

Effect of six weeks 1000 mg/day Vitamin C supplementation and healthy training in elderly women on genes expression associated with immune response - a randomized controlled trial

Małgorzata Żychowska (✉ malgorzata.zychowska@ukw.edu.pl)

Uniwersytet Kazimierza Wielkiego w Bydgoszczy <https://orcid.org/0000-0002-9475-0448>

Agata Grzybkowska

Akademia Wychowania Fizycznego i Sportu im Jędrzeja Sniadeckiego w Gdansk

Mariusz Zasada

Uniwersytet Kazimierza Wielkiego w Bydgoszczy

Anna Piotrowska

Akademia Wychowania Fizycznego im Bronisława Czecha

Danuta Dworakowska

Uniwersytet Kazimierza Wielkiego w Bydgoszczy

Olga Czerwinska-Ledwig

Akademia Wychowania Fizycznego im Bronisława Czecha

Wanda Pilch

Akademia Wychowania Fizycznego im Bronisława Czecha

Jędrzej Antosiewicz

Gdański Uniwersytet Medyczny: Gdanski Uniwersytet Medyczny

Research article

Keywords: supplementation, aging, genes involved in immune response

DOI: <https://doi.org/10.21203/rs.3.rs-113420/v1>

License:  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Abstract

Background

In this study we investigated the effects of supplementation and exercise on the expression of genes associated with inflammation like *CCL2*, *CRP*, *IL 1*, *IL 6*, *IL 10* mRNA in elderly women.

Methods

Twenty four participants divided randomly into two groups were subjected to 6 weeks of the same health training program (three times per week). SUP group (supplemented, n = 12, mean age 72.8 ± 5.26 years and mean body mass 68.1 ± 8.3 kg) received 1000 mg of Vitamin C/day during training period, while CON group (control, n = 12, mean age 72.4 ± 5.5 years and body mass 67.7 ± 7.5 kg) received placebo.

Results

No significant changes in *IL-1*, *IL-6*, *IL-10* and *CRP* mRNA were observed within and between groups. However, there was a clear tendency of a decrease in *IL-6* (two-way ANOVA, significant for time) and an increase in *IL-10* mRNA noted in the supplemented group. A significant decrease in *CCL2* mRNA was observed only in the CON group (from $2^{0.2}$ to $2^{0.1}$, $p = 0.01$).

Conclusions

It can be concluded, that 6 weeks of supplementation and exercise was too short to obtain significant changes in gene expression in leukocytes, but supplementation of 1000 mg vitamin C positively affected *IL-6* and *IL-10* expression – which are key changes in the adaptation to training. However, changes in body mass, IL1 and CCL2 was positive in CON group. It is possible that Vitamin C during 6 weeks of supplementation could have different effects on the expression of individual genes involved in immune response.

Trial registration: retrospectively registered.

Introduction

Despite many years of research on the impact of antioxidant supplementation on the human health, until now there has been no clear opinion confirming or denying the need for supplementation in healthy subjects. It seems to be obvious that antioxidants should influence antioxidative status in the body. Therefore, supplementation should be effective in any case involving a possible increase in oxidative stress, for example in diseases (i.e. cardiovascular, cancer and others) [1, 2]. Increase in oxidative stress and inflammation also occurs during physical activity [3] and aging [4]. Thus, it seems to be an easy way to improve exercise tolerance as well as preventive therapy against aging [5]. Unfortunately, in the literature the data associated with helpful actions of antioxidant supplementation are ambiguous. For example, Righ et al. [6] and Chou et al. [7] postulated that vitamin C supplementation attenuates both

oxidative stress and inflammation in young males participants or Taekwondo athletes. Some studies have shown that despite an increase in antioxidative capacity following vitamin C supplementation, pro/antioxidative status did not change [3]. No changes in the pro/antioxidative balance was associated with the possible pro-oxidative effect of vitamin C [3, 8]. A decrease in inflammation could disturb normal adaptations to training. According to Mankowski et al. [9], exercise-induced oxidative stress is an important trigger to induce training adaptations. The data from literature in which supplementation was investigated is ambiguous. There are many reasons for differences in the literature data, including different responses to supplementation depending on dose used, type of exercise and age of participants [10, 11, 12].

Fast development of biomedical sciences has increased the average duration of human life, especially in developed countries. Aging is strongly associated with an increase in oxidative stress and inflammation [13]. According to Bektas et al. [14] in most aging individuals, a chronic low-grade proinflammatory state could be observed, and this factor can influence physical and psychological dysfunction. For this reason, in the current literature there are two concepts: healthy aging and inflammation aging [15, 16]. Chronic inflammation could be a cause of inflammation aging and promote development of many diseases such as neurodegenerative disease, cancer and others [17, 18]. Unfortunately, imbalance between pro and antioxidative status accompanies both aging and physical activity. It seems that vitamin C supplementation should be especially helpful for older people subjected to physical effort due to a reduction in prolonged oxidative stress and indirectly, inflammation.

Changes in oxidation status are important for cell survival. It is a main factor that could influence expression of genes associated with cellular stress response, e.g., genes encoding heat shock proteins and interleukins [19, 20]. Cell survival under stressful conditions depends on balance between activation of pro and antiapoptotic pathways [21]. Unfortunately, in the literature there are not many studies in which changes in genes expression caused by supplementation and exercise was investigated, especially in older participants. Recently, we investigated changes in expression of genes associated with iron metabolism, which is also easily induced by oxidative stress in elderly women (part of a wider project on vitamin C supplementation during health training). We concluded that vitamin C supplementation caused a decrease in ferritin mRNA [3]. Indirectly these findings indicate that vitamin C supplementation decreased intracellular free iron and consequently oxidative intracellular status. If this is true, the expression of genes associated with immune response, such as: *CCL2*, *CRP*, *IL-1*, *IL-6* and *IL-10* should change. These genes encode proteins easily induced by oxidative stress and physical exercise and have great importance for immune response and adaptation to effort. For example, monocyte chemoattractant protein 1, (*CCL2* - chemokine (C-C motif) ligand 2) is capable of recruiting monocytes, memory T cells, and dendritic cells to induce a proinflammatory response and is produced by tissue injury [22, 23]. C-reactive protein (*CRP* - encoded by *CRP* gene) is a non-specific marker of ongoing inflammation, tissue damage and necrosis. *IL-1* and *IL-6* are proinflammatory proteins (*IL-6* is inhibited by the expression of antiinflammatory *IL-10*). According to Ziemann et al., [24] increase in antiinflammatory proteins and decrease in proinflammatory expression is crucial for adaptation to exercises.

Until now, most of the data in which inflammation, exercise and aging were investigated concerned plasma or muscle changes caused by vitamin supplementation, such as oxidative and antioxidative status, cortisol, CRP or IL-6 level only in plasma. Thus, the aim of our study was investigation of changes in oxidative stress, antioxidative capacity and the expression of chosen genes associated with inflammation, during 6 weeks of health training supported and unsupported by 1000 mg vitamin C supplementation in elderly women. We assumed, that this supplementation could cause in decrease in intracellular oxidation and a decrease in expression of proinflammatory cytokines mRNA. We also expected an increase in *IL-10* mRNA as a positive effect of vitamin C supplementation during training in older participants.

Methods

Ethics

All procedures were approved by the Bioethics Committee at the Regional Medical Chamber in Gdansk (KB-10/16). Study protocol was constructed according to the Declaration of Helsinki, and all participants gave their written consent, were fully informed prior to participation, informed about the possibility of withdrawal at any time for any reason and were given the opportunity to view their results.

Subjects

From the group of 40 volunteers, 24 women meeting the inclusion criteria were selected. Women were qualified by a doctor to participate in the experiment and were not taking any medications on a permanent basis. None of the women took any supplements for 3 months before the experiment. Participants were divided randomly into two groups: supplemented group (SUP) $n = 12$, mean age 72.8 ± 5.26 years, mean body mass 68.1 ± 8.3 kg, and control group (CON) $n = 12$, mean age 72.4 ± 5.5 years, body mass 67.7 ± 7.5 kg. All participants has never been professional athletes. The participants did not know who received vitamin C supplementation or cellulose (products in caps, look similar, double-blinded).

Body mass analysis was performed using the InBody 720 composition analysis (In body, Department Poland). The basic characteristics of the SUP and CON are summarized in the results section (Table 1). To eliminate women with poor diets we also performed diet analysis and participants were asked not to change their nutritional habits during the study.

Table 1

Anthropometric characteristics of participants before and after 6 weeks of training supported (SUP group) or not supported (CON) Vitamin C supplementation.

Parameter	Group	I	II	p I vs. II	ANOVA 2-way	dCohen I	dCohen II
VO ₂ max. [ml/kg/min]	SUP	21.4 ± 3.6	22.1 ± 3.4	0.2	*row factor	0.23	0.03
	CON	22.2 ± 3.5	22.00 ± 3.5	0.3			
$p1$ (SUP I vs CON I) = 0.3; $p2$ (SUP II vs CON II) = 0.4							
Body mass [kg]	SUP	68.1 ± 8.3	67.0 ± 7.9	0.30	*row factor, time	0.05	0.42
	CON	67.7 ± 7.5	63.8 ± 7.5*#	0.01			
$p1$ (SUP I vs CON I) = 0.30; $p2$ (SUP II vs CON II) = 0.02							
Muscle mass [kg]	SUP	22.4 ± 2.3	23.3 ± 2.6#	0.79	*row factor	0.25	0.11
	CON	23.0 ± 2.5	23.6 ± 2.7	0.33			
$p1$ (SUP I vs CON I) = 0.01; $p2$ (SUP II vs CON II) = 0.36							
Fat mass [kg]	SUP	26.6 ± 7.1	24.2 ± 7.2	0.09	*row factor, time	0.44	0.46
	CON	23.4 ± 7.5#	20.8 ± 7.6*#	0.02			
$p1$ (SUP I vs CON I) = 0.04; $p2$ (SUP II vs CON II) = 0.02							
*significant differences ($p \leq 0.05$) between I and II within the groups; #significant differences ($p \leq 0.05$) between groups							

Training

The training program was similar to the one used and described earlier [3]. Mainly the principles of health training were followed. Women participated in a 6-week multidisciplinary training program (three times per week for 60 minutes in duration) consisting of gyrokinesis, stabilization training, and Nordic walking at moderate intensity. Total duration of the training period overall was 1080 minutes. Heart rate was monitored during each training session (Polar H1). Participants were asked not to exceed a heart rate of 130/min.

Supplementation

During the 6 weeks of training, the SUP group received 1000 mg of vitamin C (Max VitaC 1000, Colfarm, Poland) and CON group received cellulose in tablets (Colfarm, Poland). People conducting the experiment and participants did not know who was receiving supplementation and who was receiving placebo. The choice of vitamin C dose was chosen due to its easy availability and high frequency of consumption in the Polish population.

Determination of VO_2 max.

VO_2 max was determined using the cycloergometer Ergoline Ergoselect 150p (Jaeger OxyconPro) and a gas analyzer (Jaeger OxyconPro) using the direct method. The measurement procedure was as follows: 2 minutes for registration of resting values followed by 5 minutes of a warm-up with 30W load and 60 rpm cadence. Participants performed the test with the load gradually increasing by 10W/min. The test was stopped if the participant was unable to continue with a given power and the required term of 60 rpm, or if symptoms occurred which indicated the need to end the trial. The results of VO_2 max testing are presented in Table 1.

Blood collection, analysis of vitamin C concentration, total oxidative status (TOS)/total oxidative capacity (TOC), total antioxidative status (TAS)/total antioxidative capacity (TAC) analysis and analysis of gene expression.

Blood samples were collected twice: immediately before and 24 hours after the training period. Venous blood (5 mL) was collected into BD Vacutainer tubes (Becton Dickinson, USA), in order to evaluate TOS/TOC and TAS/TAC. Plasma was separated by centrifugation (3000 g at 4 °C for 10 minutes). Collected plasma was stored immediately at -80 °C for further analysis. We used photometric PerOx assay kit to evaluate TOS/TOC status (Immundiagnostik AG, Germany), and ImAnOx assay kit (Immundiagnostik AG, Germany) for TAS/TAC analysis. From the obtained results prooxidative/antioxidative balance was calculated.

Vitamin C plasma concentration measurement

Vitamin C concentration was determined in plasma using the method of Robitaille and Hoffer [25] described in our earlier paper [3].

Genetic evaluation

The protocol used for gene expression evaluation has been previously described in detail by Grzybkowska [26], Żychowska [3]. Briefly, 2 ml of venous blood was collected and to remove red blood cells, the Red Blood Cell Lysis Buffer (RBCL) (A&A Biotechnology, Gdynia, Poland) was added and the samples were then centrifuged. Obtained leukocytes were lysed using Fenzol (A&A Biotechnology, Gdynia, Poland) and stored at -20 °C. The isolation of total RNA was performed based on Chomczynski and Sacchi method [27] using 200 µL of chloroform (POCH, Gliwice, Poland), centrifugation, and 500 µL of isopropanol (POCH, Gliwice, Poland). The obtained pellet was washed twice in 1 ml of 75% ethanol and spun at 7500 g at 4 °C. The dried RNA was then resuspended in the PCR-grade water, and after

spectrophotometric evaluation of the purity and concentration of obtained material, the reverse transcription procedure was performed using 1000 ng of pure RNA, 0.2 μ M oligo(dT) and the Transcriptor First Strand cDNA Synthesis Kit as per the manufacturer's instructions (Roche, Warszawa, Poland). Immediately after this step, the samples were frozen and stored at -20°C without additional freeze-thaw cycles. For the quantitative real-time polymerase chain reaction (qRT-PCR) step, the 1:10 dilution of the cDNA has been used. This step was performed on AriaMx Real-Time PCR System (Agilent Technologies, Warszawa Poland) using FastStart Universal SYBR[®] Green Master (Rox) (Roche, Warszawa, Poland) according to the manufacturer's protocol. Three replicates of 2 μ L of diluted cDNA were used for qRT-PCR analysis. For each reaction, the melt curve analysis was performed to check for non-specific amplification.

The *TUBB* was used as a reference gene. All primers sequences were designed by authors in the Primer3 web tool and then in silico PCR tool in the USCS genome browser was used. All primers were delivered by Genomed, Warszawa, Poland. Primer sequences (5'-3') used to perform this experiment were as follows:

For *TUBB* (tubulin beta class I, NM_001293213): Forward primer: CTAGAACCTGGGACCATGGA and Reverse primer: TGCAGGCAGTCACAGCTCT

For *IL1* (NM_000575.5): Forward primer: AGT GCT GCT GAA GGA GAT GCC T and Reverse primer: CCT GCC AAG CAC ACC CAG TAG

For *IL6* (NM_000600.5): Forward primer: AAT TCG GTA CAT CCT CGA CGG and Reverse primer: GAA TCC AGA TTG GAA GCA TCC

For *IL10* (NM_000572.3); Forward primer: GAC ATC AAG GCG CAT GTG AAC Reverse primer: TCC ACG GCC TTG CTC TTG TTT

For *CCL2* (NM_002982.4); Forward primer: CAG CCA GAT GCA ATC AAT GCC Reverse primer: CTTGGCCACAATGGTCTTGAA

For *CRP* (NM_000567.3): Forward primer: TCG TTA ACG GTG CTT TGA GG and Reverse primer: TCT TGG TCT TGA CCA GCC TCT

Statistical analyses

First, the presence of a normal distribution was checked with the Shapiro-Wilk's test for all results. Data obtained before and after six weeks of training period were compared within each group using paired test. The differences between groups were analyzed using parametric or non-parametric (Wilcoxon) tests, as appropriate, as well as two-way ANOVA. The pro and antioxidative balance was calculated as the pro/anti ratio. Relative expression was calculated in Microsoft Excel 2015, using Schmittgen and Livak's method [28]. Data were transformed into linear values, and subjected to the same methodology as the other parameters. To determine the significance of differences between the groups in gene

expression, t-test and two-way ANOVA were used. Calculations and figures were generated using GraphPad Prism 6.0 software. P-values were considered statistically significant when ≤ 0.05 .

Results

Changes in body composition and VO_2 max.

Table 1 near here

Despite similar total body mass in both groups, significant differences were observed in muscle mass and fat mass before experiment (Table 1). Fat mass in CON group was significantly lower after the training period, with 6 weeks of training causing a significant decrease in total body mass and fat mass only in CON group. Therefore, significantly lower total body mass and fat was observed in CON group after 6 weeks of training without supplementation. No significant changes in VO_2 max were noted in either group. However, only body mass after intervention showed medium effect size (dCohen increase to 0.42).

Figure 1 near here

After 6 weeks of supplementation, vitamin C concentration increased in the SUP group from 13.9 ± 4.2 mg/L to 18.4 ± 8.6 mg/L, $p = 0.04$ (dCohen showed strong effect size at the level 0.81). At the end of experiment there was a significant difference in plasma vitamin C concentration between groups (18.4 ± 8.58 mg/L in SUP and 13.1 ± 5.2 mg/L in CON, $p = 0.03$, Fig. 1A). These changes did not significantly influence antioxidative and prooxidative capacity, however there was a minor tendency to increase prooxidative capacity observed in the SUP group (from 489.4 ± 149.4 to 538.7 ± 132.1 $\mu\text{mol/L}$). This tendency was not observed in the CON group. Despite no significant changes in antioxidative capacity, the slight tendency to increase in SUP and decrease in CON caused a significantly lower antioxidative status in CON (278.4 ± 24.6 $\mu\text{mol/L}$ in CON and 298.5 ± 32.9 $\mu\text{mol/L}$ in SUP, $p = 0.03$). A consistently similar tendency in pro/antioxidative ratio was observed with a slight tendency to increase in the SUP group. Effect size (dCohen was strong for prooxidative and antioxidative status, 0.52 and 0.70, respectively).

Figure 2 near here

There was no significant changes in gene expression for all tested genes observed after 6 weeks training supported by 1000 mg vitamin C supplementation (Fig. 2.). There was a tendency for decreased *IL-6* mRNA (from $2^{0.035}$ to $2^{0.016}$, two-way ANOVA, significant for time) and increase in *IL-10* mRNA (from $2^{0.005}$ to $2^{0.015}$) was noted for SUP group. Similar tendency in decrease in *IL-6* mRNA occurred in CON (change from $2^{0.035}$ to $2^{0.015}$). In both groups *CRP* and *IL-1* mRNA remained unchanged. There was a significant decrease in *CCL2* mRNA observed in the CON group (from $2^{0.2}$ to $2^{0.1}$, $p = 0.01$, two-way ANOVA, significant for time and subject). However, effect size (dCohen) was strong for three of tested genes: *IL 1* mRNA (1.09), *CCL2* mRNA (1.33) and *IL10* mRNA (2.91) only at the end of intervention.

Discussion

In our research we hypothesized, that vitamin C could increase antioxidant capacity and reduce oxidative stress and in this way lead to decrease in expression of genes induced by oxidative stress, such genes associated with inflammation. Unfortunately, our hypothesis have not been confirmed by research. In order to examine changes caused by vitamin C supplementation we selected women similar in body mass. As a result, the dose of vitamin C per kg of body weight was similar within supplemented group. Although we not assumed the differences in body composition at baseline and after 6 weeks of 1000 mg vitamin supplementation, our participants differed significantly in muscle mass and fat mass at baseline, despite no significant differences in total body mass and body mass index (BMI.) Moreover, a significant decrease in body mass and fat mass was observed in CON group after 6 weeks of training without supplementation (strong effect for total body mass). Thus, in the best case vitamin C did not influence body mass and fat loss. Opposite data was reported by Johnston et al. [29]. The authors postulated that plasma vitamin C concentration inversely related to body mass and markers of obesity, however 500 mg of vitamin C supplementation did not influence the circulating concentration of adiponectin (necessary for lipolysis) in adult participants. The hindered reduction of fat mass observed in our participants receiving supplementation could be associated with the high dose of vitamin C supplementation (1000 mg/day). It is possible, that this supplementation affects IL-6 expression in adipose tissue, leading to a decrease in IL-6 expression. In the literature there are some studies in which IL-6 is considered an important factor to induce fat loss by promoting white adipose tissue and browning lipolysis [30]. However, Wong et al. [31] postulated the negative relationship between intake of vitamin C and risk of developing metabolic syndrome, associated with obesity.

The concentration of vitamin C in the serum of our subject at baseline was average, and a significant increase was noted in SUP group after 6 weeks 1000 mg vitamin C supplementation (strong effect size). The final concentration was close to the upper limit of normal in supplemented group. Unfortunately, increase in vitamin C concentration was associated with a tendency to increase prooxidative and antioxidative status (strong effect size) and finally we observed tendency to increase pro/antioxidative balance. Generally, in both groups, six weeks of training supported or unsupported by vitamin C supplementation proved to be too short for any significant changes in pro or antioxidative status. Similar data after 6 weeks of supplementation during 12 weeks of training was observed in previously published data [3]. It seems, that 6 weeks of vitamin C supplementation in a dose of 1000 mg/day does not influence pro and antiinflammatory status, independently of body mass and BMI. Similar reports can be found in the literature. No changes in pro/antioxidative balance were reported by Bunpo and Antony [32], although their participants were healthy young men who received 250–500 mg vitamin C per day during 12 weeks of exercise. Moreover, the cited authors reported a decrease in antioxidative enzyme activity in erythrocytes. Assuming no influence of this supplementation on pro/antioxidative balance it is probable that vitamin C could show oxidative action, which is often indicated by other various authors [3, 33, 34]. It is possible, that vitamin C could promote generation of reactive oxygen species i.e., OH, O₂⁻, H₂O₂ or ferryl ion [3, 8].

In terms of the obtained results, changes in genes associated with immune response may not be consistent with our assumptions. Our hypothesis, that vitamin C supplementation in a dose of 1000 mg/day will cause a decrease in proinflammatory and an increase in anti-inflammatory gene expression was not confirmed. First of all, no significant changes in gene expression of all studied genes were noted in the SUP group. Only in the CON group, *CCL2* mRNA significantly decreased after the training period. This gene is not often investigated under exercising conditions, despite its important functions in inflammation. Overexpression of *IL-6* and *CCL2* could be a reason for the development of acute inflammation and inflammatory pain [35]. The most common changes in the *CCL2* expression in humans are indicated in peripheral blood mononuclear cells (PBMC) or leukocytes. According to Strömberg et al. [36] its expression in muscle cells was marginal. *CCL2* mRNA and CCL2 protein are responsible for macrophage induction. It is well documented, that *CCL2* released by astrocytes has an impact on cognitive dysfunction and brain inflammation [37, 38]. It means that a decrease in expression may have a positive impact on cognitive function and decrease brain inflammation. Thus, obtained results showed positive changes affected by training in those not supported by vitamin C supplementation.

In the literature there is data regarding the influence of vitamin C supplementation mainly on plasma CRP. Bianias et al. [39] reported that vitamin C supplementation is associated with a decrease in CRP in patients undergoing hemodialysis. Ellulu et al. [40] reported that vitamin C (500 mg twice daily) has potential effects in alleviating inflammatory status by reducing high-sensitivity C-reactive protein (hs-CRP) and IL-6 in hypertensive and/or diabetic obese patients [41]. Unfortunately, their research was conducted on sick people, for which vitamin C supplementation may have different effects than in healthy people, when subjected to effort. Results of ultramarathon runners showed that vitamin C caused an increase in CRP by reducing the secretion of cortisol [42]. Our results (no changes in CRP mRNA in both groups) are consistent with the data published by Righi et al. [6]. The authors observed no changes in CRP following single both of acute exercise.

It is well documented that changes in IL-1 caused by exercise are less than another in cytokines, e.g. IL-6. The available data shows that the increase in IL-1 is dependent on training load. Significant increase in this cytokine was observed after ultramarathon, but only in people supplemented by 1500 mg of vitamin C (at a dose of 500 mg of vitamin C there was no such effect). This increase was small in comparison to IL-6. No changes in *IL-1* mRNA were observed in our study which confirms lower induction of this cytokine gene expression by effort and no effect of six weeks of 1000 mg/day vitamin C supplementation.

In both groups the same tendency to decrease in *IL-6* mRNA was observed. Thus, this effect is mainly associated with training. Our results are compatible with data from other literature studying the influence of aerobic exercise on the modulation of the cytokine profile. Although available data is mainly associated with changes in inflammatory (IL-6 or IL-1) protein measurement in plasma or serum. According to Elkader and All Jiffri [43], significant decrease in IL-6 and increase in IL-10 was observed following 6 months of aerobic exercise in elderly subjects. Thus, 6 weeks of training may be too little to obtain significant differences.

Moreover, Canali et al. [44] suggested a small effect of vitamin C supplementation on gene expression, especially those associated with immune response. In the cited research the authors investigated changes in gene expression in five volunteers supplemented high dose of vitamin C (1 g/day) over 5 days. They suggested that vitamin C plays important role in the modulation of *IL-10* mRNA during an inflammatory stimulus. We agree with these authors that in healthy, well nourished participants, supplementation of vitamin C is “buffered” within a homeostatic physiological equilibrium. However, tendency to increase IL-10 with a simultaneous decrease in IL-6 is important in terms of adaptation to training [24]. This was clear in the supplemented group, however, 6 weeks of supplementation did not significantly influence this change.

Conclusions

Generally, six weeks of training with moderate intensity was insufficient to obtain significant increase in VO_2 max. and it seems to be an important result for planning the training of the elderly people. It seems that 1000 mg/day vitamin C supplementation inhibited changes in body composition in elderly women. Applied six weeks of health training did not influence oxidative/antioxidative balance however the marked prooxidative effect was noticeable. Moreover, the tendency to decrease *IL-6* and increase *IL-10* mRNA in the supplemented group could indirectly indicate that oxidative stress within cells was lowered. However, most strong effect size was observed for *IL1* and *CCL2* mRNA, but these observations require further research. As there were no significant changes in pro/antioxidative balance with simultaneous change in *IL-6* and *IL-10* mRNA this change could be considered a positive effect of 1000 mg vitamin C supplementation. However, these effects were not spectacular, and it is not known whether a longer period of supplementation would have had an effect on oxidative/prooxidative balance in plasma.

Study Limitations

Our study has some limitations. Despite randomization, differences in body composition made it difficult to interpret the results. Also, only one dose (1000 mg daily) of vitamin C was used, and its effect can be expected to be dose dependent. Moreover, the results cannot be directly extrapolated to men and women of other ages.

Abbreviations

CCL2

gene encoding C-C Motif Chemokine Ligand

CRP

gene encoding C-reactive protein

IL1

gene encoding interleukin 1

IL6

gene encoding interleukin 6

IL10

gene encoding interleukin 10

CON

control group

SUP

supplemented group

TOS/TOC

total oxidative status/total oxidative capacity

TAS/TAC

total anti-oxidative status/total antioxidative capacity

TUBB

gene encoding tubulin B

RNA

ribonucleic acid

VO₂ max.

maximal oxygen consumption

Declarations

Ethics approval and consent to participate

All procedures were approved by the Bioethics Committee at the Regional Medical Chamber in Gdansk (KB-10/16). Study protocol was constructed according to the Declaration of Helsinki, and all participants gave their written consent, were fully informed prior to participation, informed about the possibility of withdrawal at any time for any reason and were given the opportunity to view their results.

Consent for publication

Not applicable

Availability of data and materials

The datasets used and/or analysed during the current study are available from the first author: Małgorzata Żychowska (malgorzata.zychowska@ukw.edu.pl) on reasonable request.

Competing interests

The authors declare that they have no competing interests.

Funding

This research was funded by Polish Ministry of Science and Higher Education nr 169/BS/KK/2018.

Authors' contributions

MŽ was the creator of the project idea, performer the genetic analysis, statistics, supervised the course of research, analysis of results and wrote the manuscript; AG conducted the research and wrote the manuscript; MZ performed statistical analysis; AP conducted biochemical analysis and revised the manuscript; DD performer genetic analysis, OCL conducted biochemical analysis – Vitamin C concentration,; reviewed the literature; WP was responsible for biochemical analysis and interpretation of results, JA data interpretation and supervision.

Acknowledgements

Not applicable.

References

1. Balsano C, Alisi, A. Antioxidant Effects of Natural Bioactive Compounds. *Curr. Pharm. Des.* 2009;15:3063-73. <https://doi.org/10.2174/138161209789058084>
2. Lee CH, Chan RSM, Wan HYL, Woo YC, Cheung CY, Fong CHY, et al. Dietary Intake of Anti-Oxidant Vitamins A, C, and E is Inversely Associated with Adverse Cardiovascular Outcomes in Chinese-A 22-Years Population-Based Prospective Study. 2018;4:10. <https://doi.org/10.3390/nu10111664>
3. Żychowska M, Grzybkowska A, Wiech M, Urbański R, Pilch W, Piotrowska A, et al. Exercise Training and Vitamin C Supplementation Affects Ferritin mRNA in Leukocytes without Affecting Prooxidative/Antioxidative Balance in Elderly Women. *Int J Mol Sci.* 2020;21:E6469. <https://doi.org/3390/ijms21186469>.
4. Milisav I, Ribarič S, Poljsak Antioxidant Vitamins and Ageing. *Subcell Biochem.* 2018,90,1-23. https://doi.org/10.1007/978-981-13-2835-0_1
5. Neuffer PD, Bamman MM, Muoio DM, Bouchard C, Cooper DM, Goodpaster BH, et al. Understanding the Cellular and Molecular Mechanisms of Physical Activity-Induced Health Benefits. *Cell Metab.* 2015,22,4-11. <https://doi.org/10.1016/j.cmet.2015.05.011>
6. Righi NC, Schuch FB, De Nardi AT, Pippi CM, de Almeida Righi G, Puntel GO, et al. Effects of vitamin C on oxidative stress, inflammation, muscle soreness, and strength following acute exercise: meta-analyses of randomized clinical trials. *Eur J Nutr.* 2020;59:2827-39. <https://doi.org/10.1007/s00394-020-02215-2>.
7. Chou CC, Sung YC, Davison G, Chen CY, Liao YH. Short-Term High-Dose Vitamin C and E Supplementation Attenuates Muscle Damage and Inflammatory Responses to Repeated Taekwondo Competitions: A Randomized Placebo-Controlled Trial. *Int J Med Sci.* 2018;15:1217-26. <https://doi.org/10.7150/ijms.26340>.
8. Stadtman ER. Ascorbic acid and oxidative inactivation of proteins. *Am. J. Clin. Nutr.* 1991;54:1125-28. <https://doi.org/1093/ajcn/54.6.1125s>
9. Mankowski RT, Anton SD, Buford TW, Leeuwenburgh C. Dietary Antioxidants as Modifiers of Physiologic Adaptations to Exercise. *Med. Sci. Sports Exerc.* 2015;47:1857-68.

<https://doi.org/1249/MSS.0000000000000620>

10. Chou CC, Sung YC, Davison G, Chen CY, Liao YH. Short-Term High-Dose Vitamin C and E Supplementation Attenuates Muscle Damage and Inflammatory Responses to Repeated Taekwondo Competitions: A Randomized Placebo-Controlled Trial. *Int. J. Med. Sci.* 2018;30:1217-26. <https://doi.org/10.7150/ijms.26340>
11. Paulsen G, Cumming KT, Holden G, Hallén J, Rønnestad B, Sveen O, et al. Vitamin C and E supplementation hampers cellular adaptation to endurance training in humans: A double-blind, randomised, controlled trial. *J. Physiol.* 2014;15:1887-901. <https://doi.org/10.1113/jphysiol.2013.267419>
12. Yfanti C, Soren AT, Anders NR, Remi M, Ole MH, Jens L, et al. Antioxidant Supplementation Does Not Alter Endurance Training Adaptation. *Med. Sci. Sports Exerc.* 2010;42:1388-95. <https://doi.org/10.1249/MSS.0b013e3181cd76be>
13. Sallam N, Laher I. Exercise modulates oxidative stress and inflammation in aging and cardiovascular diseases. *Oxidative Med. Cell. Longev.* 2016;46-54. <https://doi.org/10.1155/2016/7239639>
14. Pence BD, Yarbrow JR. Aging impairs mitochondrial respiratory capacity in classical monocytes. *Exp Gerontol.* 2018;108:112-17. <https://doi.org/10.1016/j.exger.2018.04.008>
15. Redman LM, Smith SR, Burton JH, Martin CK, Il'yasova D, Ravussin E. Metabolic Slowing and Reduced Oxidative Damage with Sustained Caloric Restriction Support the Rate of Living and Oxidative Damage Theories of Aging. *Cell Metab.* 2018;27:805-15. <https://doi.org/10.1016/j.cmet.2018.02.019>
16. El Assar M, Angulo J, Rodríguez-Mañas L. Oxidative stress and vascular inflammation in aging. *Free Radic. Biol. Med.* 2013;65:380-401. <https://doi.org/10.1016/j.freeradbiomed.2013.07.003>
17. Tsuma-Kaneko M, Sawanobori M, Kawakami S, Uno T, Nakamura Y, Onizuka M, et al. Iron removal enhances vitamin C-induced apoptosis and growth inhibition of K-562 leukemic cells. *Sci. Rep.* 2018;26:17377. <https://doi.org/10.1038/s41598-018-35730-8>
18. Gaman AM, Buga AM, Gaman MA, Popa-Wagner A. The Role of Oxidative Stress and the Effects of Antioxidants on the Incidence of Infectious Complications of Chronic Lymphocytic Leukemia. *Oxidative Med. Cell. Longev.* 2014;2014:158135. <https://doi.org/10.1155/2014/158135>
19. Kochanowicz A, Sawczyn S, Niespodziński B, Mieszkowski J, Kochanowicz K, Żychowska M. Cellular Stress Response Gene Expression During Upper and Lower Body High Intensity Exercises. *PLoS One.* 2017;12:e0171247. <https://doi.org/10.1371/journal.pone.0171247>
20. Żychowska M, Nowak-Zaleska A, Chruściński G, Zaleski R, Mieszkowski J, Niespodziński B, et al. Association of High Cardiovascular Fitness and the Rate of Adaptation to Heat Stress. *Biomed Res Int.* 2018;2018:1685368. <https://doi.org/10.1155/2018/1685368>
21. Szołtysek K, Janus P, Widlak P. The NF-κB dependent cellular signaling pathway and its interference with p53 and HSF1-dependent pathways. *Advances in Cell Biology.* 2011;38:159-75. (in Polish)
22. Carr MW, Roth SJ, Luther E, Rose SS, Springer TA. Monocyte chemoattractant protein 1 acts as a T-lymphocyte chemoattractant". *Proceedings of the National Academy of Sciences of the United States of America.* 1999;96:1081-1086.

- States of America. 1994;91:3652-56. <https://doi.org/1073/pnas.91.9.3652>.
23. Xu LL, Warren MK, Rose WL, Gong W, Wang JM. Human recombinant monocyte chemotactic protein and other C-C chemokines bind and induce directional migration of dendritic cells in vitro. *J Leukoc Biol.* 1996;60:365-71. <https://doi.org/1002/jlb.60.3.365>.
24. Ziemann E, Zembron-Lacny A, Kasperska A, Antosiewicz J, Grzywacz T, Garsztko T, et al. Exercise training-induced changes in inflammatory mediators and heat shock proteins in young tennis players. *J Sports Sci Med.* 2013;12:282-9.
25. Robilaile R, Hoffer J. A simple method for plasma total vitamin C analysis suitable for routine clinical laboratory use. *Nutr. J.* 2016;15:40. <https://doi.org/10.1186/s12937-016-0158-9>
26. Grzybkowska A, Anczykowska K, Ratkowski W, Aschenbrenner P, Antosiewicz J, Bonisławska I, et al. Changes in Serum Iron and Leukocyte mRNA Levels of Genes Involved in Iron Metabolism in Amateur Marathon Runners-Effect of the Running Pace. *Genes (Basel).* 2019;10:460. <https://doi.org/10.3390/genes10060460>.
27. Chomczynski P, Sacchi N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal. Biochem.* 1987;162:156-159. <https://doi.org/10.1006/abio.1987.9999>
28. Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative C (T) method. *Natur Protoc.* 2008;3:1101-8. <https://doi.org/1038/nprot.2008.73>
29. Johnston CS, Beezhold BL, Mostow B, Swan PD. Plasma vitamin C is inversely related to body mass index and waist circumference but not to plasma adiponectin in nonsmoking adults. *J Nutr.* 2007;137:1757-62. <https://doi.org/10.1093/jn/137.7.1757>.
30. Han J, Meng Q, Shen L, Wu G. Interleukin-6 induces fat loss in cancer cachexia by promoting white adipose tissue lipolysis and browning. *Lipids Health Dis.* 2018;17:14. <https://doi.org/10.1186/s12944-018-0657-0>.
31. Wong SK, Chin KY, Ima-Nirwana S. Vitamin C: A Review on its Role in the Management of Metabolic Syndrome. *Int J Med Sci.* 2020;17:1625-38. <https://doi.org/10.7150/ijms.47103>.
32. Bunpo P, Anthony TG. Ascorbic acid supplementation does not alter oxidative stress markers in healthy volunteers engaged in a supervised exercise program. *Appl. Physiol. Nutr. Metab.* 2016;41:175-80. <https://doi.org/10.1139/apnm-2015-0285>
33. Gomez-Cabrera MC, Domenech E, Romagnoli M, Arduini A, Borrás C, Pallardo FV, et al. Oral administration of vitamin C decreases muscle mitochondrial biogenesis and hampers training-induced adaptations in endurance performance. *Am. J. Clin. Nutr.* 2008;87:142-9. <https://doi.org/10.1093/ajcn/87.1.142>
34. Meier P, Renga M, Hoppeler H, Baum O. The impact of antioxidant supplements and endurance exercise on genes of the carbohydrate and lipid metabolism in skeletal muscle of mice. *Cell Biochem. Funct.* 2013;31:51-9. <https://doi.org/10.1002/cbf.2859>
35. Wang XM, Hamza M, Wu TX, Dionne RA. Upregulation of IL-6, IL-8 and CCL2 gene expression after acute inflammation: Correlation to clinical pain. *Pain.* 2009;142:275-83.

<https://doi.org/10.1016/j.pain.2009.02.001>.

36. Strömberg A, Olsson K, Dijksterhuis JP, Rullman E, Schulte G, Gustafsson T. CX3CL1-a macrophage chemoattractant induced by a single bout of exercise in human skeletal muscle. *Am J Physiol Regul Integr Comp Physiol*. 2016;310:R297-304. <https://doi.org/10.1152/ajpregu.00236.2015>.
37. Gruol DL. Impact of Increased Astrocyte Expression of IL-6, CCL2 or CXCL10 in Transgenic Mice on Hippocampal Synaptic Function. *Brain sciences*. 2016;6. <https://doi.org/10.3390/brainsci6020019>.
38. Xu J, Dong H, Qian Q, Zhang X, Wang Y, Jin W et al. Astrocyte-derived CCL2 participates in surgery-induced cognitive dysfunction and neuroinflammation via evoking microglia activation. *Behav. brain research*. 2017;332:145-53, <https://doi.org/10.1016/j.bbr.2017.05.066>.
39. Biniiaz V, Sadeghi Shermeh M, Ebadi A, Tayebi A, Einollahi B. Effect of Vitamin C Supplementation on C-reactive Protein Levels in Patients Undergoing Hemodialysis: A Randomized, Double Blind, Placebo-Controlled Study. *Nephrourol Mon*. 2013;6: e13351. <https://doi.org/10.5812/numonthly.13351>
40. Ellulu MS, Rahmat A, Patimah I, Khaza'ai H, Abed Y. Effect of vitamin C on inflammation and metabolic markers in hypertensive and/or diabetic obese adults: a randomized controlled trial. *Drug Des Devel Ther*. 2015;9:3405-12. <https://doi.org/10.2147/DDDT.S83144>.
41. Peters EM, Anderson R, Theron Attenuation of increase in circulating cortisol and enhancement of the acute phase protein response in vitamin C-supplemented ultramarathoners. *Int J Sports Med*. 2001;22:120-6. <https://doi.org/10.1055/s-2001-11364>.
42. Nieman DC, Peters EM, Henson DA, Nevines EO, Thompson Influence of vitamin C supplementation on cytokine changes following an ultramarathon. *J Interferon Cytokine Res*. 2000;20:1029-35. <https://doi.org/10.1089/10799900050198480>
43. Abd El-Kader SM, Al-Jiffri OH. Aerobic exercise modulates cytokine profile and sleep quality in elderly. *Afr Health Sci*. 2019;19:2198-207. <https://doi.org/10.4314/ahs.v19i2.45>.
44. Canali R, Natarelli L, Leoni G, Azzini E, Comitato R, Sancak O, et al. Vitamin C supplementation modulates gene expression in peripheral blood mononuclear cells specifically upon an inflammatory stimulus: a pilot study in healthy subjects. *Genes Nutr*. 2014;9:390. <https://doi.org/10.1007/s12263-014-0390-x>.