

# Identification and validation of RNA binding protein-associated prognostic model for Neuroblastoma

**Jun Yang**

University of Hong Kong - Shenzhen Hospital

**Jiaying Zhou**

University of Hong Kong - Shenzhen Hospital

**Cuili Li**

University of Hong Kong - Shenzhen Hospital

**Shaohua Wang** (✉ [lwwsbs22@163.com](mailto:lwwsbs22@163.com))

Department of Pediatrics, Women and Children Institute of Futian, University of South China, ShenZhen, China

---

## Research Article

**Keywords:** Neuroblastoma, RNA binding proteins, prognostic, TARGET, GTEx

**Posted Date:** January 7th, 2021

**DOI:** <https://doi.org/10.21203/rs.3.rs-117557/v1>

**License:** © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License. [Read Full License](#)

---

# Abstract

**ABSTRACT** Background: The abnormal expression of RNA binding protein (RBP) may be related to the development and progress of cancer. However, little is known about the mechanism of RBP in neuroblastoma (NB). Methods: We downloaded the RNA expression data of NB and normal nervous tissues from the (TARGET) database and GTEx database, and determined the differential expression of RBP between normal and cancerous tissues. Then the function and prognostic value of these RBPs were systematically studied. Results: A total of 348 differentially expressed RBPs were identified, together with 166 up-regulated RBPs and 182 down-regulated RBPs. Two hub RBPs (CPEB3 and CTU1) were identified as prognostic-related genes and chose to build prognostic risk score models. Further analysis showed that based on this model, the overall survival rate of patients in the high-risk subgroup was lower ( $P=2.152e-04$ ). The area under the curve(AUC) of the receiver-operator characteristic curve(ROC) of the prognostic model is 0.720 in the TARGET cohort. There is a significant difference in the survival rate of patients in the high and low risk subgroups in the validation data set GSE85047 ( $P = 0.1237e-08$ ), the AUC is 0.730. Conclusions: RNA binding protein (CPEB3 and CTU1) can be used as molecular markers of NB. Keywords: Neuroblastoma, RNA binding proteins, prognostic, TARGET, GTEx

## Introduction

Neuroblastoma is still the main cause of tumor-related deaths in children worldwide [1]. As diagnosis and treatment methods have made great progress in the past twenty years, the average 5-year relative survival rate of neuroblastoma has reached 50%[2]. Currently, the diagnosis of NB mainly relies on histopathological examination, imaging results and tumor molecular biomarkers.[3] It is difficult to detect neuroblastoma early. This may be the most important reason affecting the mortality of patients with neuroblastoma[4]. Therefore, further study of the molecular mechanism of neuroblastoma and identification of effective molecular markers for early cancer screening are essential to enhance the therapeutic outcomes and quality of life of child.[5].

RNA binding protein (RBP) is a pleiotropic protein that can regulate gene expression to reach post-transcriptional levels by interacting with target RNA[6]. It can interact with various types of RNA, such as rRNA, ncRNA, snRNA, miRNA, mRNA, tRNA and snoRNA [7]. Now, over 1,500 different types of RBP have been found in the human genome through whole-genome screening, [8]. RBP regulates cell functions by interacting with RNA, and plays an important function in post-transcriptional gene expression regulation.[9] The ribonucleoprotein complex formed by the binding of RBP and target RNA regulates the stability of mRNA on post-transcriptional level, thereby affecting RNA processing, splicing, localization, export and translation.[10, 11] Regulate and determine various important physiological processes of cells [12]. Existing research have found that RBPs play an important responsibility in many human diseases which are key regulators of the development and progression of cardiovascular diseases[13], myotonic muscular dystrophy[14], neurological diseases[15] and cancer[16].

Therefore, we use high-throughput bioinformatics analysis to identify RBPs that are differentially expressed between cancerous samples and normal samples, and systematically investigated their expression patterns, functional effects and potential mechanisms to understand their role in tumors. This study will deepen our

understanding of the molecular mechanism of NB and provide potential diagnostic or prognostic biomarkers for NB.

## Materials And Methods

### Data sets and preprocessing

We get RNA expression datasets and corresponding clinical datasets of NB patients from the Therapeutically Applicable Research To Generate Effective Treatments project database (TARGET, <https://ocg.cancer.gov/programs/target>), and normal neural tissues samples datasets from Genotype-Tissue Expression Database (GTEx, <https://gtexportal.org/>), respectively. All data comes from an open public big data platform, this study does not require ethics approval. In order to determine the differentially expressed genes between NB tissue and normal sample, the Limma software package was used for analysis. The GSE85047 dataset were downloaded from Gene Expression Omnibus (GEO) (<https://www.ncbi.nlm.nih.gov/geo/>) and used as a validation cohort.

### Gene Ontology (GO) enrichment and KEGG pathway analysis

Gene enrichment analysis and pathway analysis was carried out by the R package “clusterProfiler”. [17]

### Protein- protein interaction (PPI) network building and subnet detection.

Significant differential protein-protein interaction information of RBP is evaluated using STRING database (<http://www.string-db.org/>) [18] and further building and visualization of the PPI network by Cytoscape 3.7.0 software. The application the molecular complexity detection (MCODE) plug-in clusters genes in the PPI network and constructs functional modules.  $P < 0.05$  was considered a statistically significant difference [19].

### Prognostic model construction

The R package “survival” was applied to carry out univariate Cox regression analysis on all differential RBPs to identify the prognostic genes, and lasso regression was performed to further screen important key genes. Finally, based on the preliminary screening of the above key candidate genes, we built a multivariate Cox proportional hazard regression model, and evaluated the survival of patients through risk scores. The sample risk score formula is like this:

$$\text{Risk score} = \beta_1 * \text{Exp1} + \beta_2 * \text{Exp2} + \dots + \beta_i * \text{Exp}_i$$

Among them,  $\beta$  was the value of risk coefficient, and Exp represented the value of expression in a certain gene. In accordance with the median value of risk score, NB patients were divided into two groups: low-risk group and high-risk group, and the survival difference between the two subgroups was compared through survival analysis. In addition, the prognostic ability of the above model is estimated through receiver operating characteristic curve (ROC) analysis. A sample of 276 NB patients with dependable follow up information from the GSE85047 data set was used as a validation group to evaluate the predictive power of the prognostic model.  $P < 0.05$  was considered a statistically significant difference.

# Results

## Identification of differently expressed RBPs in NB patients

In this research, we performed a methodical analysis of the key role and prognostic value of RBP in NB. The NB data is downloaded from TARGET, which contains 144 tumor samples, and the normal nerve tissue data is downloaded from the GTEx database, which contains 278 samples. After analyzing the currently known 1542 RBPs, 348 RBPs with significant differences ( $P$  adjusted  $<0.05$ ,  $|\log_2FC|>1.0$ ) were screened out, together with 166 up-regulated RBPs and 182 down-regulated RBPs. (Figure1)(Table S1)

## Enrichment analysis of the differently expressed RBPs

In order to study the functions and mechanisms of the selected RBP, we use the R package clusterProfiler for enrichment analysis. The results show that biological processes are mainly enriched in mRNA processing, RNA splicing, ncRNA metabolic process, RNA phosphodiester bond hydrolysis, RNA splicing, via transesterification reactions with bulged adenosine as nucleophile, mRNA splicing, via spliceosome, RNA splicing, via transesterification reactions, nucleic acid phosphodiester bond hydrolysis, RNA catabolic process, and MF in catalytic activity, acting on RNA ribonuclease activity, nuclease activity, mRNA 3'-UTR binding, endonuclease activity, translation regulator activity, catalytic activity, acting on a tRNA, mRNA binding, double-stranded RNA binding, endoribonuclease activity, single-stranded RNA binding, and CC in ribonucleoprotein granule, cytoplasmic ribonucleoprotein granule, ribosome, ribosomal subunit, organellar ribosome, mitochondrial ribosome, P-body, mitochondrial matrix, P granule, pole plasm. The KEGG chiefly enriched in RNA transport, mRNA surveillance pathway, Ribosome biogenesis in eukaryotes, RNA degradation, Ribosome, Aminoacyl-tRNA biosynthesis, Spliceosome, RNA polymerase, Influenza A. (Figure 2)(Table1,2)

## PPI network building and subnet detection

In order to more study the function of differential RBP and its role in the development of NB, we used Cytoscape software to create a PPI network, which contains 311 nodes and 1766 edges. The co-expression network was analysis with the MCODE to recognize potential key section. (Figure 3) The RBPs in the subnet 1 were mainly enriched in ribosome biogenesis in eukaryotes pathway, ribosome biogenesis, rRNA processing, ncRNA processing, ,maturation of SSU-rRNA, ribosomal small subunit biogenesis, rRNA metabolic process ,maturation of SSU-rRNA from tricistronic rRNA transcript (SSU-rRNA, 5.8S rRNA, LSU-rRNA), ribosomal large subunit biogenesis.

## Prognosis-related RBPs selecting

The difference analysis identified a total of 348 key RBPs. In order to learn the prognostic significance of these RBPs and their effect on clinical outcome and survival time, we conducted univariate Cox regression analysis and get 4 candidate center RBPs related to prognosis(Figure 4 ). Subsequently, through lasso regression, the prognostic risk equation of multi-factor Cox regression was established. (Figure5, Table3).

## Prognosis-related RBPs model building and analysis

At last, CPEB3 and CTU1 were identified as the key prognostic genes by the multivariate Cox regression analysis. We used this two hub genes to construct the predictive model. The risk score of every child was calculated in accordance with the following formula:

$$\text{Risk score} = (-0.60901 * \exp \text{CPEB3}) + (0.851637 * \exp \text{CTU1}).$$

Then, based on median value of riskscore, 144 NB patients were divided into two groups: low-risk group and high-risk group. The results showed that compared with patients in the low-risk group, patients in the high-risk group had poorer survival, which was statistically significant ( $P=2.152e-04$ ). The value of area under curve(AUC) in the TARGET model is 0.720. (Figure6A, 6B, Figure 7A)

### Validation of hub RBPs

With the purpose of evaluation of the prognostic value of the RBPs prediction model, we used the GSE85047 patient cohort to verify the relationship between risk score and survival time. In the GSE85047 cohort, groups were also grouped based on the median value of risk score in the TARGET model. The survival time of patients with high risk scores was poorer for patients with lower risk scores, which was significant ( $P = 0.1237e-08$ ), and the AUC was 0.730. (Figure6C,6D, Figure 7B)

### RBPs as an independent prognostic factor

We assessed the prognostic value of risk scores of RBPs. For NBL-TARGET, the risk score in univariate analysis was significantly correlated with overall survival (OS) ( $\text{HR}=1.535$ ,  $95\% \text{ CI}=1.368-1.722$ ,  $P=2.69E-13$ ) (Figure 8A). Multivariate analysis showed that the risk score was an independent prognostic indicator ( $\text{HR}=1.518$ ,  $95\% \text{ CI}=1.344-1.715$ ,  $P=1.91E-11$ ) (Figure 8B). Nomogram integrates multiple risk factors to quantify individual risks in the clinical environment. We conducted a nomogram to predict the probability of OS in 1, 2, and 3 years. Figure 8C

## Discussion

The prognosis of different neuroblastoma patients varies greatly, that is, there is extensive tumor heterogeneity among neuroblastoma. For low-risk neuroblastoma patients (most commonly in infants), simple observation or surgical treatment can often achieve good results; but for high-risk neuroblastoma patients, even if a variety of intensive treatment options are combined.[20] The prognosis is still not ideal. The true cause of neuroblastoma is still unclear. In recent years, with the emergence of immunotherapy and new drugs, the survival of patients in the high-risk group has improved to a certain extent[21].

RBP has always been with RNA life. It is not an exaggeration to say: Without RBP, RNA can't do anything. Its main role is to mediate RNA maturation, transport, localization and translation; one RBP may have multiple target RNAs; and its expression defects can cause multiple diseases. Recently, the importance of RBP in tumor occurrence, development, and metastasis has gradually been noticed[22].

In our research, we identified 348 RBPs based on NB datasets from TARGET. We systematically analyzed the related biological functions and built these RBP PPI networks and its subnets. In addition, we also performed

univariate Cox regression analysis, survival analysis, lasso regression analysis and multivariate Cox regression analysis of differential RBP to more investigate its biological function and prognostic value.

The GO enrichment and KEGG pathway analysis of these differentially expressed RBPs indicate that RBP is used in mRNA monitoring pathways, RNA transport, ribosomal biogenesis in eukaryotes, RNA degradation, ribosomes, aminoacyl-tRNA biosynthesis, and spliceosome. It is significantly enriched in RNA polymerase pathway, and plays an critical function in mRNA processing, RNA splicing, ncRNA metabolism, RNA phosphodiester bond hydrolysis, catalytic activity, and acting on RNA ribonuclease activity and nuclease activity. At present, many research have reported its role in various forms of RBPs in metabolism and disease. It plays a dual and opposite role in tumorigenesis, regulating the proliferation of early tumor cells and promoting tumor progression and metastasis of advanced cancer. According to reports, abnormal expression of multiple RBPs had been found in many malignant tumors[10, 23, 24]. However, the impact of RBP on the occurrence and development of cancer is still poorly understood.

A total of 2 RBPs were identified as hub RBPs related to NB prognosis, including CPEB3 and CTU1.

CPEB3 (Cytoplasmic Polyadenylation Element Binding Protein) is an RBP that shuttles through the cytoplasm. It mainly exists in the cytoplasm and can inhibit the translation of target RNA. CPEB3 can disrupt the crosstalk between cancer cells and tumor-associated macrophages through IL-6R/STAT3 signaling, thereby inhibiting epithelial-mesenchymal transition. Studies have found that CPEB3 is related to the tumorigenesis and development of glioma[25], high-grade serous ovarian cancer[26], colorectal cancer[27], hepatocellular carcinoma[28], cervical cancer[29].

CTU1 (cytoplasmic sulfurylase subunit 1) is a protein-coding gene. The diseases associated with CTU1 include spinal cord septal membrane tumors and spinal gliomas. The related pathways include gene expression and tRNA processing, and its function is related to tRNA binding and nucleotide transferase activity. CTU1 promotes cancer resistance to targeted therapy[30], related to Spinal Cord Gliomas' high rates of morbidity and mortality[31], and up-regulation of CTU1 is involved in human breast cancer metastasis.

Based on the two hub RBPs trained by the TARGET cohort, multi-step Cox regression analysis produced a riskscore model that can predict the prognosis of NB. In the TARGET-NB cohort and GSE85047 cohort, the survival results of the high and low risk subgroups were significantly different, and the ROC values of the training set and validation set were 0.72 and 0.73, respectively, indicating that the 2-gene marker prognostic model is used to evaluate the prognosis of NB patients has a certain value. However, the molecular mechanisms of these two RBPs are still little known, and further study of their underlying function may be valuable.

In summary, we systematically study the function and prognostic value of RBPs differently expressed in NB. These RBPs may be related to the occurrence, development, invasion and metastasis of NB. The establishment of a prognostic model of NB gene based on two RBP coding genes is conducive to clinical application. Our results help explain the pathogenesis of NB and develop new molecular markers of therapeutic and prognostic targets.

## Declarations

## Availability of data and materials

The datasets used to support the findings of this study are available in Therapeutically Applicable Research To Generate Effective Treatments project database (TARGET, <https://ocg.cancer.gov/programs/target>) and Gene Expression Omnibus (GEO) (<https://www.ncbi.nlm.nih.gov/geo/>).

## Ethics approval and consent to participate

This study was approved by the Academic Committee of HKU-SZH, and conducted according to the principles expressed in the Declaration of Helsinki. All the datasets were retrieved from the publishing literature, so it was confirmed that all written informed consent was obtained.

## Competing interest

The authors declare no conflicts of interest.

## Funding

There is no fund support for this work.

## Authors' contributions

Jun Yang and Shaohua Wang conceived and designed the study and wrote the manuscript.

Jiaying Zhou and Cuili Li analyzed the data.

All authors reviewed and approved the final manuscript.

## Acknowledgements

Not applicable.

## References

1. Kamihara J, Bourdeaut F, Foulkes WD, Molenaar JJ, Mossé YP, Nakagawara A, Parareda A, Scollon SR, Schneider KW, Skalet AH *et al*: **Retinoblastoma and Neuroblastoma Predisposition and Surveillance**. *Clin Cancer Res* 2017, **23**(13).
2. Matthay KK, Maris JM, Schleiermacher G, Nakagawara A, Mackall CL, Diller L, Weiss WA: **Neuroblastoma**. *Nat Rev Dis Primers* 2016, **2**:16078.
3. Pinto NR, Applebaum MA, Volchenboun SL, Matthay KK, London WB, Ambros PF, Nakagawara A, Berthold F, Schleiermacher G, Park JR *et al*: **Advances in Risk Classification and Treatment Strategies for Neuroblastoma**. *J Clin Oncol* 2015, **33**(27):3008-3017.
4. Oldridge DA, Truong B, Russ D, DuBois SG, Vaksman Z, Mosse YP, Diskin SJ, Maris JM, Matthay KK: **Differences in Genomic Profiles and Outcomes Between Thoracic and Adrenal Neuroblastoma**. *J Natl Cancer Inst* 2019, **111**(11):1192-1201.

5. Jones DTW, Banito A, Grünewald TGP, Haber M, Jäger N, Kool M, Milde T, Molenaar JJ, Nabbi A, Pugh TJ *et al*: **Molecular characteristics and therapeutic vulnerabilities across paediatric solid tumours.** *Nat Rev Cancer* 2019, **19**(8):420-438.
6. Hentze MW, Castello A, Schwarzl T, Preiss T: **A brave new world of RNA-binding proteins.** *Nat Rev Mol Cell Biol* 2018, **19**(5):327-341.
7. Ramanathan M, Majzoub K, Rao DS, Neela PH, Zarnegar BJ, Mondal S, Roth JG, Gai H, Kovalski JR, Siprashvili Z *et al*: **RNA-protein interaction detection in living cells.** *Nat Methods* 2018, **15**(3):207-212.
8. Gerstberger S, Hafner M, Tuschl T: **A census of human RNA-binding proteins.** *Nat Rev Genet* 2014, **15**(12):829-845.
9. Chu C, Zhang QC, da Rocha ST, Flynn RA, Bharadwaj M, Calabrese JM, Magnuson T, Heard E, Chang HY: **Systematic discovery of Xist RNA binding proteins.** *Cell* 2015, **161**(2):404-416.
10. Pereira B, Billaud M, Almeida R: **RNA-Binding Proteins in Cancer: Old Players and New Actors.** *Trends Cancer* 2017, **3**(7):506-528.
11. Dominguez D, Freese P, Alexis MS, Su A, Hochman M, Palden T, Bazile C, Lambert NJ, Van Nostrand EL, Pratt GA *et al*: **Sequence, Structure, and Context Preferences of Human RNA Binding Proteins.** *Mol Cell* 2018, **70**(5).
12. Lee FCY, Ule J: **Advances in CLIP Technologies for Studies of Protein-RNA Interactions.** *Mol Cell* 2018, **69**(3):354-369.
13. Gupta SK, Garg A, Bär C, Chatterjee S, Foinquinos A, Milting H, Streckfuß-Bömeke K, Fiedler J, Thum T: **Quaking Inhibits Doxorubicin-Mediated Cardiotoxicity Through Regulation of Cardiac Circular RNA Expression.** *Circ Res* 2018, **122**(2):246-254.
14. Zu T, Cleary JD, Liu Y, Bañez-Coronel M, Bubenik JL, Ayhan F, Ashizawa T, Xia G, Clark HB, Yachnis AT *et al*: **RAN Translation Regulated by Muscleblind Proteins in Myotonic Dystrophy Type 2.** *Neuron* 2017, **95**(6).
15. Bennett CL, Dastidar SG, Ling S-C, Malik B, Ashe T, Wadhwa M, Miller DB, Lee C, Mitchell MB, van Es MA *et al*: **Senataxin mutations elicit motor neuron degeneration phenotypes and yield TDP-43 mislocalization in ALS4 mice and human patients.** *Acta Neuropathol* 2018, **136**(3):425-443.
16. Chatterji P, Rustgi AK: **RNA Binding Proteins in Intestinal Epithelial Biology and Colorectal Cancer.** *Trends Mol Med* 2018, **24**(5):490-506.
17. Yu G, Wang L-G, Han Y, He Q-Y: **clusterProfiler: an R package for comparing biological themes among gene clusters.** *OMICS* 2012, **16**(5):284-287.
18. Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, Simonovic M, Doncheva NT, Morris JH, Bork P *et al*: **STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets.** *Nucleic Acids Res* 2019, **47**(D1):D607-D613.
19. Bader GD, Hogue CWV: **An automated method for finding molecular complexes in large protein interaction networks.** *BMC Bioinformatics* 2003, **4**:2.
20. Ackermann S, Cartolano M, Hero B, Welte A, Kahlert Y, Roderwieser A, Bartenhagen C, Walter E, Gecht J, Kerschke L *et al*: **A mechanistic classification of clinical phenotypes in neuroblastoma.** *Science (New York, NY)* 2018, **362**(6419):1165-1170.



21. Fletcher JI, Ziegler DS, Trahair TN, Marshall GM, Haber M, Norris MD: **Too many targets, not enough patients: rethinking neuroblastoma clinical trials.** *Nat Rev Cancer* 2018, **18**(6):389-400.
22. Wang E, Lu SX, Pastore A, Chen X, Imig J, Chun-Wei Lee S, Hockemeyer K, Ghebrechristos YE, Yoshimi A, Inoue D *et al*: **Targeting an RNA-Binding Protein Network in Acute Myeloid Leukemia.** *Cancer Cell* 2019, **35**(3).
23. Lujan DA, Ochoa JL, Hartley RS: **Cold-inducible RNA binding protein in cancer and inflammation.** *Wiley Interdiscip Rev RNA* 2018, **9**(2).
24. Degrauwe N, Suvà M-L, Janiszewska M, Riggi N, Stamenkovic I: **IMPs: an RNA-binding protein family that provides a link between stem cell maintenance in normal development and cancer.** *Genes Dev* 2016, **30**(22):2459-2474.
25. Chen Y, Bao C, Zhang X, Lin X, Huang H, Wang Z: **Long non-coding RNA HCG11 modulates glioma progression through cooperating with miR-496/CPEB3 axis.** *Cell Prolif* 2019, **52**(5):e12615.
26. Liu F, Zhang G, Lv S, Wen X, Liu P: **miRNA-301b-3p accelerates migration and invasion of high-grade ovarian serous tumor via targeting CPEB3/EGFR axis.** *J Cell Biochem* 2019, **120**(8):12618-12627.
27. Zhong Q, Fang Y, Lai Q, Wang S, He C, Li A, Liu S, Yan Q: **CPEB3 inhibits epithelial-mesenchymal transition by disrupting the crosstalk between colorectal cancer cells and tumor-associated macrophages via IL-6R/STAT3 signaling.** *J Exp Clin Cancer Res* 2020, **39**(1):132.
28. Zou C-D, Zhao W-M, Wang X-N, Li Q, Huang H, Cheng W-P, Jin J-F, Zhang H, Wu M-J, Tai S *et al*: **MicroRNA-107: a novel promoter of tumor progression that targets the CPEB3/EGFR axis in human hepatocellular carcinoma.** *Oncotarget* 2016, **7**(1):266-278.
29. Zhang Y, Yu R, Li L: **LINC00641 hinders the progression of cervical cancer by targeting miR-378a-3p/CPEB3.** *J Gene Med* 2020:e3212.
30. Rapino F, Delaunay S, Rambow F, Zhou Z, Tharun L, De Tullio P, Sin O, Shostak K, Schmitz S, Piepers J *et al*: **Codon-specific translation reprogramming promotes resistance to targeted therapy.** *Nature* 2018, **558**(7711):605-609.
31. Zhang M, Iyer RR, Azad TD, Wang Q, Garzon-Muvdi T, Wang J, Liu A, Burger P, Eberhart C, Rodriguez FJ *et al*: **Genomic Landscape of Intramedullary Spinal Cord Gliomas.** *Sci Rep* 2019, **9**(1):18722.

## Tables

Tables 1 - 3 are available as downloads in the Supplementary Files.

## Figures



(caption included in image)

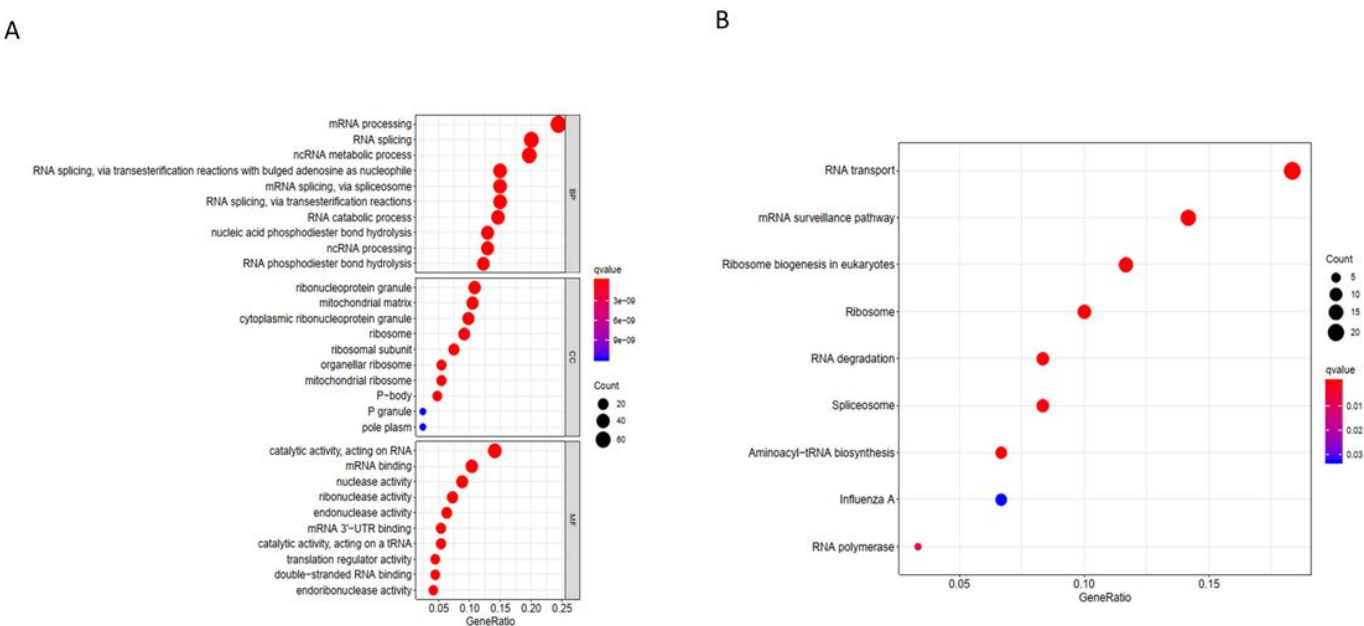


Figure 3

(caption included in image)

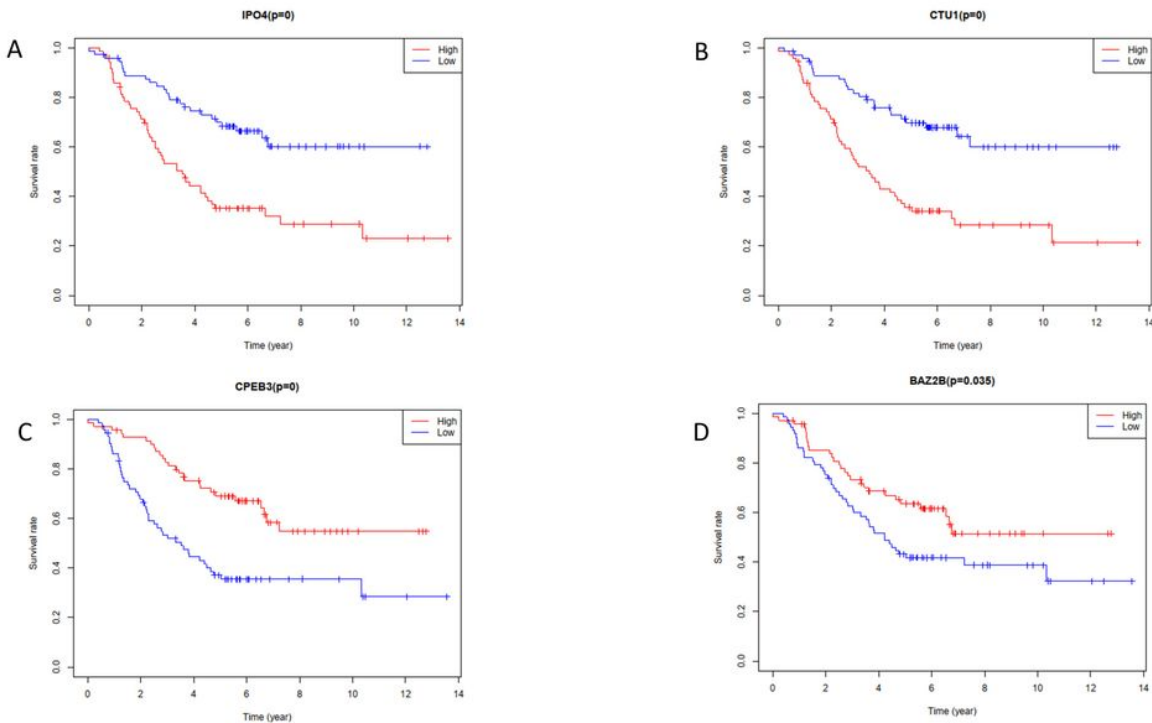


Figure 4

(caption included in image)

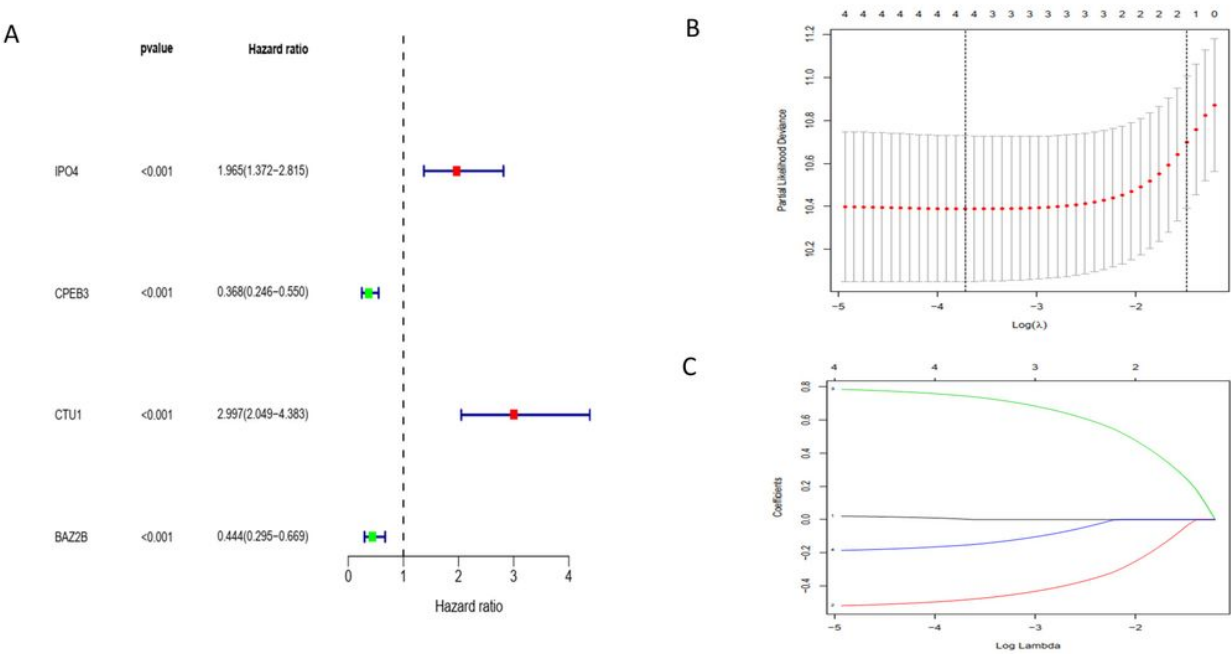


Figure 5

(caption included in image)

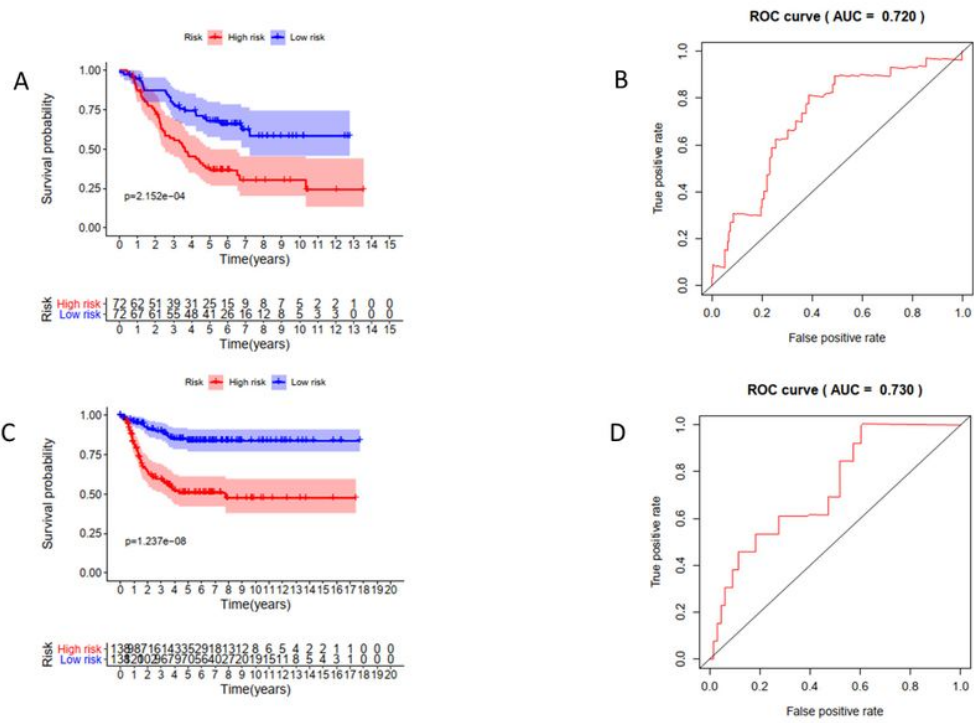


Figure 6. Survival analysis and prognostic risk assessment of 2-hub genes model for NB patients. (A,B) TARGET cohort, (C,D) GSE85047 cohort.

Figure 6

(caption included in image)

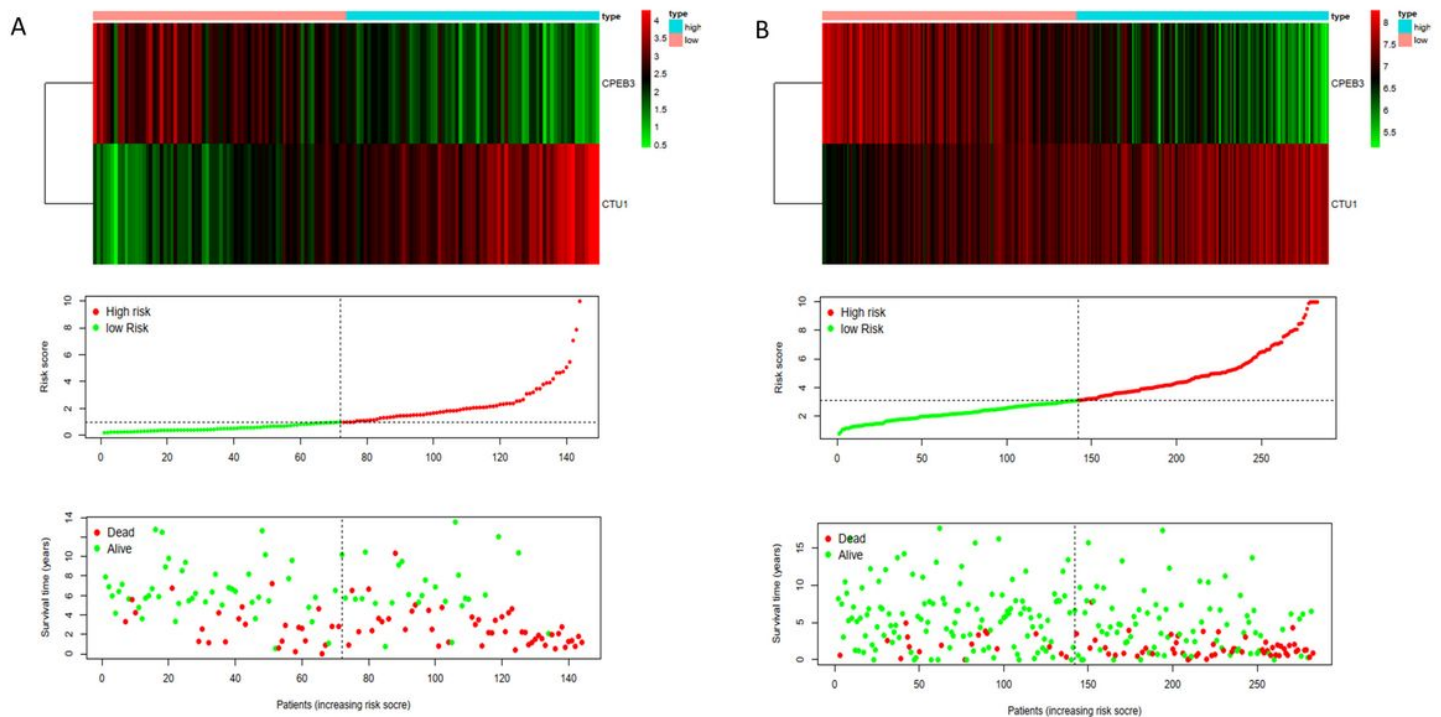
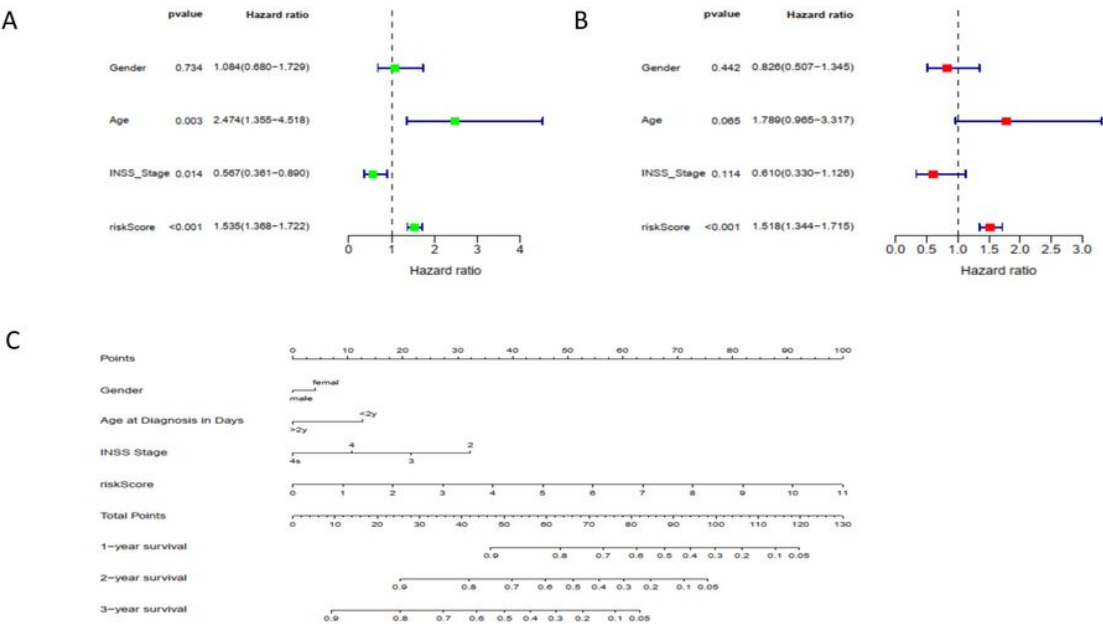


Figure 7. Risk score analysis of 2-gene prognostic model.(A) TARGET cohort. (B) GSE85047 cohort.

Figure 7

(caption included in image)



**Figure 8** The nomogram can predict the prognosis probability in NB. A. Univariate Cox regression analysis. Forest plot of associations between risk factors and the survival of NB. B. Multiple Cox regression analysis. The RBPs gene signature is an independent predictor of NB. C. Nomogram of the NB cohort used to predict the OS.

**Figure 8**

(caption included in image)

# Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [Table1GO.docx](#)
- [Table2KEGG.docx](#)
- [Table3.docx](#)
- [TableS1IdentificationandvalidationofRNAbindingproteinassociatedprognosticmodelforNeuroblastoma.xls](#)