plant extract, stirring rate, and reaction time were evaluated. The polydispersity value of nanoparticles was measured by a dynamic light scattering (DLS) instrument after each synthesis to analyze the effect of chosen experimental parameters. The zeta potential and UV-Vis spectrum was monitored over 3 months period for analysis of colloidal stability. Furthermore, the antibacterial activity of powder gIONPs were tested against Gram-negative (Escherichia coli) and Gram-positive (Staphylococcus aureus) bacteria.
2. Materials And Methods

Materials

Carob pod was purchased from the local market in Kayseri, Turkey. Ferric chloride (FeCl₃·6H₂O), sodium hydroxide (NaOH) and hydrochloric acid (HCl) were purchased from Merck. Ultrapure water was used in the experiments (Elga Purelab Option-Q 7).

Synthesis and characterization of gIONPs

The method for the extraction of the carob pod was used as similar to the procedure followed in our previous study [14]. 10 gr of carob pod pieces were extracted in 150 mL of ultrapure water for 2 hours. The resultant extract was filtered through Whatman No.1 filter paper and was used as a reducing and capping agent for the green synthesis of iron oxide nanoparticles. 3.8 g of ferric chloride (FeCl₃·6H₂O) was dissolved in 100 mL of ultrapure water for 15 min. The extract to precursor iron solution was mixed at a 1:1 volume ratio. The addition of the plant extract to ferric chloride aqueous solution led to turning initially yellowish solution to a brown color indicating the formation of iron nanoparticles. The resultant colloidal nanoparticle solution was stirred for 3 hours and kept in a +4 °C refrigerator to do stability analysis. Nanoparticles were precipitated by adjusting the pH value of gIONPs colloidal solution to 7 and the pellet was washed with ultrapure water and ethanol twice. It was dried in a laboratory oven at 70°C for 10 h. The brownish-black powder of gIONPs (Fig.S1) was used for characterization and antibacterial analysis.

The characterization of the gIONPs was investigated using X-ray powder diffraction (XRD Bruker AXS D8), SEM energy dispersive (SEM – LEO 440) X-ray (EDX) mapping, and Fourier-transform infrared spectroscopy (FTIR Perkin Elmer 400). Instrumental analysis was performed by the Nanotechnology Research Center (ERNAM) and Technology Research Center (TAUM), Erciyes University. Transmission electron microscopy (TEM) images were recorded using the JEOL 1220 JEM TEM of the Eskişehir Osmangazi University Research Laboratory Application and Research Center (ARUM). The size and dispersity of nanoparticles were analyzed using a dynamic light scattering (DLS) instrument (Horiba nanoPartica SZ-100V2), NanoBiotech Research Laboratory.

The average crystallite size of gIONPs was determined by Debye–Scherrer equation \( d = \frac{k\lambda}{\beta \cos \theta} \), where \( k \) is the Debye–Scherrer constant (0.94 for FWHM of spherical crystals with cubic symmetry), \( \lambda \) is the X-ray wavelength (0.154 nm), \( \beta \) is the width of the peak with the maximum intensity in half height, \( d \) is the thickness of the crystal, and \( \theta \) is the diffraction angle (radian). Origin 2021b (© 1991-2021 OriginLab Corporation) was used for the calculation of the FWHM of XRD peaks for the calculation of \( \beta \).

Design of experiment and analysis

In this study, the design of experiment (DOE) strategy and the Taguchi method for the experimental design approach was used to find optimum experimental conditions of the gIONPs with the minimum
polydispersity index value (PDI). The goal was the investigation of the six parameters of synthesis including temperature (Temp), pH of carob pod extract, extract concentration of carob pod, the concentration of iron ion, stirring rate (RPM), and reaction time (time). Six experimental parameters used for the green synthesis of iron oxide nanoparticles and their corresponding three levels are represented in Table 1. The analysis of variance (ANOVA) was used to determine the significance of individual parameters for the control of the final response.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Levels</th>
</tr>
</thead>
<tbody>
<tr>
<td>pH of carob pod extract</td>
<td>Low 5</td>
</tr>
<tr>
<td>Temperature (°C)</td>
<td>Center 8</td>
</tr>
<tr>
<td>Carob pod extract concentration (% mg/mg)</td>
<td>High 11</td>
</tr>
<tr>
<td>Concentration of iron ion (M)</td>
<td>Low 0.07</td>
</tr>
<tr>
<td>Stirring rate (rpm)</td>
<td>Center 0.14</td>
</tr>
<tr>
<td>Reaction Time (h)</td>
<td>High 0.28</td>
</tr>
</tbody>
</table>

The S/N ratio and the analysis of variance (ANOVA) with a significance level of 0.05 were employed to study the contributions of the parameters to the response. The Minitab software version 13 (Ankara, Turkey) as a tool for statistical design of experiment was used. The equivalent volume of the gIONPs suspension (2 mL) was diluted with a constant volume (2 mL) of deionized water and then subsequently all measurements were performed at room temperature at the DLS instrument. The experiments of 27 runs with 3 trials were conducted for three levels of six parameters.

**Stability analysis of gIONPs**

The zeta potential measurement of the colloidal gIONPs samples was determined using a zeta potential analyzer (Horiba nanoPartica SZ-100V2). The method used by the SZ-100V2 is known as laser Doppler electrophoresis. The sample was irradiated with a laser light of 532 nm at 3.3 electrode voltage. A cell with an electrode was used to introduce an electric field into the sample. All presented zeta potentials were reported as the mean value of three independent measurements at 25°C temperature. UV–Visible spectroscopy (Hach DR 6000 UV-Vis spectrophotometer) was used for monitoring the maximum absorbance of nanoparticles over time. The UV-Vis spectrum of the gIONPs sample at a dilution ratio of 1:10(v/v) was monitored using a 1 cm glass cuvette. The absorbance spectrum was obtained between 200 – 700 nm. The stability analysis was performed for 3 months.

**Antibacterial activity analysis of gIONPs by well-diffusion method**
The bacterial culture and antibiotic discs were obtained from the Veterinary Medicine Faculty Laboratory of Erciyes University. The bacterial activity of glONPs was performed against both Gram-negative (Escherichia coli 0157:H7) and Gram-positive (Staphylococcus aureus ATCC 25923). The antibacterial activity analysis was performed due to the Kirby-Bauer well diffusion method. Briefly, the selected bacterial pathogenic strains were subcultured on Mueller–Hinton nutrient broth. Each strain was spread uniformly on nutrient agar plates using a sterile cotton swab. A sterile cork borer was used to create a well of about 8 mm diameter for testing the antibacterial activity of glONPs. 250 mg of glONPs was dispersed in 1 mL of water. 100 µL of glONPs was loaded into the well (8 mm diameter) on the Muller–Hinton agar plate. Antibiotics of Cefoxitin (30 µg /disc), Cefotaxime (30 µg /disc), Gentamicin (10 µg /disc), and Tetracycline (30 µg / disc) were used as a positive control. The extract of Ceratonia siliqua L. (100 µL) was used as a negative control. Then the plates were incubated at 37 °C for 24 h. The different zone levels of inhibition were measured and analyzed according to the Kirby Bauer Interpretation chart (Table S3). The average values of triplicate measurements were calculated as an outcome of antibacterial activity.

3. Results And Discussion

Optimization of glONPs synthesis parameters

Taguchi experimental design was employed based on the L27 orthogonal array for three levels of six parameters. The goal was the investigation of the six parameters of synthesis including the concentration of carob pod extract, the concentration of iron ion, temperature, pH of carob pod extract, reaction time, and stirring rate. Supplementary Table S1 shows the 27 experimental runs with their responses of hydrodynamic size and polydispersity (PDI) value according to the DLS measurements. The term “polydispersity “is used to describe the degree of non-uniformity of size distribution. The numerical value of PDI ranges from 0.0 (perfectly uniform sample) to 1.0 (multiple particle size populations) and is measured by the dynamic light scattering (DLS) technique [15].

The S/N ratio results of each parameter effect on the PDI of glONPs are presented in Fig.1 as graphs. The highest S/N ratios were selected for the smallest PDI value response. The response table for the S/N ratios of parameters is shown in Table 2. According to their larger value of delta, the parameters were assigned as rank 1, 2, 3, 4, 5, and 6. Rank 1 means the highest contribution parameter for the lowest PDI value while rank 6 means the lowest contribution parameter.

The reaction time is the highest contribution parameter while the pH of the carob pod extract is the least contribution parameter. The optimum parameter levels for the smallest PDI value were determined as 0.14 M iron ion concentration, 10% extract concentration, extract pH of 5, 25 °C temperature, stirring rate of 400 rpm, and reaction time of 3 hours (Fig.1). Plant extract is the dominant parameter in nanoparticle synthesis, as it is the source of reducing agents and stabilizing molecules. Increasing the plant concentration from 10% to 20% (w / w) increased the PDI of the nanoparticles, while also causing agglomeration at 30%. Meantime, enhancement of the reaction time from 1 to 3 hours lead to...
nanoparticles with the smallest size. The stirring rate and pH of carob pod extract did not make a huge difference to the response. The direct effect of temperature may be due to enhancing the rate of nanoparticle formation [16]. The pH change at the carob pod extract might not have altered the electrical charges of biomolecules and capping agents and their ability to bind and reduce metal ions [17]. The stirring rate of 400 rpm was observed as a transition condition between the stirring rate of 200 and 600 rpm for the smallest size and polydispersity index. The low stirring rate was proved unable to produce iron oxide nanoparticles. The high mixing rate might have caused the nanoparticles to have high surface energy and caused their agglomeration [18]. Analysis of variance (ANOVA) showed that the significance level of the studied parameters in Table 3. The results indicate that iron ion concentration, reaction time, temperature, and carob pod extract concentration had a significant effect on PDI value (p-value < 0.05).

There are several studies about the procedure optimization of the green synthesis of several nanoparticles such as silver, zinc, and gold, etc. in the literature. The green synthesis of nanoparticles is related to various factors, including types of plant biomolecules and concentration, pH, temperature, stirring rate, and exposure time. These factors are found to control the sizes and morphologies of the nanoparticles according to the literature [19] [20] [21] [22]. The effects of various parameters in the green synthesis method of iron nanoparticles such as iron precursor, plant extract concentration, reaction temperature, and pH have been studied little when the studies in the literature are reviewed. Kheshtzar et.al. reported that the quantity of leaf extract and concentration of iron precursor is the most effective parameters in the green synthesis reaction [23]. It was revealed by Gholami et.al. that the iron precursor to extract ratio has a significant effect on green iron nanoparticles synthesis [24].

Table 2. Response table for S/N ratios of PDI

<table>
<thead>
<tr>
<th>Level</th>
<th>Extract Concentration</th>
<th>Iron Ion</th>
<th>Time</th>
<th>Temp</th>
<th>RPM</th>
<th>pH</th>
</tr>
</thead>
<tbody>
<tr>
<td>Delta</td>
<td>6.0092</td>
<td>7.1950</td>
<td>8.4651</td>
<td>8.0705</td>
<td>1.6846</td>
<td>1.4451</td>
</tr>
<tr>
<td>Rank</td>
<td>4</td>
<td>3</td>
<td>1</td>
<td>2</td>
<td>5</td>
<td>6</td>
</tr>
</tbody>
</table>

Table 3. The results of ANOVA for the designed orthogonal array design in the optimization of gIONPs synthesis parameters.
Analysis of Variance (ANOVA)

<table>
<thead>
<tr>
<th>Parameters</th>
<th>DF</th>
<th>Adj SS</th>
<th>Adj MS</th>
<th>F-Value</th>
<th>P-Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Extract Concentration (w/w %)</td>
<td>2</td>
<td>5.310</td>
<td>5.310</td>
<td>5.15</td>
<td>0.021</td>
</tr>
<tr>
<td>Iron Ion (M)</td>
<td>2</td>
<td>28.023</td>
<td>28.023</td>
<td>27.18</td>
<td>0.000</td>
</tr>
<tr>
<td>Time (h)</td>
<td>2</td>
<td>37.887</td>
<td>37.887</td>
<td>36.75</td>
<td>0.000</td>
</tr>
<tr>
<td>Temp (°C)</td>
<td>2</td>
<td>46.776</td>
<td>46.776</td>
<td>45.37</td>
<td>0.000</td>
</tr>
<tr>
<td>RPM</td>
<td>2</td>
<td>1.921</td>
<td>1.921</td>
<td>1.86</td>
<td>0.192</td>
</tr>
<tr>
<td>pH of extract</td>
<td>2</td>
<td>2.410</td>
<td>2.410</td>
<td>2.34</td>
<td>0.133</td>
</tr>
<tr>
<td>Error</td>
<td>14</td>
<td>7.216</td>
<td>7.216</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Total</td>
<td>26</td>
<td>129.543</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Characterization of gIONPs at optimum synthesis parameters

A follow-up characterization analysis was carried out considering the optimum values of synthesis parameters for gIONPs. The hydrodynamic size and PDI value of gIONPs at the optimum parameters were measured as 78 ±22 nm and 0.42 ±0.06, respectively (Fig.2(a)). gIONPs sample was measured at an average zeta potential of + 40.75 ± 1.22 mV (Fig.2(b)). The stability of nanoparticle solutions can be obviously predicted by zeta potential measurements. The particles tend to agglomerate as the electric potential approaches zero. Particles tend to repel other particles at the zeta potential of either more than +20 mV or less than -20 mV [25].

The formation of gIONPs in the aqueous colloidal solution was confirmed by the peak at 299 nm using UV–visible spectral analysis (Fig.3). As per the previously reported literature, this is due to the green biosynthesis of iron oxide nanoparticles [26]. The absorbance peak of iron oxide nanoparticles synthesized using plant extract ranged from 240 to 360 nm [27] [28]. The X-ray diffraction of gIONPs was recorded at 2θ values ranging from 10°–80° (Fig. 4). XRD spectrum showed distinct peaks at 24.20°, 33.18°, 35.68°, 40.90°, 49.54°, 54.17°, 62.47°, and 64.12°, which indicates the crystalline nature of the iron oxide nanoparticles and corresponds to the hematite (JCPDS No. 013-0534) (Table S2) [29] [30]. The average crystallite size of the nanoparticles was calculated as 5.09 nm using Debye-Scherrer’s equation.

Fig.5(b) and Fig.5(c) present the images of iron oxide nanoparticles captured by transmission electron microscopy (TEM). The TEM images showed that the nanoparticles are in a spherical shape and some of the particles were agglomerated. In the absence of coating materials, iron oxide nanoparticles that form part of a colloid tend to agglomerate. These agglomerates may be caused by either strong or weak physical interactions [33].

The gIONPs at optimum conditions were also characterized by energy-dispersive X-ray spectroscopy (EDX) to examine their composition. EDX was built in the microscope by which the SEM image of the
sample was recorded. The EDX spectrum is presented in Fig.6. The atomic percentages are 33.14% of oxygen and 64.26% of iron. The EDX confirmed the presence of both iron and oxygen which indicates the reduction of iron ions to iron oxide by carob pod extract. The Cl signal was detected originating from iron chloride precursor salt used in the synthesis protocol and from the plant biomaterial used for the synthesis of nanoparticles. The signals for K were observed which may originate from the plant biomaterial [34].

FTIR spectroscopy for the extract and gIONPs solution was carried out. It was observed that the soluble elements present in plant extract acted as capping agents of gIONPs due to the FTIR analysis. The spectra of the carob pod extract revealed absorption bands at 3493, 3120, 1708, 1377, 1040, 803, and 626 cm\(^{-1}\), while the absorption bands of synthesized gIONPs were observed at 3028, 2080, 1680, 1390, 1047, 917, and 470 cm\(^{-1}\) (Fig.7).

The band at 3493 cm\(^{-1}\) is an indicator of stretching in the OH group within free hydroxyl [35]. The band located at 3120 cm\(^{-1}\) is assigned to stretching of O-H group bond in galactose sugar compound in carob pod extract [36]. The band in 1708 cm\(^{-1}\) shows the presence of unsaturated carbonyl groups [37]. The band in 1377 cm\(^{-1}\) is attributed to the symmetrical stretch of the methyl functional group [38]. The band at 1040 cm\(^{-1}\) can be assigned to the stretching vibration of C-N function groups in plant polyphenolic species [39]. The band in 626 cm\(^{-1}\) shows the C-S stretching [40]. The band in 3028 cm\(^{-1}\) is attributed to the C–H and C–O bending and stretching of oleic acids [41]. The band in 2080 cm\(^{-1}\) is also a typical band of stretching bands of unbound CO [42]. The absorption bands at 1047, 1680, and 1390 cm\(^{-1}\) were assigned to the strong C=O stretching band related to the carbonyl group and medium C-H bending, respectively [43]. The band at the 917 cm\(^{-1}\) is attributed to the C-H bending due to the carbohydrate. The formation of gIONPs is characterized by the absorption bands from 428 to 686 cm\(^{-1}\). The band at 470 cm\(^{-1}\) corresponds to the Fe-O vibration bonds related to the octahedral units of the magnetite phase [44] [45]. This band was absent in plant extract which indicates the formation of iron oxide nanoparticles. A similar observation has been reported by several studies [46] [47].

**Stability analysis of colloidal gIONPs at optimum synthesis parameters**

The pH and zeta potential of colloidal gIONPs solution was measured as 1.9 and +42.5 mV after synthesis, respectively. The schematic representation of zeta potential is seen in Fig.8. The pH of the isoelectric point of the gIONPs was determined as 6.8 by the zeta potential measurements (Fig.9(a)). The gIONPs began to agglomerate at pH values greater than 5 and less than 11 (Fig.9(b)). The stability of the colloidal gIONPs solution was evaluated by monitoring the zeta potentials over 3 months (Table 4). The high stability of gIONPs is defined by their high electrostatic potential. A similar study has been performed for the iron oxide nanoparticles synthesized using the aqueous coprecipitation method and it was observed that the zeta potential of nanoparticles was constant at around −39 mV for long-term storage (over 16 months) [48].
Table 4. Zeta potential measurement of gIONPs nanoparticles over 3 months

<table>
<thead>
<tr>
<th>Time (month)</th>
<th>Zeta Potential (mV)</th>
<th>pH</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>+ 42.5</td>
<td>1.92</td>
</tr>
<tr>
<td>1</td>
<td>+ 41.0</td>
<td>1.91</td>
</tr>
<tr>
<td>2</td>
<td>+ 40.4</td>
<td>1.95</td>
</tr>
<tr>
<td>3</td>
<td>+ 40.8</td>
<td>1.90</td>
</tr>
</tbody>
</table>

The UV−vis absorbance spectrum of gIONPs colloidal solution over 3 months was monitored and the maximum absorbance did not change during this period (Fig.10). Metal nanoparticles exhibit surface plasmon resonance (SPR) absorption dependent on the size, shape, and composition [49]. According to this phenomenon, it could be explained that gIONPs were formed with good stability and the composition did not change due to the formation of agglomerates during the 3 months period [50]. Shoaib et al. have carried out similar stability experiments with green silver nanoparticles using UV−vis spectroscopy and they indicated the change of the maximum absorbance due to the presence of agglomerations [51].

**Antibacterial activity evaluation of gIONPs**

Agar well diffusion method was employed to evaluate the antibacterial activity of powder form of gIONPs against the Gram-negative (*Escherichia coli*) and Gram-positive (*Staphylococcus aureus*) bacteria species. gIONPs showed clear zones of inhibition of 24 mm and 21 mm in 250 mg/mL concentration against *S. aureus* and *E. coli* bacteria respectively. The negative control of *Ceratonia siliqua* L. extract had no effect on microbial growth whereas the standard antibiotics selected as positive control showed inhibition on the selected microorganisms. The inhibition activity of the gIONPs were compared with standard antibiotics Cefoxitin, Cefotaxime, Gentamicin, and Tetracycline shown in Table 5. The results were compared with the standard Kirby Bauer interpretation chart (Table S3). The Gram-positive *Staphylococcus aureus* bacteria was found to be more susceptible to gIONPs than the Gram-positive pathogens *Escherichia coli*. gIONPs showed the highest inhibition zone against *S. aureus* when compared with 4 standard antibiotics. The antibacterial property of gIONPs showed a greater inhibition activity against Gram positive than Gram negative bacteria (Fig.11). This might be because of the differences in cell wall composition of bacteria [52].

Table 5. Zone inhibition of gIONPs, *Ceratonia siliqua* L. extract and standard antibiotics against selective bacterial strains.
### Zone of Inhibition (Diameter in mm)

<table>
<thead>
<tr>
<th></th>
<th>S. aureus</th>
<th>E. coli</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ceratonia siliqua L. Extract</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Cefoxitin (FOX) (30 µg/disc)</td>
<td>11.87 ± 0.14</td>
<td>28.46 ± 0.11</td>
</tr>
<tr>
<td>Cefotaxime (CTX) (30 µg/disc)</td>
<td>-</td>
<td>14.27 ± 0.09</td>
</tr>
<tr>
<td>Gentamicin (CN) (10 µg/disc)</td>
<td>23.62 ± 0.16</td>
<td>22.83 ± 0.08</td>
</tr>
<tr>
<td>Tetracycline (TE) (30 µg/disc)</td>
<td>10.5 ± 0.15</td>
<td>23.92 ± 0.14</td>
</tr>
<tr>
<td>gIONPs (250 mg/mL)</td>
<td>24.27 ± 0.12</td>
<td>20.83 ± 0.11</td>
</tr>
</tbody>
</table>

The inhibitory action of gIONPs may be attributed to the oxidative stress by reactive oxygen species generation and the Fenton reaction [53]. When iron nanoparticles penetrate cells through disrupted membranes, it may cause further damage. Li et al. found that the iron oxide nanoparticles not only inhibited *E. coli* cell growth, but also caused bacterial cell death due to the production of significantly elevated reactive oxygen species (ROS) such as hydroxyl radicals, superoxide radicals, singlet oxygen, and hydrogen peroxide levels [54]. Das et al. performed the ROS measurement studies of *S. aureus* in presence of iron oxide nanoparticles. They found that iron oxide nanoparticles enhance the fluorescence intensity significantly which is directly correlated with the quantity of ROS developed at *S. aureus* bacterial cells. [55]. Similar results of the antibacterial effect of iron nanoparticles on *S. aureus* and *E. coli* have been reported in recent studies. Kanagasubbulakshmi and Kadirvelu evaluated the antibacterial property of iron oxide nanoparticles against Gram negative – *E. coli* and Gram positive- *S. aureus*. They found the zone of inhibition of 10 mm and 8 mm for *Escherichia coli* and *Staphylococcus aureus*, respectively [56]. Suganya et al. reported considerable antibacterial activity of iron oxide nanoparticles from leaf extract of *Passiflora foetida* towards Gram negative and positive bacteria. They found the zone of inhibition for *Staphylococcus aureus* and *Escherichia coli* as 11 mm and 18 mm, respectively [57]. Amutha and Sridhar used the *Glycosmis mauritiana* leaf extract for the formation of iron oxide nanoparticles and evaluated its application on antibacterial activity against the tested pathogens of *E. coli* and *S. aureus*. They found the minimum zone of inhibition as 12 mm and 8 mm for the 10 µg/disc of green synthesized iron oxide nanoparticles [58].

### 4. Conclusion

The potential of *Ceratonia siliqua* L. aqueous extract for the simple and one-pot green synthesis of iron oxide nanoparticles was investigated. This study focused on the optimization of the green synthesis parameters for the stable colloidal form of gIONPs and evaluation of the antibacterial effect of the gIONPs powder against Gram-negative (*Escherichia coli*) and Gram-positive (*Staphylococcus aureus*) bacteria. Optimum synthesis parameters were found to be efficient in terms of synthesis of narrow-sized as well as stable nanoparticles. Reaction time and temperature were the most effective parameters in the iron oxide nanoparticle synthesis reaction. The pH of extract and stirring rate showed no significant effects. The biomaterials from the Ceratonia siliqua L. extract successfully achieved the monodispersed
and stable gIONPs for 3 months period. The particles were effectively surrounded by biomaterials without any external addition of surfactant and polymers. Powder form of gIONPs also showed antibacterial activity against both Gram-negative (*Escherichia coli*) and Gram-positive (*Staphylococcus aureus*). The gIONPs inhibited the growth of *Staphylococcus aureus* with high susceptibility when compared with the standard antibiotics of Cefotaxime, Tetracycline, Gentamicin, and Cefoxitin and also the growth of *Escherichia coli* when compared with the standard antibiotics of Tetracycline and Gentamicin. gIONPs could serve as a potential antibacterial agent.

**Declarations**

**Author Contributions**

Derya Aksu Demirezen: Methodology, Data curation, Investigation, Writing - original draft. Şeyda Yılmaz: Methodology, Data curation. Dilek Demirezen Yılmaz: Methodology, Resources. Yalçın Şevki Yıldız: Resources, Funding.

**Conflict of Interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work.

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**References**


Figures

Figure 1
Main effects plot of S/N ratio for smallest PDI value.

Figure 2
(a) The particle size distribution and (b) zeta potential of colloidal gIONPs.

Figure 3
UV/Vis absorption spectrum of colloidal gIONPs.
Figure 4

XRD pattern of glONPs.

Figure 5

(a) SEM image and TEM images of (b) distribution and (c) size and shape of glONPs.

Figure 6

EDX analysis of glONPs.

Figure 7

FTIR spectra of colloidal glONPs and carob pod extract.

Figure 8
Schematic representation of zeta potential of colloidal gIONPs.

Figure 9

(a) Zeta potential versus pH plot for colloidal gIONPs and (b) images of gIONPs colloidal solution at different pH values.

Figure 10

UV-Vis absorption spectra of colloidal gIONPs over 3 months.

Figure 11

(a) Antibacterial activity of gIONPs, Ceratonia siliqua L. Extract, and standard broad-spectrum antibiotics (Cefoxitin (FOX), Cefotaxime (CTX), Gentamicin (CN), and Tetracycline (TE)) against S. aureus and E. coli bacteria (b)-(c) The plates showing the inhibition zones of gIONPs and Ceratonia siliqua L. Extract, and standard broad-spectrum antibiotics (Cefoxitin (FOX), Cefotaxime (CTX), Gentamicin (CN), and Tetracycline (TE)) against S. aureus and E. coli bacteria.

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