

Reference Intervals for 26 Common Biochemical Analytes in Term Neonates in Jilin Province, China

Kaijin Wang

First Hospital of Jilin University

Xuetong Zhu

First Hospital of Jilin University

Qi Zhou

First Hospital of Jilin University

Jiancheng Xu (✉ xjc@jlu.edu.cn)

First Hospital of Jilin University

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Abstract

Background: Biochemical analytes provide information for neonatal disease management and therapy, and population-based reference intervals (RIs) are essential to accurately interpret laboratory test results. This study aimed to establish local RIs for biochemical assays in term neonates.

Methods: A total of 195 healthy term neonates from birth to 3rd day were recruited as reference individuals prospectively. Analytes of 26 common biochemistries were measured using the VITROS 5600 Integrated System. The 3-level nested ANOVA was performed to assess the need for partitioning RIs of each analytes, and RIs were derived by a nonparametric method or robust method. Multiple regression analysis was used to evaluate specific correlations between the analytes and individual characteristics including age, gender, gestational age, birthweight and delivery mode.

Results: There were no between-sex differences in all analytes, whereas there were significant between-day-age differences in 6 analytes. Small between-delivery-mode differences were observed in the results for potassium, phosphate, and urea. The major related factor of most analytes was postnatal age. During the first 3 days, values of iron, lipids and lipoproteins increased; creatinine, urea, uric acid, creatine kinase and lactate dehydrogenase decreased; other analytes showed slight changes or relatively stable trends. Reference limits of some analytes, particularly lactate dehydrogenase and alkaline phosphatase, were significant different from adult and pediatric groups.

Conclusions: RIs of 26 common biochemical analytes are established for term neonates aged 0 to 3 days in northeast China. Additionally, it is suggested that age-related changes should be valued in the clinical decision-making process for newborns.

Introduction

Reference intervals (RIs) are determined from individuals in healthy status, serving as tools for interpreting test results. Despite the well standardized measurements of major laboratory test, it is difficult to establish harmonized and uniform RIs on a global scale. Previous research has found that the differences in RIs of analytes are due to their complex patterns of biological sources of variations, such as region, race, diet, and more individual characteristic [1, 2]. Data from the studies we have published confirms that establishing age- and sex-appropriate local RIs is more accurate and reliable for providing clinical decision support, especially in pediatrics [3-6].

Recently, there has been renewed interest in neonatal RIs with increased attention to neonatal health screening and management [7, 8]. Anthropometry and laboratory data are useful for neonatal growth and nutrition assessment [9]. The common biochemical analytes are related to measuring the functional maturity of organ systems, monitoring the changes of electrolyte and acid-base status, and evaluating the ability of material absorption and metabolism. Hence they can not only help diagnose relevant diseases but also be engaged in population pharmacokinetic research. However, the Health Occupation Standards of China does not yet provide the RIs of common biochemical analytes for newborns, and most clinical laboratories in China adopt RIs from textbooks, reagent inserts, or adult reference standards. These data might be obsolete, lack of any traceability, or established on mismatched population in race or age, and then would be misleading to interpretate laboratory results. Thus, this study aimed to investigate the RIs for newborns during the first days in Jilin Province, China.

The Clinical and Laboratory Standards Institute (CLSI) recommends a priori method to determine RIs, namely selecting healthy individuals defined by specific reference criteria as reference populations in advance [10]. Another commonly used method, posteriori selection process, is able to establish RIs using clinical large databases. One drawback of the retrospective study is that its result would not be ideal for newborns, because it is limited to complicated pathologic condition and unclear information about specimen collection. However, recruitment of healthy infants is often difficult because of ethical constraints for collecting unnecessary blood samples. We conducted this research with written informed consent from parents, and the specimen collection procedure was evaluated by qualified pediatricians to ensure rationality and applicability in local medical practice.

It is now widely acknowledged that term infants are different from preterm infants in the measured values of most biochemistry analytes [11-13]. Meanwhile, the two groups of infants have quite different causes of hospitalization. The premature are more susceptible to severe diseases for their immature organ function; whereas the common reasons for term infants to be hospitalized are birth injury, feeding intolerance, jaundice, sepsis and other medical complications. With this background, doctors should choose rational sets of tests and assess the results according to different criteria. In addition, most studies focus on establishing RIs and analyzing the correlations between individual parameters and analytes in newborns, who are born preterm or admitted to neonatal intensive care unit. However, detailed data about term infants are limited.

Therefore, this prospective study aimed to determine RIs of 26 common biochemical analytes for term newborns during the first 3 days, and investigate the associations between the analytes and neonatal characteristics.

Materials And Methods

Ethics

This study was approved by the Ethics Committee of the First Hospital of Jilin University, and written informed consent was obtained from all the babies' parents.

Study participants

Recruitment of subjects was from the obstetric unit of the First Hospital of Jilin University between July 2018 and October 2018, with 195 healthy term neonates enrolled prospectively. Neonatal assessment were performed by neonatal physicians, and maternal medical records were also reviewed. Eligibility criteria included gestational age between 37 and 42 weeks, birthweight ≥ 2500 g and < 4000 g, Apgar score at 1 and 5 minutes ≥ 7 , and the newborn was delivered by normal delivery or elective caesarean section. Infants were excluded from this study if they had congenital infections, chromosomal abnormalities or severe neonatal diseases diagnosed in the perinatal period, as well as infants of mothers suffering from hypertension, thyroid dysfunction or other severe illnesses. All newborns were routinely vaccinated with hepatitis B and Bacillus Calmette-Guérin (BCG) after birth, and given vitamin K1 1 mg intramuscularly.

Sample collection and laboratory procedure

Serum samples of newborns were taken within the first 3 days of life after obtaining informed consent from parents. A 4 ml sample of blood was collected into serum separation tubes in the morning from radial artery using disposable butterfly needle. The samples were transported to the laboratory and centrifuged at $1200 \times g$ for 10 min within 1 h after collection. Specimens that were icteric, hemolyzed, or lipemic were removed prior to analysis, and then the activity or concentration of 26 biochemical analytes were determined on the VITROS 5600 Integrated System (Ortho Clinical Diagnostics/ Johnson & Johnson, Raritan, NJ, USA) within 2 h after separation. The analytes were as follows: total carbon dioxide (CO_2), chloride (Cl), potassium (K), sodium (Na), calcium (Ca), iron (Fe), magnesium (Mg), phosphate (P), creatinine (Cr), urea (BUN), uric acid (UA), alanine aminotransferase (ALT), alkaline phosphatase (ALP), aspartate aminotransferase (AST), cholinesterase (CHE), creatine kinase (CK), creatine kinase MB form (CK-MB), gamma glutamyl transferase (GGT), lactate dehydrogenase (LDH), albumin (Alb), total protein (TP), transferrin iron-binding capacity (TIBC), cholesterol (TCHO), high-density lipoprotein-cholesterol (HDL-C) and triglycerides (TG). The low-density lipoprotein-cholesterol (LDL-C) result was calculated with the Friedewald equation for SI units [$\text{LDL-C} = \text{TC} - \text{HDL-C} - (\text{TG}/2.2)$]. All reagents and quality control products for the assays were purchased from Ortho Clinical Diagnostics/ Johnson & Johnson company. Qualified laboratory proficiency and staff training were described in the research conducted in the same period [3, 4]. Analytical procedures were performed according to the manufacturer's instructions and laboratory protocols, including regular maintenance, function checks, calibration, and internal quality control. Details of analytical methods and performance were listed in Supplemental Table 1.

Statistical analysis

The study population was divided into subgroups according to delivery mode, sex and day age. Histogram and Shapiro-Wilk test was performed to detect data distribution before statistical analysis, and non-normal data were logarithmically transformed. Demographic data were expressed as mean \pm standard deviation or number when appropriate. Mann-Whitney test or Wilcoxon rank sum test was used to compare numerical variables; Chi-square test was used for categorical variables.

For all the biochemical analytes, outliers were eliminated using the Dixon D/R ratio rule first, and data were expressed as median and interquartile range. Percentile curves were generated for all analytes using the LMS method. Three-level nested ANOVA was applied to evaluate sources of variation of RIs [14]. The magnitude of between-subgroup differences partitioned by the factors was computed as the standard deviation (SD) ratio (SDR), which corresponded to the SD from between-subgroup variation divided by SD between individuals. The variations of between-delivery-mode, between-sex and between-day-age components were expressed as SDR-dm, SDR-sex and SDR-age respectively. A ratio greater than 0.40 was considered as the criterion to partition RIs. According to CLSI EP28-A3c guidelines [10], the lower limits (LL) and upper limits (UL), referring to 2.5th percentiles and 97.5th percentiles, were derived by the nonparametric method or robust method after normalizing data with the Box-Cox method. The 90% confidence intervals (CIs) for reference limits were estimated on the bootstrap method through iterative resampling 100 times.

Multiple linear regression was used to investigate the associations between the analytes and individual characteristics, including GA, birthweight, delivery mode, sex and postnatal age. The significance of each variable was evaluated as standard partial regression coefficient (r_p), which corresponded to partial correlation coefficient and takes value between -1.0 and 1.0 . Based on an objective classification system in previous studies [15, 16], the practical correlation was characterized as slight for $0.20 \leq |r_p| < 0.30$, moderate for $0.30 \leq |r_p| < 0.50$ and strong for $0.50 \geq |r_p|$. All statistical analyses were performed using the *R* statistic software (version 3.6.0) and MedCalc Software (version 15.2). A bilateral *P*-value < 0.05 indicated statistical significance.

Results

Characteristics of study participants

A total of 195 term infants with postnatal age of 1-3 days fulfilled the inclusion criteria, and demographic information was shown in Table 1. Overall, both sexes were approximately equally represented in the study population, whereas the percentage of babies by caesarean section and on the second day was relatively large, accounting for 86.7% and 61.5% respectively. In brief, sex, birthweight, gestational age, maternal age and delivery mode did not show significant differences among the day-age groups.

Trends and RI establishment for common biochemical analytes

The division of RIs was codetermined by the scatter plot and 3-level nested ANOVA. Twenty-six common biochemical analytes had been measured during the first 3 days of life, and variation tendency depicted with the smoothed percentile curves for all analytes. Figure 1 and supplement Figures presented some interesting patterns, 1) BUN, Cr and UA significantly decreased over time, whereas Fe increased gradually within the 3 days; 2) there was an overall slight decrease in AST, CK, GGT, LDH, CK-MB, CO₂ and TIBC, while ALP showed a minor increase; 3) during the first day of life, Alb, TP, CHE and K decreased slightly, whereas Cl and Na increased slightly, and then they held steady in the following days; 4) TCHO, HDL-C, LDL-C and TG seemed to be steady during the first two days and began to rise on the 3rd day; 5) Ca appeared to decline within 2 days and then leveled off, yet Mg and P represented an opposite trend; 6) ALT remained steady all the time.

By adopting 0.40 as a significant effect size for SDR, the result of 3-level nested ANOVA showed that none of the analytes was affected by sex. The SDR values for delivery mode (SDR-dm) were higher than 0.40 for K, P and BUN. As shown in Figure 2, the upper limit (UL) and lower limit (LL) of subgroups divided by delivery mode were similar for BUN; instead, the BUN decreased significantly in a time-dependent manner. In this case, the RI of BUN was not partitioned by the mode of delivery. When the RIs of K and P were analyzed for vaginal births (VD) and cesarean-section (C/S) groups respectively, the UL of VD group (RI: 3.2-5.9 mmol/L) for K was higher than that of C/S group (RI: 3.3-5.4 mmol/L); the LL of VD group (RI: 1.56-2.79 mmol/L) for P was lower than that of C/S group (RI: 1.78-2.77 mmol/L). A small numerical distance was acceptable according to the clinician's recommendation and the level of clinical decision limit for both, the combined RIs of K and P were calculated to be 3.3-5.4 mmol/L and 1.70-2.75 mmol/L.

From the Table 2, the differences among day-age groups expressed as SDR-age were highlighted for 6 analytes. The medians and ULs of Fe, TCHO and TG gradually increased with age, however the changes of LLs were not very noticeable; the RIs for renal function tests (BUN, Cr and UA) went an apparent decline (Figure 1). Age-specific RI for each analyte was summarized in Table 4.

MRA to evaluate the correlations between individual characteristics and biochemical analytes

Multiple linear regression analysis (MRA) showed consistent results with nested ANOVA analysis (Table 3). For newborns, there was no actually significant association between sex and analyte concentrations during perinatal period. Instead, age was the main source of variation related to most analytes. In the regression models, some analytes showed slight associations with the delivery mode, birthweight and gestational age ($|r_p| \leq 0.3$). Infants born by vaginal delivery had a slightly high level of K and BUN and a slightly low level of P, compared to cesarean delivery. For BW-related changes, birthweight was positively correlated with P and TIBC and negatively correlated with ALT and ALP; for GA-related changes, GA showed positive correlations with ALT, CK and TP.

RIs comparison to other studies

The RIs of the 26 biochemical analytes showed differences among this study, reagent inserts and other published data (see Figure 3 and Supplemental Table 2). Comparing with the adult RIs provided by manufacturers, Cl, K, Na, and Mg did not show significant differences; lower ULs and LLs for CO₂, Ca, Fe, TIBC, BUN, Cr, UA, ALT, CHE, Alb and TP were observed in healthy neonates, however higher values for P, ALP, AST, GGT, CK, LDH could be seen. In particular, the analysis of CK-MB, lipid or lipoprotein was complex in adults, therefore the direct comparisons of the RIs were omitted.

In comparison with the RIs derived from other experimental designs, sex partitions were not emphasized in all the researches, but different age divisions and reference limits were able to be observed. These results showed that some differences in LDH were quite marked among the RIs listed for similar age group. ULs of Ca, Mg, Fe, TCHO and HDL-C are significantly lower than those from the experiment based on the cord blood in Korean [17]. The RIs of most analytes are similar to those from the study in Lanzhou, China [18], however the RIs of liver enzymes except CHE showed great differences compared with those from Zhengzhou, China [19].

Discussion

This study described the distribution and dynamic changes of biochemistry analytes from healthy term infants during the first 3 days of life in northeast China. There was heterogeneity in analytes with different biological sources of variations, however, day age showed a predominant correlation with the concentration for most analytes. Premature and term infants were prone to different diseases, and the demand for clinical evaluation and nursing intervention varied [20]. Therefore, the local RIs were derived for ensuring a high accuracy of clinical management.

Age-dependent RIs for 26 common biochemical analytes

In reviewing the literature, gestational age and birthweight were regarded as key factors for establishing RIs for preterm infants, but little difference of the two would be found in healthy term newborns. Although the guidance documents for establishing RIs only set the lowest limit for the number of subgroup participants, there was no single, specific definition for age intervals. If the age intervals were too wide, the rapid changes in a short time might be otherwise overlooked. Therefore, this study statistically depended upon SDR-dm, SDR-sex and SDR-age to analyze differences between subgroups, and then judged whether it was appropriate for clinical practice to partition RIs according to delivery mode, sex and postnatal age. Regarding the test results of SDR-dm and SDR-sex, RIs of all items did not require specific partitions (SDR <0.40) except for K, P and BUN. It was suggested that the combination of scatter plots and SDR values decide whether to partition RIs for test results comparing across different groups. This helps avoid exaggerated SDR for the actual narrow range of RIs or masked differences caused by the interaction between the factors.[2, 21] In effect, no sex difference for all analytes was detected, which was consistent with data in different regions [8, 17, 22].

However, there were some considerable differences in postnatal age. This might be due to the rapid physiological adaptation to extrauterine life for newborns. A possible explanation for this might be that perinatal data were mainly affected by maternal and neonatal physiological factors during the first days of life. The concentration of each analyte reflected the balance between the production, metabolism and clearance. First, some easy-to-understand mechanisms may cause short-term fluctuations of most analytes, such as proteins, electrolytes, lipids [23] and renal function markers [24], listed as follows: 1) the exchange of substances based on placental transfer between the fetus and mother ceased abruptly after delivery; 2) developmental and maturational changes occurred in the organs during the perinatal period, especially liver and kidney; 3) the intake of external nutrients directly and indirectly affected the concentration of metabolites. Furthermore, the stress response during labour can also have an impact. Enzyme levels evaluated in our study were consistent with Lackmann et al. who found that the cytoplasmic and mitochondrial enzymes presented similar activity curves, whereas the membrane-bound enzymes showed the opposite. The differences in enzymes were considered the result of minor cell damage caused by physiological hypoxia during labour [25]. However, the changes in our study may be not obvious enough because the release of enzymes was also affected by uterine contractions and physical stress through the birth canal, while the small number of natural delivery were included [26]. The regulation of substances by hormones was equally important after birth, which can be demonstrated by bone metabolism status in neonates. In addition to maternal vitamin D during pregnancy [27], serum Ca, Mg and P homeostasis was also regulated by foetal parathyroid hormone (PTH), calcitriol, calcitonin, calcium sensing receptor (CaSR) and fibroblast growth factor-23 (FGF-23), which affecting bone physiology, intestinal absorption and renal excretion [28].

Other variables that related to RIs of biochemical analytes

Generally the main considerations for sources of variations of each tests were sex, age and BMI in adults and children, besides races and regions. In the case of neonates, even more factors came into play, including maternal and infant health, delivery mode, gestational age and weight at birth. The comparison between the MLA model and the nested ANOVA method indicated that the results of delivery mode and sex were almost the same in present study, and postnatal age was the main influence on the concentration of most analytes. Birthweight and gestational age were widely used in evaluating neonatal clinical outcomes, while both had limited effects on a few items. A possible explanation for this might be that the premature or overdue babies and the low birthweight or fetal macrosomia were not included in this study.

In this study, there was poor negative correlation between birthweight and ALT and ALP, but positive correlation with P and TIBC although in term healthy newborns. Surprisingly, the small but consistent inverse associations of birthweight with ALT and ALP remained in late adolescent [29] and adult [30]. In fact, an elevated level of ALT within the normal range was proven to predict the hepatic steatosis of the metabolic syndrome [30, 31]. It may be assumed that birthweight is a proxy for exposures such as intrauterine nutrition or genetic factors that directly affect the liver. In addition, the opposite correlation of birthweight with ALP and P may reflect the inverse association between birthweight and bone strength. For another, the observed increasing levels in CK, TP and ALT with GA could be attributed to the rate of the babies' muscular mass, or to the development of their metabolism in advanced pregnancy [13, 32]. It is noteworthy that the expression of LC3-II and p62 in cord blood was associated with serum total protein in infants, perhaps suggesting that the autophagy reaction introduced by postnatal starvation played a crucial part in the maintenance of protein or amino acid metabolism during the perinatal period [33].

Comparison to Other Studies

RIIs in our study were compared with those of kit inserts and other studies, which covered different experimental types and designs, assay systems, characteristics of the studied population and specimen types. The manufacturer RIIs were less reliable in the evaluation of clinical applicability before use in Chinese laboratories, because they were generally based on Caucasian people and lacked data on children. The RIIs of Cl, K and Na were similar to those from reagent inserts and previous findings for children and adolescents [4]. Therefore, it seemed that the three electrolytes remained relatively stable in one's life. The differences of RIIs for Ca, Mg, P and lipids between this study and a study in Korea, to some extent, might be explained by the fact that umbilical cord contained maternal substances for fetal development. Compared to another two studies in other parts of China, there might be some factors that contributed to the differences between RIIs, including actual differences between cities, laboratories, analytical systems and statistical calculation methods reported in these studies. Moreover, Liu et al [19]. used an indirect method for determination of RIIs, which based on the stricter exclusion criteria and wider age intervals, and then little data about the rapid changes was presented during the first days.

As mentioned in the Caliper's report about transferred RIIs from Abbott to Ortho assays [34], Ca, CO₂, Mg and LDH did not meet transference or verification criteria. Except these, the RIIs of most corresponding analytes showed higher values, which might be attributed to racial and diet differences. This finding further support that laboratories should verify transferred RIIs for local population and analytical platform—that covers as many partitioned RIIs as possible—according to CLSI guidelines.

Limitations

Unfortunately, several limitations existed. First, being limited to recruiting healthy newborns, this study just determined the RIIs in newborns aged 0-3 days, and new data on infant period should be supplemented in the future. Second, once the RI was partitioned, the small sample size would be hard to assure a highly accurate RI with narrow confidence intervals. Finally, all the analytes should be evaluated the validity of routine use in clinics and further investigated in preterm infants. The percentile charts provided might be integrated into the hospital and laboratory information system to implement new strategies for result display.

Conclusion

This study used a priori method to investigate analyzer and reagent-appropriate RIIs in term infants, with clear inclusion and exclusion criteria, standardized sample processing procedures and novel data analysis methods. Moreover, the research of establishing RIIs on VITROS 5600 Integrated System was unique, and this study complemented those of earlier studies on children and adolescents. In summary, this study preliminarily established serum RIIs for 26 common biochemical analytes in healthy term infants, investigated their dynamic changes during the first 3 days of life and evaluated the associations of individual characteristics with these analytes. The results of this study indicated that day-age-appropriate RIIs should be considered for newborns in the field of perinatal-neonatal medicine. Continued efforts are needed to further assess the significance of these RIIs in routine clinical work and disease diagnosis.

Abbreviations

CO₂: total carbon dioxide; Cl: chloride; K: potassium; Na: sodium; Ca: calcium; Fe: iron; Mg: magnesium; P: phosphate; Cr: creatinine; BUN: urea; UA: uric acid; ALT: alanine aminotransferase; ALP: alkaline phosphatase; AST: aspartate aminotransferase; CHE: cholinesterase; CK: creatine kinase; CK-MB: creatine kinase MB form; GGT: gamma glutamyl transferase; LDH: lactate dehydrogenase; Alb: albumin; TP: total protein; TIBC: transferrin iron-binding capacity; HDL-C: high-density lipoprotein-cholesterol; TCHO: cholesterol; TG: triglycerides. LDL-C: low-density lipoprotein-cholesterol; LL: lower limit; MRA: multiple regression analysis; UL: upper limit; SDR: standard deviations ratio; r_p= partial correlation coefficient; RI: reference interval.

Declarations

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Authors' contributions

WKJ gathered and analysed the data, and wrote the first draft of the manuscript. ZXT was involved in gaining ethical approval, patient recruitment and experimental operation. ZQ and XJC conceived the study and contributed to protocol development. XJC revised the article. All authors have reviewed and edited the manuscript and approved the final version of the manuscript.

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Availability of data and materials

The datasets used and/or analysed during the current study are available from the corresponding author on reasonable request.

Ethical approval and consent to participate

All procedures performed in this study were in accordance with the ethical standards of the Ethics Committee of the First Hospital of Jilin University (reference number: 2016-306), as well as the 1964 Helsinki declaration and its subsequent amendments or comparable ethical standard. Written informed consent was obtained from all parents for the participants included in the study.

Consent for publication

N/A.

Competing interests

The authors declare that they have no competing interests.

Author details

¹Department of Laboratory Medicine, First Hospital of Jilin University, Changchun, China. ²Department of Pediatrics, First Hospital of Jilin University, Changchun, China

References

1. Ichihara K, Itoh Y, Lam CW, Poon PM, Kim JH, Kyono H, Chandrawening N, Muliaty D: Sources of variation of commonly measured serum analytes in 6 Asian cities and consideration of common reference intervals. *Clin Chem* 2008, 54(2):356-365.
2. Xia L, Chen M, Liu M, Tao Z, Li S, Wang L, Cheng X, Qin X, Han J, Li P *et al*: Nationwide Multicenter Reference Interval Study for 28 Common Biochemical Analytes in China. *Medicine (Baltimore)* 2016, 95(9):e2915.
3. Guo W, Zhou Q, Jia Y, Xu J: Age- and Sex-Specific Reference Intervals for Myocardial Enzyme Activity in Healthy Chinese Han Population Aged 1 approximately <18 years. *Biomed Res Int* 2019, 2019:2018598.
4. Zhu X, Wang K, Zhou Q, Guo W, Jia Y, Xu J: Age- and Sex-Specific Pediatric Reference Intervals of Serum Electrolytes in Jilin Province of China Using the A Priori Approach. *American journal of clinical pathology* 2020.
5. Guo W, Zhou Q, Jia Y, Xu J: Division of Myocardial Enzyme Reference Intervals in Population Aged 1 to <18 Years Old Based on Fisher's Optimal Segmentation Method. *Comput Math Methods Med* 2020, 2020:2013148.
6. Li X, Wang D, Yang C, Zhou Q, Zhuoga SL, Wang LQ, Yao HX, Zhang Q, Ai Q, Yang CX *et al*: Establishment of age- and gender-specific pediatric reference intervals for liver function tests in healthy Han children. *World J Pediatr* 2018, 14(2):151-159.
7. Ianni B, McDaniel H, Savilo E, Wade C, Micetic B, Johnson S, Gerkin R: Defining Normal Healthy Term Newborn Automated Hematologic Reference Intervals at 24 Hours of Life. *Arch Pathol Lab Med* 2020.
8. Schmidt BM, Tameris M, Geldenhuys H, Luabeya A, Bunyasi E, Hawkrigde T, McClain JB, Mahomed H, Scriba TJ, McShane H *et al*: Comparison of haematology and biochemistry parameters in healthy South African infants with laboratory reference intervals. *Trop Med Int Health* 2018, 23(1):63-68.
9. Moyer-Mileur LJ: Anthropometric and laboratory assessment of very low birth weight infants: the most helpful measurements and why. *Semin Perinatol* 2007, 31(2):96-103.
10. Institute CaLS: Defining, Establishing, and Verifying Reference Intervals in the Clinical Laboratory; Approved Guideline—3rd ed. CLSI document C28-A3c. Wayne, PA: Institute CaLS; 2010.
11. Dokos C, Tsakalidis C, Manaridou K, Karayianni P, Kyrkos I, Roussos I: Clinical-laboratory findings of bone metabolism in healthy premature and full-term neonates: preliminary results. *Clin Cases Miner Bone Metab* 2017, 14(2):167-172.
12. Donega S, Oba J, Maranhao RC: [Concentration of serum lipids and apolipoprotein B in newborns]. *Arq Bras Cardiol* 2006, 86(6):419-424.
13. Elizabeth KE, Krishnan V, Zachariah P: Auxologic, biochemical and clinical (ABC) profile of low birth weight babies- a 2-year prospective study. *J Trop Pediatr* 2007, 53(6):374-382.

14. Ichihara K, Boyd JC, Intervals ICoR, Decision L: An appraisal of statistical procedures used in derivation of reference intervals. *Clinical chemistry and laboratory medicine* 2010, 48(11):1537-1551.
15. Borai A, Ichihara K, Masaud A, Tamimi W, Bahijri S, Armbuster D, Kawano R, Baarmah Z, Joatar F, Almohammadi M: Establishment of reference intervals for immunoassay analytes of adult population in Saudi Arabia. *Clinical chemistry and laboratory medicine* 2020, 58(8):1302-1313.
16. Evgina S, Ichihara K, Ruzhanskaya A, Skibo I, Vybornova N, Vasiliev A, Kimura S, Butlitski D, Volkova E, Vilenskaya E *et al*: Establishing reference intervals for major biochemical analytes for the Russian population: a research conducted as a part of the IFCC global study on reference values. *Clin Biochem* 2020, 81:47-58.
17. Choi SJ, Lee S, Lee B, Jang JY, Cho J, Uh Y: Comparison of neonatal reference intervals for 23 biochemical analytes in the cord blood-A single center study in South Korea. *Turk J Pediatr* 2019, 61(3):337-344.
18. Zhao C, Wang H, Zhang C, He L: Research on reference values of 26 items of serum biochemical parameters in Lanzhou healthy neonates. *Int J Lab Med* 2007, 28(3):211-213.
19. Liu J, Dai Y, Lee Y, Yuan E, Wang Q, Wang L, Su Y: Pediatric reference intervals of liver and renal function tests from birth to adolescence in Chinese children as performed on the Olympus AU5400. *Clin Chim Acta* 2019, 490:142-146.
20. Cheng YW, Kaimal AJ, Bruckner TA, Halloran DR, Caughey AB: Perinatal morbidity associated with late preterm deliveries compared with deliveries between 37 and 40 weeks of gestation. *BJOG* 2011, 118(12):1446-1454.
21. Ichihara K, Yomamoto Y, Hotta T, Hosogaya S, Miyachi H, Itoh Y, Ishibashi M, Kang D, Committee on Common Reference Intervals JSOCC: Collaborative derivation of reference intervals for major clinical laboratory tests in Japan. *Ann Clin Biochem* 2016, 53(Pt 3):347-356.
22. Zierk J, Arzideh F, Rechenauer T, Haeckel R, Rascher W, Metzler M, Rauh M: Age- and Sex-Specific Dynamics in 22 Hematologic and Biochemical Analytes from Birth to Adolescence. *Clinical Chemistry* 2015, 61(7):964-973.
23. Yip PM, Chan MK, Nelken J, Lepage N, Brotea G, Adeli K: Pediatric reference intervals for lipids and apolipoproteins on the VITROS 5,1 FS Chemistry System. *Clin Biochem* 2006, 39(10):978-983.
24. Novo AC, Sadeck Ldos S, Okay TS, Leone CR: Longitudinal study of Cystatin C in healthy term newborns. *Clinics (Sao Paulo)* 2011, 66(2):217-220.
25. Lackmann GM: Reference values for selected enzyme activities in serum from healthy human neonates. *Clin Biochem* 1996, 29(6):599-602.
26. Nakajima J, Tsutsumi N, Nara S, Ishii H, Suganami Y, Sunohara D, Kawashima H: Correlations of Enzyme Levels at Birth in Stressed Neonates with Short-Term Outcomes. *Fetal Pediatr Pathol* 2018, 37(3):157-165.
27. Larque E, Morales E, Leis R, Blanco-Carnero JE: Maternal and Foetal Health Implications of Vitamin D Status during Pregnancy. *Ann Nutr Metab* 2018, 72(3):179-192.
28. Sethi A, Priyadarshi M, Agarwal R: Mineral and bone physiology in the foetus, preterm and full-term neonates. *Semin Fetal Neonatal Med* 2020, 25(1):101076.
29. Liu JX, Au Yeung SL, Kwok MK, Leung JYY, Lin SL, Hui LL, Leung GM, Schooling CM: Birth weight, gestational age and late adolescent liver function using twin status as instrumental variable in a Hong Kong Chinese birth cohort: "Children of 1997". *Prev Med* 2018, 111:190-197.
30. Fraser A, Ebrahim S, Smith GD, Lawlor DA: The associations between birthweight and adult markers of liver damage and function. *Paediatr Perinat Epidemiol* 2008, 22(1):12-21.
31. Chen S, Guo X, Yu S, Zhou Y, Li Z, Sun Y: Metabolic Syndrome and Serum Liver Enzymes in the General Chinese Population. *Int J Environ Res Public Health* 2016, 13(2):223.
32. Bellieni CV, Tomasini B, Bracciali C, Buonocore G: Normal values of creatine kinase and of MB-creatine kinase at birth in healthy babies. *Minerva Pediatr* 2017.
33. Sekiguchi K, Miyahara H, Inoue M, Maeda T, Ihara K: The autophagy reaction in the human umbilical cord: a potential marker for estimating fetal nutrition and neonatal growth. *J Matern Fetal Neonatal Med* 2020:1-5.
34. Higgins V, Truong D, Woroch A, Chan MK, Tahmasebi H, Adeli K: CLSI-based transference and verification of CALIPER pediatric reference intervals for 29 Ortho VITROS 5600 chemistry assays. *Clin Biochem* 2018, 53:93-103.

Tables

Table 1 Participant demographics.

Characteristics	Total	Day 1	Day 2	Day 3	Pvalue
	n=195	n=21	n=120	n=54	
Gestational age (weeks)	38.7±1.1	39.0±0.86	38.6±1.1	38.7±1.1	0.17
Birthweight (kg)	3.37±0.43	3.58±0.44	3.33±0.45	3.37±0.39	0.13
Boys/girls	104/91	12/9	62/58	30/24	0.83
Vaginal delivery/caesarean section	26/169	5/16	14/106	7/47	0.31

Data represented as mean ± standard deviation or number when appropriate.

Table 2 Result of 3-level Nested ANOVA and summary of statistical description for 28 biochemical analytes according to day age.

Analytes	3-level Nested ANOVA			Day1		Day 2		Day 3		Total	
	SDR-dm	SDR-sex	SDR-age	median	IQR	median	IQR	median	IQR	median	IQR
CO ₂	0.221	0.000	0.100	24	22-25	21	20-23	21	19-25	22	20-24
Cl	0.222	0.000	0.319	101	95-104	103	100-104	102	97-105	102	98-104
K	0.431	0.000	0.231	3.9	3.6-4.3	4.0	3.7-4.3	4.2	3.8-4.5	4.0	3.7-4.4
Na	0.000	0.000	0.021	135	133-136	135	133-137	135	133-136	135	133-137
Ca	0.285	0.000	0.000	1.97	1.88-2.01	1.92	1.83-2.03	1.96	1.81-2.04	1.95	1.83-2.03
Mg	0.264	0.000	0.323	0.77	0.76-0.80	0.80	0.76-0.84	0.80	0.78-0.87	0.80	0.76-0.84
P	0.673	0.139	0.091	2.11	1.85-2.36	2.28	2.10-2.43	2.18	2.07-2.39	2.25	2.07-2.42
Fe	0.151	0.000	0.411	8.4	7.1-9.4	9.4	8.3-11.0	10.6	8.7-12.8	9.6	8.3-11.3
TIBC	0.278	0.000	0.000	42.0	40.0-44.4	40.4	37.8-43.0	39.9	37.2-43.7	40.5	37.7-43.3
BUN	0.580	0.000	0.519	2.53	1.75-3.14	1.65	1.14-2.25	1.08	0.78-1.63	1.57	1.05-2.15
Cr	0.121	0.000	0.670	61	52-67	52	48-60	46	42-52	51	45-60
UA	0.000	0.000	0.675	278	183-308	209	164-244	146	130-189	186	145-241
ALT	0.164	0.000	0.000	30	27-32	30	22-35	31	25-34	30	24-34
ALP	0.096	0.228	0.000	143	129-155	150	123-167	156	141-172	151	127-168
AST	0.000	0.272	0.180	56	52-83	52	44-63	52	42-63	53	44-63
CHE	0.325	0.000	0.167	4326	4060-4983	4266	3871-4696	4272	4069-4458	4279	3897-4655
GGT	0.000	0.000	0.000	133	91-169	129	86-169	113	88-142	122	86-169
CK	0.000	0.213	0.251	412	297-470	318	233-414	238	157-382	304	203-415
CK-MB	0.066	0.116	0.000	22	18-26	24	18-30	22	18-29	23	18-30
LDH	0.000	0.146	0.246	1247	972-1330	1149	1022-1270	1039	928-1230	1120	985-1285
Alb	0.263	0.215	0.116	33.7	30.8-36.3	32.7	30.8-34.3	32.3	30.3-34.7	32.7	30.7-34.7
TP	0.314	0.190	0.000	47.1	45.6-50.8	48.0	45.5-52.0	49.6	45.6-52.2	48.2	45.5-52.1
HDL-C	0.117	0.000	0.315	0.62	0.55-0.80	0.67	0.58-0.81	0.71	0.59-0.83	0.68	0.58-0.82
LDL-C	0.000	0.000	0.319	0.56	0.51-0.80	0.57	0.45-0.80	0.90	0.56-1.15	0.63	0.48-0.91
TCHO	0.125	0.000	0.618	1.57	1.29-1.85	1.60	1.38-1.92	2.08	1.65-2.39	1.73	1.39-2.05
TG	0.000	0.000	0.517	0.64	0.56-0.77	0.70	0.58-0.86	1.02	0.77-1.26	0.74	0.58-0.95

SDR \geq 0.4 was used as a criterion for partition of reference values (highlighted in grey).

SDR, standard deviation ratio; SDR-dm, SDR for between-delivery-mode differences; SDR-sex, SDR for between-sex differences; SDR-age, SDR for between-day-age differences; IQR, interquartile range.

Table 3 Multiple linear regression of individual characteristics on the biochemical analytes.

Analytes	Delivery mode	Sex	Postnatal age	Birth weight	Gestational age	Analytes	Delivery mode	Sex	Postnatal age	Birth weight	Gestational age
CO ₂	0.099	0.079	-0.118	0.114	0.014	ALP	-0.051	0.119	0.099	-0.200	-0.085
Cl	-0.053	0.046	0.057	-0.171	-0.108	AST	0.079	0.141	-0.179	-0.137	0.094
K	-0.200	-0.088	0.133	-0.059	-0.001	CHE	-0.133	0.007	-0.073	-0.006	-0.198
Na	-0.050	0.078	-0.136	-0.060	-0.043	GGT	-0.072	-0.029	-0.095	-0.148	-0.139
Ca	-0.171	0.001	-0.028	-0.029	0.137	CK	-0.083	0.131	-0.246	-0.042	0.301
Mg	0.102	-0.072	0.192	-0.196	0.103	CKMB	0.083	0.133	-0.063	-0.067	0.068
P	0.262	-0.118	0.014	0.231	-0.195	LDH	0.040	0.080	-0.195	-0.014	0.028
Fe	-0.108	0.090	0.297	-0.119	0.133	Alb	-0.160	-0.161	-0.083	0.094	0.042
TIBC	0.074	0.025	-0.085	0.232	-0.048	TP	-0.130	-0.114	0.022	-0.124	0.260
BUN	-0.231	0.007	-0.388	-0.123	0.050	HDL-C	-0.120	-0.142	0.265	0.157	-0.144
Cr	-0.075	0.092	-0.445	-0.130	-0.121	LDL-C	-0.171	-0.134	0.408	0.142	-0.074
UA	-0.001	0.059	-0.457	-0.091	-0.004	TCHO	-0.143	-0.110	0.147	0.002	-0.079
ALT	-0.077	-0.104	-0.021	-0.304	0.240	TG	-0.051	-0.030	0.378	-0.020	0.178

Listed values are standardized partial regression coefficients (r_p). $|r_p|$ values that exceed 0.2 was shown in three graded black background colors: light $0.2 \leq |r_p| < 0.3$, moderate $0.3 \leq |r_p| \leq 0.5$, dark $|r_p| \geq 0.5$ respectively.

Table 4 Age-dependent reference intervals for 28 biochemical analytes in newborns.

Analyte	Age	Lower Limit (90% CI)	Upper Limit (90% CI)	Analyte	Age	Lower Limit (90% CI)	Upper Limit (90% CI)
Chemistry				Enzymes			
CO ₂ , mmol/l	1-3 d	16 (16-17)	28 (28-29)	ALT, U/L	1-3 d	6	44 (41-46)
Cl, mmol/l	1-3 d	92 (91-93)	108 (108-109)	ALP, U/L	1-3 d	91 (83-103)	229 (214-240)
K, mmol/l	1-3 d	3.3 (3.2-3.4)	5.4 (5.1-5.6)	AST, U/L	1-3 d	29 (29-33)	101 (92- 115)
Na, mmol/l	1-3 d	131 (130-131)	140 (140-141)	CHE, U/L	1-3 d	3299 (3272-3397)	6491 (5838-6736)
Ca, mmol/l	1-3 d	1.53 (1.42-1.61)	2.15 (2.12-2.16)	GGT, U/L	1-3 d	42 (39-54)	302 (280-316)
Mg, mmol/l	1-3 d	0.68 (0.65-0.70)	0.94 (0.91-0.96)	CK, U/L	1-3 d	95 (89-109)	715 (566-773)
P, mmol/l	1-3 d	1.70 (1.64-1.78)	2.75 (2.68-2.84)	CK-MB, U/L	1-3 d	12 (11-13)	40 (39-41)
Fe, µmol/L	1 d	5.2 (4.6- 5.9)	14.6 (12.1-17.2)	LDH, U/L	1-3 d	651 (570-720)	1577 (1524-1620)
	2 d	6.8 (6.6-7.0)	15.9 (14.6-17.4)	Proteins			
	3 d	6.4 (5.9- 7.1)	18.1 (16.3-19.8)	Alb, g/L	1-3 d	28 (27-28)	40 (39-43)
TIBC, µmol/L	1-3 d	32.9 (31.9-35.1)	52.4 (50.5-55.5)	TP, g/L	1-3 d	41 (40-41)	59 (57-61)
BUN, mmol/l	1 d	0.71*	5.72 (4.58-6.87)	Lipids/Lipoproteins			
	2 d	0.71*	3.77 (3.58-4.23)	HDL-C, mmol/l	1-3 d	0.43 (0.39-0.46)	1.13 (1.04-1.19)
	3 d	0.71*	3.55 (2.56-5.04)	LDL-C, mmol/l	1-3 d	0.20 (0.17-0.22)	1.48 (1.26-1.76)
Cr, µmol/L	1 d	41 (35-47)	82 (76-87)	TG, mmol/l	1-2 d	0.38 (0.33-0.43)	1.25 (1.12-1.55)
	2 d	38 (37-40)	76 (72-80)		3 d	0.32 (0.22-0.44)	1.84 (1.66-2.01)
	3 d	31 (29-34)	65 (60-69)	TCHO, mmol/l	1-2 d	1.29*	2.57 (2.46-3.11)
UA, µmol/L	1 d	99 (68-151)	454 (388-512)		3 d	1.29*	3.16 (2.96-3.37)
	2 d	97 (87-110)	335 (318-351)				
	3 d	78 (69-88)	271 (236-300)				

*Lowest detection limit. Actual values could be lower.

Figures

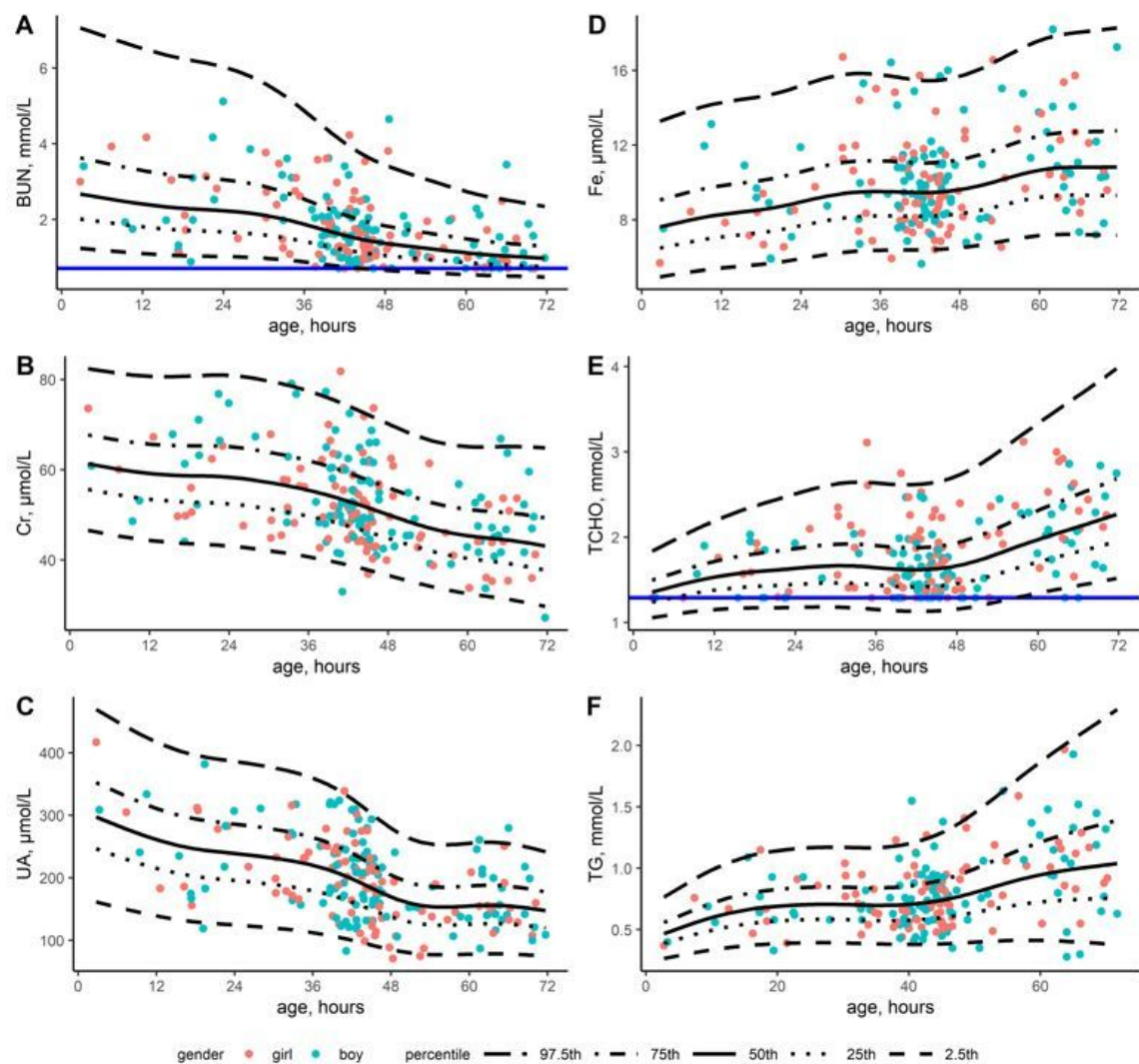


Figure 1

Percentile charts for BUN, Cr, UA, Fe, TCHO and TG. Horizontal blue lines shown in the graphs for BUN and TCHO represent lowest detection limit.

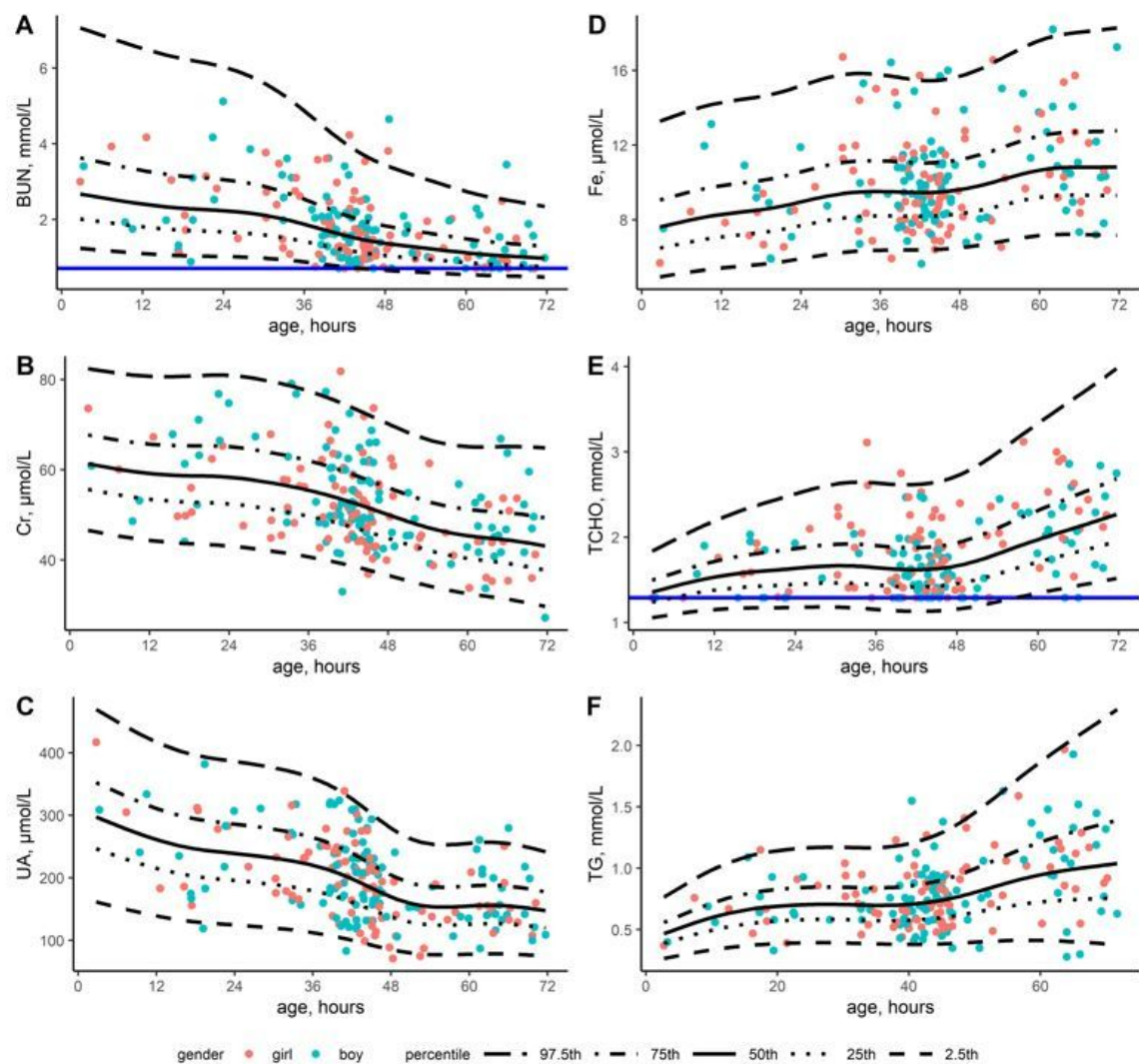


Figure 1

Percentile charts for BUN, Cr, UA, Fe, TCHO and TG. Horizontal blue lines shown in the graphs for BUN and TCHO represent lowest detection limit.

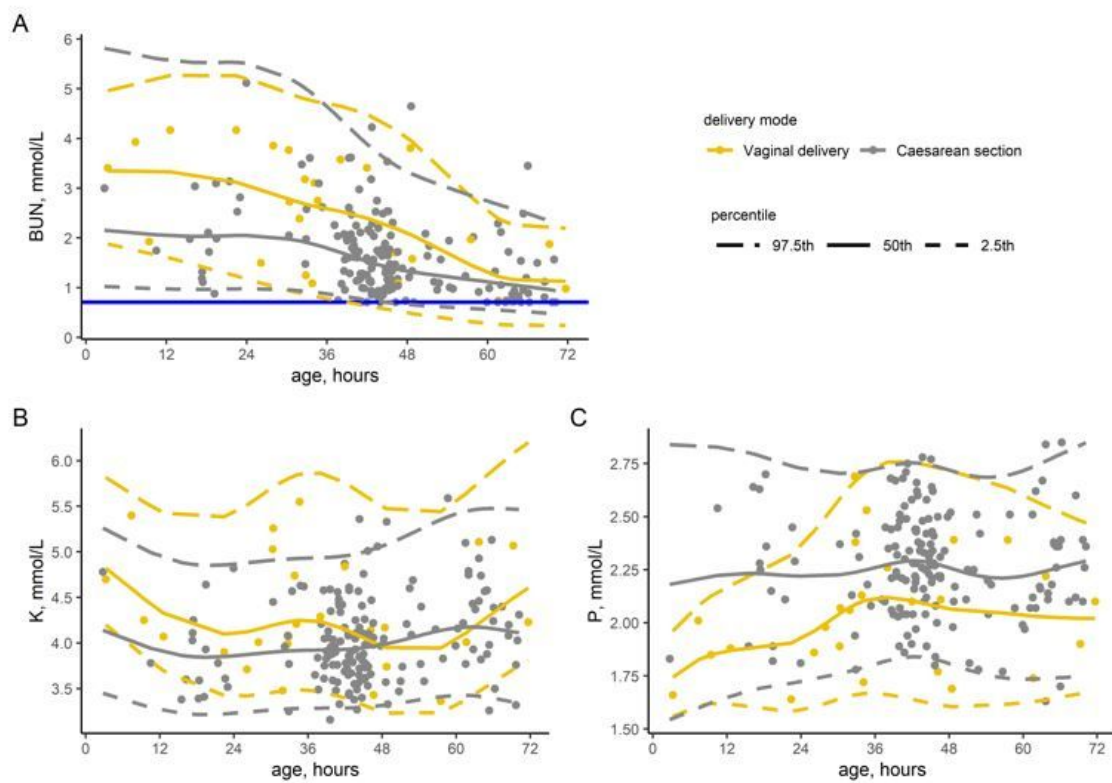


Figure 2

Percentile charts for K, P and BUN between subgroups in different delivery modes. Horizontal blue lines shown in the graphs for BUN represent lowest detection limit.

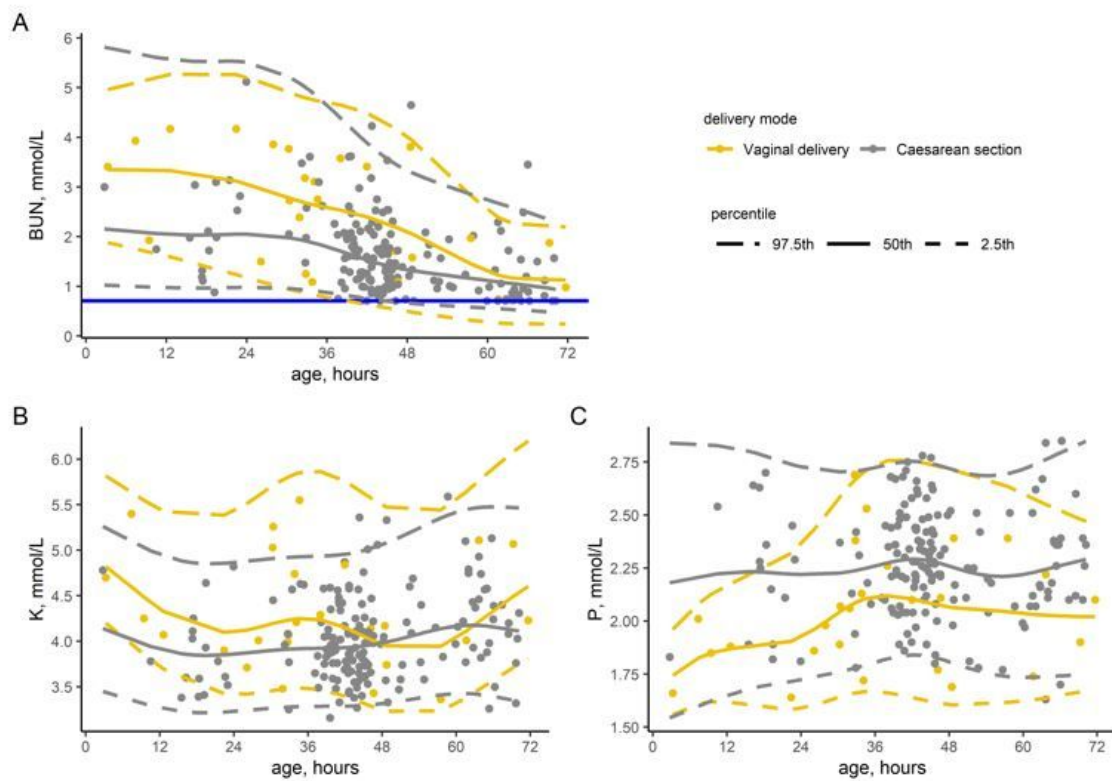


Figure 2

Percentile charts for K, P and BUN between subgroups in different delivery modes. Horizontal blue lines shown in the graphs for BUN represent lowest detection limit.

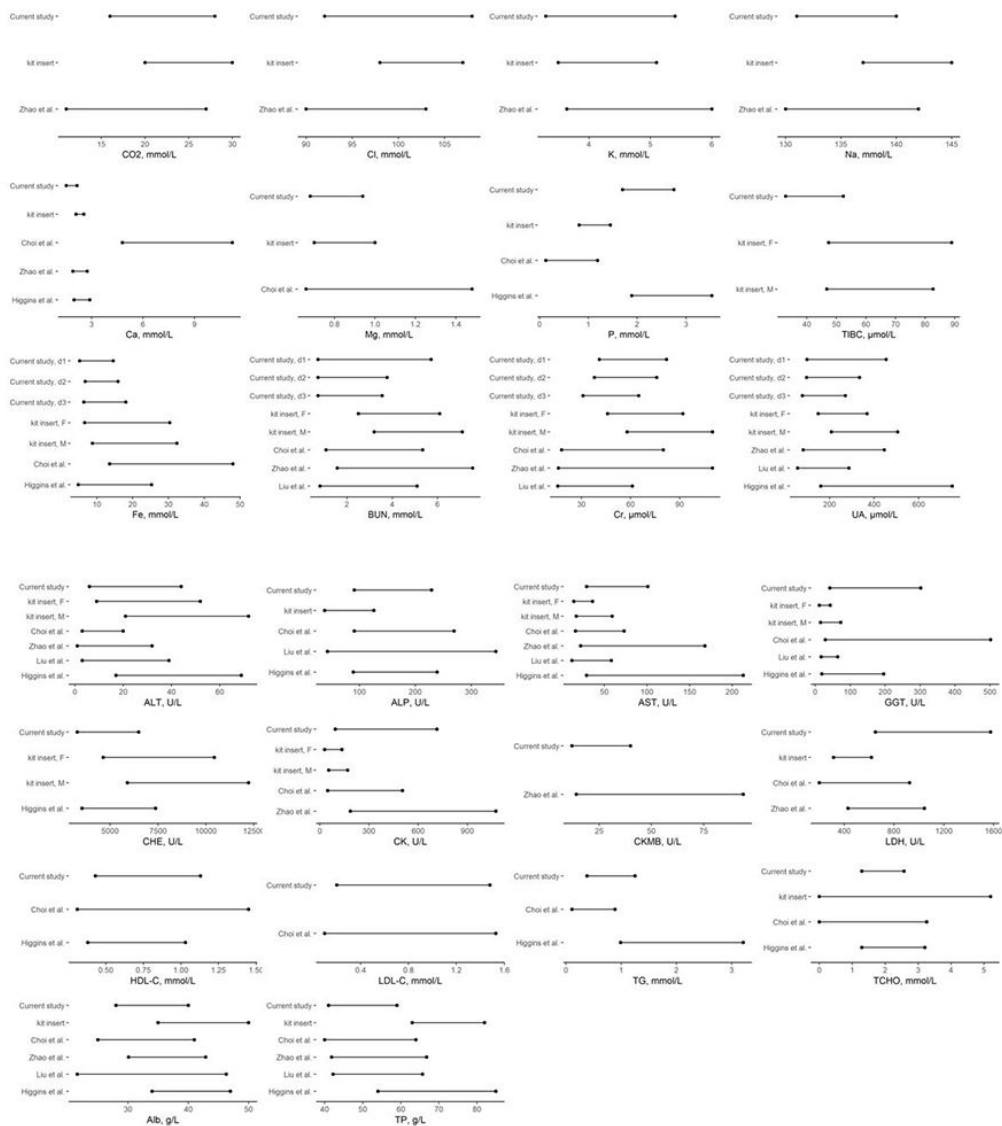


Figure 3

a Biochemical reference intervals in comparison with other studies. b Biochemical reference intervals compared with other studies.

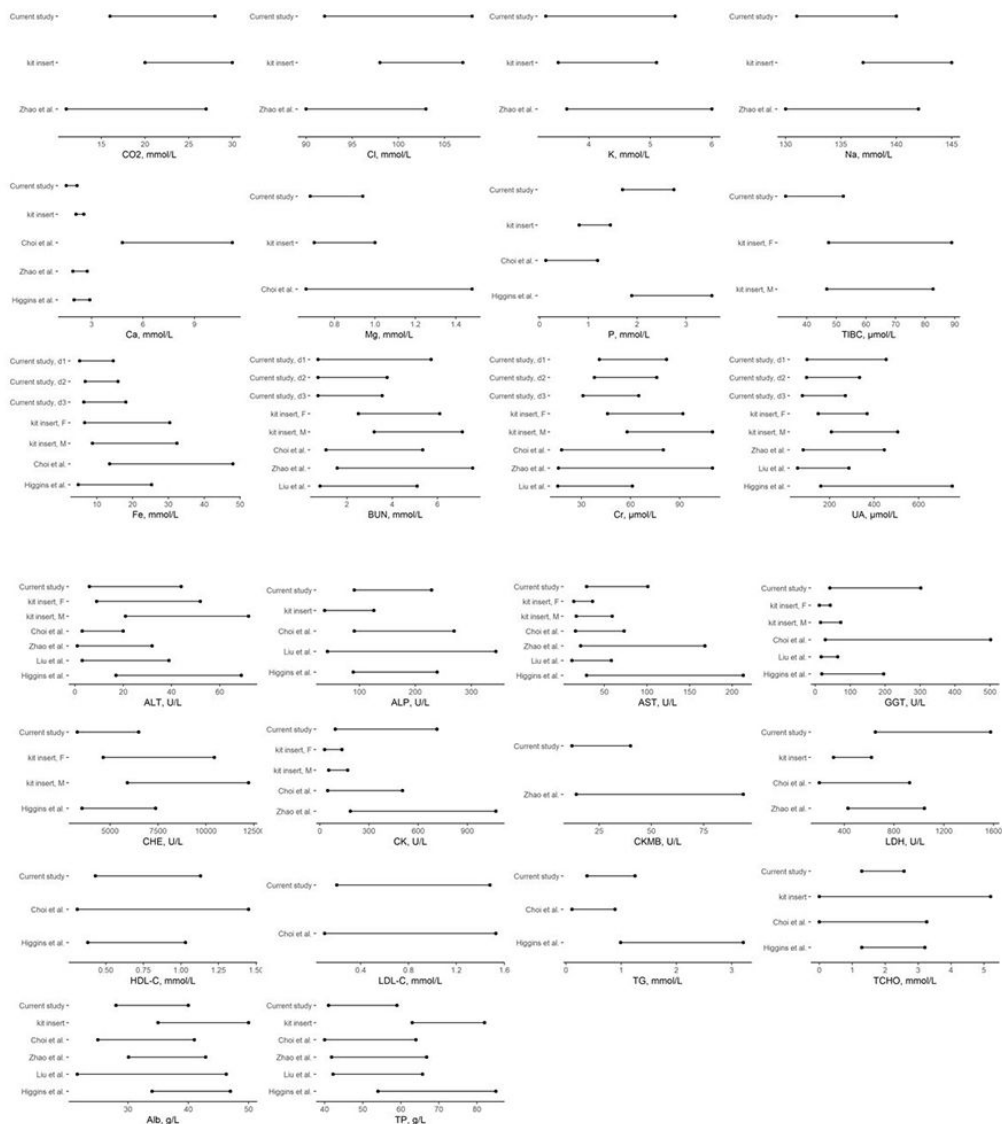


Figure 3

a Biochemical reference intervals in comparison with other studies. b Biochemical reference intervals compared with other studies.

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