

Sampling Strategies to Evaluate the Prognostic Value of a New Biomarker on a Time-to-Event End-Point

Francesca Graziano

University of Milan–Bicocca: Università degli Studi di Milano-Bicocca

Maria Grazia Valsecchi

University of Milano-Bicocca

Paola Rebora (✉ paola.rebora@unimib.it)

University of Milano-Bicocca <https://orcid.org/0000-0003-0606-5852>

Research article

Keywords: case-control design, cohort studies, power, two-phase sampling, weighted Cox model

Posted Date: November 9th, 2020

DOI: <https://doi.org/10.21203/rs.3.rs-102892/v1>

License:  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

Version of Record: A version of this preprint was published at BMC Medical Research Methodology on April 30th, 2021. See the published version at <https://doi.org/10.1186/s12874-021-01283-0>.

1 **TITLE: Sampling strategies to evaluate the prognostic value of a new biomarker on a**
2 **time-to-event end-point**

3 **RUNNING TITLE: Two phase sampling strategies**

4 Francesca Graziano¹, Maria Grazia Valsecchi¹ and Paola Rebora^{1*}

5 **Author affiliations:** ¹ BICOCCA BIOINFORMATICS BIOSTATISTICS AND
6 BIOIMAGING CENTRE-B4, School of Medicine and Surgery, University of Milano -
7 Bicocca, Via Cadore 48, 20900 Monza, Italy

8

9 *Correspondence:

10 paola.rebora@unimib.it, Tel. +39 02 6448 8165

11 BICOCCA BIOINFORMATICS BIOSTATISTICS AND BIOIMAGING CENTRE-B4,
12 School of Medicine and Surgery, University of Milano - Bicocca, Via Cadore 48, 20900
13 Monza, Italy

14

15

16

17

18

19

20

21

22

23

24

25

26 **ABSTRACT**

27 **Background:** The availability of large epidemiological or clinical data storing biological
28 samples allow to study the prognostic value of novel biomarkers, but efficient designs are
29 needed to select a subsample on which to measure them for parsimony and economical
30 reasons. Two-phase stratified sampling is a flexible approach to perform such sub-sampling,
31 but literature on stratification variables to be used in the sampling and power evaluation is
32 lacking especially for survival data.

33 **Methods:** We compared the performance of different sampling designs to assess the
34 prognostic value of a new biomarker on a time-to-event end-point, applying a two-phase Cox
35 model weighted by the inverse of the empirical inclusion probability.

36 **Results:** Our simulation results suggest that case-control stratified (or post stratified) by a
37 surrogate variable of the marker can yield higher performances than simple random,
38 probability proportional to size and case-control sampling. In the presence of high censoring
39 rate, results showed an advantage of nested case-control and counter-matching designs in
40 term of design effect, but their constrain in using a fixed ratio between cases and controls
41 might be disadvantageous. On real data on childhood acute lymphoblastic leukemia, we also
42 found that optimal sampling using pilot data is greatly efficient.

43 **Conclusions:** Our study suggests that, in our sample, case-control stratified by surrogate and
44 nested case-control yield estimates and power comparable to estimates obtained in the full
45 cohort while strongly decreasing the number of patients required. We recommend to plan the
46 sample size and using sampling designs for exploration of novel biomarker in clinical cohort
47 data.

48 **Keywords:** case-control design, cohort studies, power, two-phase sampling, weighted Cox
49 model

50

51 **1. BACKGROUND**

52 In the past decades, there has been a growing number of epidemiological (1-3) and
53 longitudinal studies storing biological samples (4) to allow retrospective evaluation of new
54 research questions, such as evaluating the prognostic value of new biomarkers. This approach
55 is convenient, significantly reducing the time of the study. However, the analysis of novel
56 biomarkers could be expensive. Sub-sampling strategies result in considerable cost savings
57 and parsimonious use of biological specimens, by restricting data extraction to an informative
58 subgroup of the original sample. Instead of choosing the subgroup at random, it may be
59 carefully sampled to obtain unbiased and more precise results (5, 6).

60 Two-phase sampling is a general approach to perform such sub-sampling, including case-
61 control and case-cohort designs (7, 8). This approach considers the entire cohort as the first-
62 phase sample from the population of interest. In the second phase, subsamples are drawn from
63 the cohort to measure additional information, such as new biomarkers of interest (9). An
64 optimal sampling strategy was proposed for stratified two-stage studies with binary outcome,
65 however it needs the availability of pilot data on the biomarker of interest that are not always
66 available. Moreover, there is no literature on the choice of variables to be used as strata and
67 no extension for time-to-event outcomes.

68 The sample size of retrospective studies is often planned considering only budget constrain
69 without a proper evaluation of the statistical power (10), also due to the lack of methodologies
70 to evaluate power in this setting. Cai and Zeng (11) focused on the power in case-cohort
71 design without any stratification; Haneuse et al. (12, 13) focused on binary outcomes, but a
72 general strategy for power evaluation is missing for survival data.

73 In this study we compared different approaches of sampling designs in the two-phase setting
74 to assess the prognostic value of a new biomarker on a time-to-event end-point and provide a
75 tool to estimate power by simulations. In particular, we focused on the sampling design of the

76 sub-cohort on which to measure the new biomarker. In this context, we explored the
77 contribution of different types of stratification variables (e.g. surrogate, risk factor,
78 confounder...) and of different sampling designs on power and design effect. We performed
79 a power evaluation varying the sub-cohort sample size and using real data from a study
80 carried out using data from a randomized trial in childhood acute lymphoblastic leukemia
81 (ALL). Briefly, this study was performed to evaluate the role of different genetic
82 polymorphisms on treatment failure due to relapse (14, 15). Clinical data and other
83 information were available for the whole trial cohort and biological samples were stored at
84 diagnosis. The genetic polymorphisms were retrospectively evaluated on these specimens
85 using a two-phase design.

86 2. METHODS

87 2.1 Notation settings

88 Due to a survival analysis framework, commonly survival notation has been used. Let T_i be
89 the failure time and C_i the censoring time of subject i ($i = 1 \dots N$) in a cohort (phase I) of size
90 N followed-up to time τ . T_i and C_i are assumed to be independent, $T \perp C$. Define $Z =$
91 $\min(T, C)$ and $\Delta = I(T \leq C)$. Let $h_i(t)$ be the hazard rate for the i th individual. The hazard
92 function, modelled using the Cox proportional hazards model, is equal to $h_i(t) =$
93 $h_0(t)\exp(\beta X_i)$ where $h_0(t)$ is the baseline hazard, X_i the vector of the explanatory
94 variables for individual i and β 's the corresponding regression coefficients. The classical
95 approach for estimating β is to maximize the partial likelihood (16). Suppose that information
96 on one variable of interest, i.e. X_{BM} , is available only for a subset $n < N$ of subjects drawn
97 from the phase I data and let ξ_i indicate whether subject i is selected into this sample. We will
98 refer to the $n = \sum_{i=1}^N \xi_i$ subjects as the phase II sample. Let $\pi_i = P(\xi_i = 1 | X_i, \Delta_i, Z_i)$ being
99 the inclusion probability of subject i for the phase II sample, conditional on being selected at
100 the first phase. In a random sample this probability is equal for every subject ($\pi = n/N$).

101 However, in a stratified sampling this probability is common for all subjects in the same
102 stratum and differs between strata. In particular, it is usually higher for the more informative
103 strata (e.g. strata including subjects with the event of interest as in case-control studies).

104

105 **2.2 Simulation context**

106 **First phase sample**

107 To mimic a realistic context, we hypothesized a cohort of subjects of size N (i.e. clinical trial
108 cohort, register, clinical cohort) followed up to τ , in which we aim to evaluate the prognostic
109 value of a new biomarker (X_{BM}) on a time-to-event endpoint (T) in the presence of a possible
110 confounder (X_{Conf}), a risk factor ($X_{Risk\ Fact}$) and a possible auxiliary/surrogate variable
111 (X_{Surr}) of the marker of interest. To describe and illustrate relationships between these, a
112 Directed Acyclic Graph (DAG) was displayed in **Figure 1**. In particular, we assumed the
113 confounder to have an impact on both the biomarker and the event of interest, the risk factor
114 to be associated only with the event of interest, and the surrogate to be associated only with
115 the biomarker. We believe that these variables might well represent the majority of variables
116 usually available in practice, even though in a simplified setting for simulation.

117 *Insert Figure 1*

118 **Second Phase Sample**

119 We assumed that the risk factor, the confounder and the surrogate are known for all subjects
120 in the first phase (N), while the biomarker (X_{BM}) is measured only on the subset of n
121 individuals (second phase sample).

122 To sample the subset a stratified two-phase sampling approach was used. Strata for the first
123 phase sample were defined using the following variables: event, event and risk factor, event
124 and confounder, event and surrogate.

125 By note, in this work, we consider only sampling done at the end of the follow-up (τ).

126 Subjects who developed the event during the follow-up are defined as cases and subjects
127 event-free at τ as controls.

128 The sample size of the second phase is fixed (n), but the sampling probabilities depend on
129 different designs, as described below:

130 (i) **Simple Random Sample (SRS)**: all possible subsamples have an equal probability to be
131 chosen.

132 (ii) **Probability Proportional to Size (PPS)**: The inclusion probability for cases and controls
133 is constant ($\pi_i=n/N$) applied within each stratum without replacement (17). Thus, the size
134 for each stratum is given by the total size of the stratum in the original cohort multiplied by
135 n/N .

136 (iii) **Case-Control (CC)** is performed by separately sampling cases and controls (18). As we
137 aimed to compare different sampling strategies with a fixed sample size, we did not
138 necessarily select all cases from the full cohort as often done. We fixed a total sample size
139 (n) and an equal number of cases ($n/2$) and controls ($n/2$) were selected. We also
140 considered stratified CC by using the variables available in phase I (see **Figure 1**):
141 separated simple random sampling was performed in each stratum. A balance design was
142 considered (19).

143 (iv) **Nested case-control (NCC)** can be considered as a particular case of case-control designs
144 in which controls are sampled by the set of subjects event-free at the time of event of the
145 case (20-22). Sampling probabilities for controls were derived by Samuelsen (23), while
146 for cases they were equal to 1 when the second phase sample size n was at least twice the
147 total number of event in the entire cohort ($\sum_{i=1}^N \Delta_i$) and $\pi_i = (n/2)/(\sum_{i=1}^N \Delta_i)$ otherwise.

148 (v) **Counter matching (CM)** is an alternative stratified version of the NCC. In this design, the
149 selection of controls is conducted by sampling from the set at risk in the opposite stratum

150 at the time of event of the case. Inclusion probabilities for controls within strata were
151 derived by Samuelsen (24) while for cases, the π_i was derived as in NCC design. As the
152 aim is to maximize the “discordance” of exposure within case-controls sets (25-27), the
153 variables used to define strata must be a proxy for the variables of interest, thus we used
154 only the surrogate variable X_{Surr} as strata for this design.

155 **Figure 2** illustrates an example of each sampling design method described above.
156 Specifically, in the upper part of the figure we displayed PPS and CC considering a
157 stratification for a binary variable; in the lower part NCC and CM designs are displayed. By
158 note we considered a 1:1 matching ratio between cases and controls in NCC and CM designs.

159

160 *Insert Figure 2*

161

162 **2.3 Evaluation of Biomarker impact on the event**

163 The following Cox model was applied to assess the influence of the biomarker on the event
164 adjusting for the confounder variable X_{Conf} (following the minimal set of adjustment
165 suggested in **Figure 1**):

$$166 \quad h_i(t) = h_0(t) \exp(\beta_{BM}X_{BM_i} + \beta_{Conf}X_{Conf_i}) \quad (1)$$

167

168 where β_{BM} and β_{Conf} represent the regression coefficients of the biomarker and of the
169 confounder, respectively. Given the availability of the biomarker only for the sub-cohort
170 (phase II), we applied a two-phase Cox model, in which regression coefficients are estimated
171 by maximizing the partial likelihood weighted by the inverse of the empirical inclusion
172 probability ($w_i = 1/\pi_i$) that accounts for the specific sampling design (6, 28, 29). In SRS,
173 CC and PPS designs (17, 30) empirical inclusion probabilities (π_i) were calculated using a

174 standard approach implemented in the twophase function in the `survey` package. Instead,
175 π_i 's were calculated following Samuelsen (23) for NCC and following Rivera for CM (25).
176 As surrogate variables are rarely available for new biomarkers at the design stage, we
177 considered also a situation in which the surrogate variable was identified only at the analysis
178 stage. Thus we ran a Cox model post-stratifying for the surrogate variable to take advantage
179 of it in the analysis stage (8).

180

181 **2.4 Simulations parameters**

182 The performance of the different designs was investigated through simulations. The number
183 of simulated data-set was set at $B=2000$ assuming a level of accuracy equal to 0.0046 and a
184 variance of the X_{BM} equal to 0.011 with a 5% significance level (31). For each scenario we
185 drew $B=2000$ random first-phase samples of $N=2000$ subjects to generate the hypothetical
186 cohort described above. We considered a common and rare marker with frequency in the
187 entire cohort of nearly 25% and 5%, respectively. In order to cover different levels of
188 accuracy of the surrogate in “predicting” the value of the biomarker, we varied the simulation
189 parameter to get different specificity and sensitivity.

190 The time-to-event end-point was generated (31, 32) from a Weibull hazard model as $T =$
191 $(-\log U / \lambda \exp(\beta' X))^{1/p}$, where $p=0.9$, $\lambda=0.1$, with U following a uniform distribution on the
192 interval from 0 to 1 and with the matrix of covariates X including the biomarker value (X_{BM}),
193 the risk factor ($X_{Risk\ Factor}$), and the confounder (X_{Conf}). The regression coefficients (β)
194 were inspired by the observed values in ALL data (15, 33). We started by simulating the
195 confounder variable as a dichotomous variable with $P(X_{Conf} = 1) = 0.5$; the biomarker was
196 simulated by a binomial distribution with $P(X_{BM} = 1 | X_{Conf}) = \exp(a + b * X_{Conf}) / (1 +$
197 $\exp(a + b * X_{Conf}))$ resulting in a frequency in the entire cohort of nearly 25% ($a=-2$ and $b=1.7$)
198 and 5% ($a=-4$ and $b=1.5$) for common and rare biomarker, respectively. The surrogate or

199 auxiliary variable, using X_{BM} as gold-standard, was simulated as $P(X_{Surr} = 1|X_{BM}) =$
 200 $\exp(c + d * X_{BM}) / (1 + \exp(c + d * X_{BM}))$. In order to cover different levels of accuracy of the
 201 surrogate in “predicting” the value of the biomarker, we set different values of parameters c
 202 and d . For example, we set $c=-0.87$ and $d=1.74$, resulting in specificity, $P(X_{Surr} = 0|X_{BM} =$
 203 $0)$, and sensitivity, $P(X_{Surr} = 1|X_{BM} = 1)$, values equal to 70%; $c= -0.9$ and $d=2.6$ resulting in
 204 specificity and sensitivity values equal to 70% and 80%, respectively; $c=-1.1$ and $d=3.3$
 205 resulting in specificity and sensitivity values equal to 70% and 90%, respectively. Finally, an
 206 additional binary risk factor $X_{Risk\ factor}$ was generated with a probability of $P(X_{Risk\ factor} =$
 207 $1) = 0.4$. Details of all specific parameters were reported in the Additional file Table S1.
 208 Censoring was generated with an exponential function with different rates $\rho = 0, 0.1, 0.4$,
 209 resulting in 0%, 15% and 50% of censoring at the end of follow-up τ . Minimum between
 210 time-to-event T_i and censoring C_i ($Z_i = \min(T_i, C_i)$) was calculated, with $\Delta_i = I(T_i < C_i)$.
 211 Administrative censoring was set at $\tau=2$. An average of 500 events for each phase I dataset
 212 was reached at the end of follow-up.
 213 To compare the performance of different sampling designs we sampled from each one of the
 214 B phase I data, a phase II sample with size n , as described in the chapter “**Second-phase**
 215 **sample**”.
 216 Information on X_{BM} was disregarded for subjects not included in the phase II sample and a
 217 weighted Cox model was applied to estimate β_{BM} as described in **Evaluation of Biomarker**
 218 **impact on the event** section.
 219 The performance of the estimate of β_{BM} over the B simulation has been assessed by the
 220 following measures (31):
 221 (i) Absolute bias, given by $= \bar{\hat{\beta}}_{BM} - \beta_{BM}$, where $\bar{\hat{\beta}}_{BM} = \sum_{i=1}^B \hat{\beta}_{iBM} / B$,
 222 (ii) $SE(\hat{\beta}_{BM})$, the empirical Standard Error (SE) of β_{BM} over all simulations,

- 223 (iii) Design effect, defined as the ratio between the estimated variance of β_{BM} in each
224 sampling design by the one in SRS (34),
- 225 (iv) Mean Square Error, MSE, given by $(\bar{\hat{\beta}}_{BM} - \beta_{BM})^2 + (SE(\hat{\beta}_{BM}))^2$,
- 226 (v) Coverage of the 95% confidence interval (CI) of β_{BM} and 95%CI length,
- 227 (vi) Power, number of times in which the null hypothesis ($\beta_{BM} = 0$) was rejected by the
228 Wald test at 5% significance level in the two-phase Cox regression model.

229 All analyses were performed using R software (version 3.5.2) (35).

230

231 3. RESULTS

232 3.1 Design comparison

233 General results considering both common (~25% frequency) and rare biomarker (~5%
234 frequency) are shown in **Table 1a-b**, respectively, under three censoring schemes (absent,
235 low and high). Overall, the simulations showed that the β_{BM} was estimated without any
236 noticeable bias for all designs. The standardized bias was always lower than 5% and the
237 distributions of $\hat{\beta}_{BM}$ were symmetric for all sampling designs (Additional file Figure S1).

238 As shown in **Table 1a**, the PPS did not show much advantage respect to the SRS design. The
239 empirical Standard Error of SRS and PPS were about the same, indicating no gain in
240 efficiency. We found a small but not relevant increase of power in PPS stratified by the
241 surrogate (2c in **Table 1a**) compared with traditional PPS.

242 On the other hand, the CC design improved power as compared with SRS reducing MSE and
243 empirical Standard Error (for each scenario), with a further advantage when also the surrogate
244 variable was used for stratification (3c in **Table 1a**). The stratification for risk factor and
245 confounder (3a and 3b in **Table 1a**) showed a slight loss of efficiency with respect to the
246 classical CC (3 in **Table 1a**).

247 When matching on time in the NCC and CM designs (4 and 5 in **Table 1**), a decrease of Mean
248 Square Error and empirical Standard Error (increase in design effect) were obtained if
249 compared with CC, PPS, and SRS in the presence of a censoring rate of $\rho=0.4$. CM presented
250 higher design effect and shorter confidence intervals with respect to CC stratified by the
251 surrogate for any censoring rate. Among all the considered designs, the CC stratified by the
252 surrogate was showing the highest power. By note, in NCC and CM design, the final sample
253 size of the second phase was slightly lower than the fixed one (i.e. $n=600$, see **Table 1**) as
254 resampling of controls is possible in these designs.

255 Similar performance results were obtained when a rare exposure ($\sim 5\%$) was considered
256 (**Table 1b**). In general, with a rare exposure, performance of NCC and CM, in term of
257 absolute bias, design effect and length of 95%CI, had an improvement with increasing
258 censoring rates. The estimate of regression coefficient ($\hat{\beta}_{BM}$), length and coverage of its 95%
259 confidence interval considering common and rare biomarker are given as Additional file
260 Table S2.

261
262 *Insert Table 1*
263

264 As sensitivity analysis, we evaluated the performance of the different designs modifying
265 variables included in the weighted Cox model in (1). Results are presented in the Additional
266 file (Table S3) and are consistent with previous results.

267 Interestingly, when the weighted Cox model was adjusted also for the risk factor variable
268 (Table S3a), $X_{Risk\ Factor}$, there was an increase of power for all designs as compared with
269 results of **Table 1a**. On the contrary, when the Cox model was adjusted for all variables
270 available in our setting (i.e. confounder, X_{Conf} , risk factor $X_{Risk\ Factor}$ and surrogate, X_{Surr} ,
271 see Table S3b), power decreased.

272

273 **3.2 Impact of surrogate**

274 In this section, we explore the impact of the accuracy level of the surrogate variable in designs
275 that use the surrogate as a stratification variable. As expected, higher sensitivity increased
276 power and design effect in the CC design stratified by the surrogate and in the CM design
277 (**Figure 3**). Interestingly, also results on the performance of post stratification by the
278 surrogate variable (not used as strata in the design) showed an increase of power and design
279 effect with an increase of the surrogate accuracy with a slightly lower performance than the
280 CC design stratified by the surrogate.

281

Insert Figure 3

282 **3.3 Power evaluation**

283 In **Figure 4** we have explored the power by the size of the phase II sample in different
284 designs. CM and NCC were more powerful up to a second phase sample size of nearly 500
285 individuals (1/4 of the entire cohort). For larger sample sizes, CC stratified for surrogate was
286 the most powerful design. By note, both NCC and CM were sampled considering a 1:1 case-
287 control ratio and controls could be resampled, thus the sample size of phase II was
288 constrained to not exceed twice the number of events in the entire cohort.

289

Insert Figure 4

290

291 **3.4 Application on the real data**

292 The study that motivated our work was performed to evaluate the role of different genetic
293 polymorphisms on treatment failure due to relapse (14, 15) and used data from a large Italian
294 clinical trial (ClinicalTrials.gov identifier NCT00613457) (36). Clinical data and other
295 information were available for the whole trial cohort (first phase) of 1999 consecutive patients
296 newly diagnosed with childhood acute lymphoblastic leukemia between 2000 and 2006.

297 Biological samples were stored at diagnosis and were used to measure the genetic
298 polymorphism of interest (second phase). In the study of Franca et al. (15) the subsample on
299 which to measure the genetic polymorphism was chosen after classifying patients into six
300 strata according to the event of interest (relapse/no relapse) and a three-level risk group
301 stratification defined by prognostic features in the treatment protocol. Patients were sampled
302 at random without replacement from each stratum, according to an optimal sampling strategy
303 (37). In particular, the sampling fractions for each stratum were chosen proportionally to the
304 genetic variability reported within each of the strata to maximize the precision of the estimate
305 of the genotype effect on the outcome. Of note, this was possible only due to the availability
306 of pilot data on the genetic polymorphism of interest, that actually are not often available.
307 Overall, out of the 766 children for whom genotyping was required (approximately 1.5
308 controls for each case), the biomarker of interest (GST- θ) was obtained on 601 patients,
309 getting a hazard ratio (HR) of 1.34 (95%CI: 0.90-2.00). By breaking up the variance of the
310 coefficient of GST- θ into phase I and II contributions, we derived the efficiency of the design
311 with respect to the expected one in the full cohort (estimate of the minimum irreducible
312 uncertainty for the cohort) that resulted 54% by having genotyped 1/3 of the sample.
313 Interestingly, the efficiency we got was higher than the expected one in any of the CC designs
314 considered (see **Table 2**), as computed by simulations developed in this paper. Thus, the use
315 of pilot data for an optimal sampling strategy compensated the lack of a surrogate variable.
316 Power evaluation was not done in this study at the design stage, but according to our
317 simulations results, a sample size of n=601 subjects would have reached a power of 55% and
318 68% to detect an HR_{BM} of 1.3 and 1.5, respectively with a CC design. If CC stratified by
319 surrogate would have been considered, an increase of power would have been obtained (60%
320 and 71% respectively for an HR_{BM} of 1.3 and 1.5), but still not reaching a reasonable value
321 (i.e. 80%).

322

Insert Table 2

323 **4. DISCUSSION**

324 This work underlined the importance of a careful study design in retrospective studies
325 evaluating a new research question using available cohort data on which to measure
326 additional characteristics, such as a new biomarker. It showed the advantages we can get in
327 terms of efficiency and power by using available data and the importance of power evaluation
328 in order to avoid useless studies. We also provide a tool to compute power by simulations.
329 From the simulation results, we found that the weighted Cox model provided valid estimates
330 of biomarker effect and good coverage probabilities in the considered designs. The
331 availability of auxiliary/surrogate variables of the biomarker of interest in phase I, the amount
332 of censoring and the prevalence of the biomarker, together with power considerations could
333 help researchers to identify the most efficient design. As expected, the CC provided better
334 efficiency with respect to the SRS design, while PPS did not show much advantage (5). If
335 some covariates are expected to be associated with the new variable of interest, it is
336 advantageous to use them to define strata in a two-phase design, especially if they have a
337 good accuracy in predicting it and in the presence of biomarkers with low frequency. Of note,
338 simulation results showed that using these surrogate variables of the biomarker just in the
339 analysis stage (and not in the stratified design) is also improving efficiency and power.
340 Interestingly, if a variable is associated both with the new variable and with the event of
341 interest, such as the “confounder”, or just with the event of interest, such as the “risk factor”,
342 using it to define strata did not show any advantage in power. Nevertheless, the inclusion of
343 the “risk factor” in the Cox model on the outcome is beneficial.
344 In the presence of censoring, sampling designs matching on time (NCC and CM) have shown
345 higher performance in term of design effect than CC and CC stratified by the surrogate
346 designs, respectively. Similar results were found by Borgan and Olsen, that also suggested to

347 combine the simple and counter-matching designs (sampling some controls by simple random
348 sampling and others by stratified random sampling) (38). Higher design effect is not always
349 followed by an improvement in power as the last one depends also on the direction of bias
350 that actually is favouring the CC design (as shown in Additional file Table S2). Moreover,
351 matched designs are constrained to have a fixed integer number for the case/control ratio and
352 this could result as a disadvantage in some settings. In the absence of censoring, results
353 showed that CC is more powerful compared to the all other designs. Stoer and colleagues
354 have found similar results and have called this particular condition as “CC extreme” design
355 (39), as in this setting controls have the longest possible follow-up (subjects event-free at the
356 end of follow-up in the absence of censoring). We also found, similarly to (22, 26), that CM
357 has a marked efficiency advantage especially when the biomarker is rare, as surrogate
358 information help to sample subjects with the biomarker.

359 One limitation of our work is that we have considered only 1:1 matching ratio for NCC and
360 CC designs, but we did a fair evaluation by comparing the performance of different designs at
361 the same sample size. Moreover, to emulate the ALL data, we have considered only a
362 moderate effect of the biomarker on the event and we have assumed time-constant
363 coefficients. However the general indications coming from our work are consistent with
364 previous results and among different setting, as well as for different specification of the
365 outcome model, as explored with sensitivity analysis. Moreover, the code developed,
366 available at <https://github.com/Fgraziano?tab=repositories>, is helpful to investigate the power
367 of different sampling designs in various setting.

368 **CONCLUSIONS**

369 Summarizing, for efficient selection of the subcohort, we recommend the use of the
370 information available on the entire cohort as suggested in the flowchart of Additional file
371 Figure S2 in supporting information. If a surrogate variable of the biomarker of interest is

372 available, we suggest to use a case-control study stratified for the surrogate variable or a
373 counter-matching design. The latter choice should be driven by the rate of censoring: if
374 censoring is low we might opt for CC stratified, otherwise CM is more convenient. If the
375 surrogate variable is not available, we should consider using a CC or a NCC design depending
376 on the censoring rate. As NCC and CM designs are constrained by a fixed ratio between cases
377 and controls (1:1, 1:2 ...), the overall sampling fraction with respect to the number of cases
378 should be also considered together with power evaluation. In fact, if sufficient power would
379 be reached with a phase II sample size n of nearly all cases plus a number of controls equal to
380 1.5 the number of cases, a CC design would be more convenient with respect to the matched
381 designs. Moreover, we found that optimal sampling strategies using potentially available
382 pilot data are greatly efficient. Thus, optimal sampling strategies for survival data would be
383 very useful together with an user-friendly instrument to compute power and efficiency for
384 different sampling designs (also considering stratification variable) in the two-phase setting.

385

386

387 **List of abbreviations:** Acute Lymphoblastic Leukemia (ALL); Case-Control (CC), Counter-
388 matching (CM), Hazard Ratio (HR), Nested Case-Control (NCC), Simple Random Sample
389 (SRS), Probability Proportional to size (PPS)

390

391 **DECLARATIONS**

392 **ETHICS APPROVAL AND CONSENT TO PARTICIPATE**

393 Not applicable

394 **CONSENT FOR PUBLICATION**

395 Not applicable

396 **AVAILABILITY OF DATA AND MATERIALS**

397 The simulation codes that support the findings of this study are openly available at
398 <https://github.com/Fgraziano?tab=repositories>.

399 **COMPETING INTERESTS**

400 The authors declare that they have no competing interests

401 **FUNDING**

402 This work was supported by the Italian Ministry of Education, University and Research [grant
403 SIR-MIUR RBSI14LOVD, 2017-NAZ-0446 (PRIN 2017) and Department of Excellence
404 project PREMIA (PREcision MedIcine Approach: bringing biomarker research to clinic)].

405 The sponsor has no role in the design of the study and collection, analysis, and interpretation
406 of data and in writing the manuscript.

407 **AUTHORS' CONTRIBUTIONS**

408 All authors commented on the draft and the interpretation of the findings, read and approved
409 the final manuscript.

410 FG: drafting the article, data analysis and interpretation, critical revision of the article, final
411 approval of the version to be published.

412 PR: conception the work (PI), supervision of the data collection, data analysis and
413 interpretation, drafting the article, critical revision of the article, final approval of the version
414 to be published.

415 MGV: critical revision of the article, final approval of the version to be published.

416 **ACKNOWLEDGMENT**

417 We thank the Italian Ministry of Education, University and Research

418

419 **REFERENCES**

420 1. Kannel WB. Bishop lecture. Contribution of the Framingham Study to preventive
421 cardiology. J Am Coll Cardiol. 1990;15(1):206-11.

- 422 2. Buist AS. The US Lung Health Study. *Respirology*. 1997;2(4):303-7.
- 423 3. Evans A, Salomaa V, Kulathinal S, Asplund K, Cambien F, Ferrario M, et al.
424 MORGAM (an international pooling of cardiovascular cohorts). *Int J Epidemiol*.
425 2005;34(1):21-7.
- 426 4. Lee J-E. How Should Biobanks Collect Biosamples for Clinical Application? A 20-year
427 Biomarker-related Publication and Patent Trend Analysis. *Osong public health and research*
428 *perspectives*. 2018;9(3):105-11.
- 429 5. Borgan Ø. Cohort sampling in epidemiological studies. Preprint series Statistical
430 Research Report [http://urn](http://urn.nb.no/URN:NBN:no-23420) nb no/URN: NBN: no-23420. 2001.
- 431 6. Borgan Ø, Samuelsen SO. A review of cohort sampling designs for Cox's regression
432 model: Potentials in epidemiology. *Norsk Epidemiologi*. 2003;13(2).
- 433 7. Breslow NE, Lumley T, Ballantyne CM, Chambless LE, Kulich M. Using the whole
434 cohort in the analysis of case-cohort data. *Am J Epidemiol*. 2009;169(11):1398-405.
- 435 8. Lumley T. *Complex surveys: a guide to analysis using R*: John Wiley & Sons; 2011.
- 436 9. Langholz B. Use of cohort information in the design and analysis of case-control studies.
437 *Scandinavian Journal of Statistics*. 2007;34(1):120-36.
- 438 10. Langholz B, Thomas DC. Nested case-control and case-cohort methods of sampling
439 from a cohort: a critical comparison. *American journal of epidemiology*. 1990;131(1):169-76.
- 440 11. Cai J, Zeng D. Sample size/power calculation for case-cohort studies. *Biometrics*.
441 2004;60(4):1015-24.
- 442 12. Haneuse S, Saegusa T, Lumley T. *osDesign: An R Package for the Analysis, Evaluation,*
443 *and Design of Two-Phase and Case-Control Studies*. *J Stat Softw*. 2011;43(11).
- 444 13. Rivera-Rodriguez C, Spiegelman D, Haneuse S. On the analysis of two-phase designs
445 in cluster-correlated data settings. *Stat Med*. 2019;38(23):4611-24.

- 446 14. Rebora P, Valsecchi MG. Survival estimation in two-phase cohort studies with
447 application to biomarkers evaluation. *Stat Methods Med Res.* 2016;25(6):2895-908.
- 448 15. Franca R, Rebora P, Basso G, Biondi A, Cazzaniga G, Crovella S, et al. Glutathione S-
449 transferase homozygous deletions and relapse in childhood acute lymphoblastic leukemia: a
450 novel study design in a large Italian AIEOP cohort. *Pharmacogenomics.* 2012;13(16):1905-16.
- 451 16. Marubini E, Valsecchi MG. *Analysing survival data from clinical trials and*
452 *observational studies: John Wiley & Sons; 2004.*
- 453 17. Rosén B. On sampling with probability proportional to size. *Journal of Statistical*
454 *Planning and Inference.* 1997;62(2):159-91.
- 455 18. Vandembroucke JP, Pearce N. Case-control studies: basic concepts. *Int J Epidemiol.*
456 2012;41(5):1480-9.
- 457 19. Haneuse S, Rivera-Rodriguez C. On the analysis of case-control studies in cluster-
458 correlated data settings. *Epidemiology.* 2018;29(1):50-7.
- 459 20. Delcoigne B, Stoer NC, Reilly M. Valid and efficient subgroup analyses using nested
460 case-control data. *Int J Epidemiol.* 2018.
- 461 21. Ernster VL. Nested case-control studies. *Prev Med.* 1994;23(5):587-90.
- 462 22. Langholz B, Clayton D. Sampling strategies in nested case-control studies. *Environ*
463 *Health Perspect.* 1994;102 Suppl 8:47-51.
- 464 23. Samuelsen SO. A pseudolikelihood approach to analysis of nested case-control studies.
465 *Biometrika.* 1997;84(2):379-94.
- 466 24. Samuelsen SO, Ånestad H, Skrandal A. Stratified case-cohort analysis of general cohort
467 sampling designs. *Scandinavian journal of statistics.* 2007;34(1):103-19.
- 468 25. Rivera C, Lumley T. Using the whole cohort in the analysis of countermatched samples.
469 *Biometrics.* 2016;72(2):382-91.

- 470 26. Cologne JB, Sharp GB, Neriishi K, Verkasalo PK, Land CE, Nakachi K. Improving the
471 efficiency of nested case-control studies of interaction by selecting controls using counter
472 matching on exposure. *Int J Epidemiol.* 2004;33(3):485-92.
- 473 27. Steenland K, Deddens JA. Increased precision using countermatching in nested case-
474 control studies. *Epidemiology.* 1997;8(3):238-42.
- 475 28. Lin DY. On fitting Cox's proportional hazards models to survey data. *Biometrika.*
476 2000;87(1):37-47.
- 477 29. Binder DA. Fitting Cox's proportional hazards models from survey data. *Biometrika.*
478 1992;79(1):139-47.
- 479 30. Laitila T, Olofsson J. A two-phase sampling scheme and π ps designs. *Journal of*
480 *Statistical Planning and Inference.* 2011;141(5):1646-54.
- 481 31. Burton A, Altman DG, Royston P, Holder RL. The design of simulation studies in
482 medical statistics. *Stat Med.* 2006;25(24):4279-92.
- 483 32. Bender R, Augustin T, Blettner M. Generating survival times to simulate Cox
484 proportional hazards models. *Stat Med.* 2005;24(11):1713-23.
- 485 33. Reborá P, Antolini L, Glidden DV, Valsecchi MG. Crude incidence in two-phase
486 designs in the presence of competing risks. *BMC Med Res Methodol.* 2016;16:5.
- 487 34. Kish L. *Survey Sampling* Wiley. New York. 1965.
- 488 35. TeamR RC. *A Language and Environment for Statistical Computing.* 2015. R
489 Foundation for Statistical Computing, Vienna, Austria:[URL <https://www.R-project.org/>]
490 Accessed November. 2018;26.
- 491 36. Moricke A, Zimmermann M, Valsecchi MG, Stanulla M, Biondi A, Mann G, et al.
492 Dexamethasone vs prednisone in induction treatment of pediatric ALL: results of the
493 randomized trial AIEOP-BFM ALL 2000. *Blood.* 2016;127(17):2101-12.

494 37. Reilly M. Optimal sampling strategies for two-stage studies. *American journal of*
495 *epidemiology*. 1996;143(1):92-100.

496 38. Borgan O, Olsen EF. The Efficiency of Simple and Counter-matched Nested Case-
497 control Sampling. *Scandinavian journal of statistics*. 1999;26(4):493-509.

498 39. Støer N, Salim A, Bokenberger K, Karlsson I, Reilly M. Is the matched extreme case-
499 control design more powerful than the nested case-control design? *Statistical Methods in*
500 *Medical Research*.0(0):0962280218778624.

501

Table 1. Bias, empirical standard error, mean square error, power and design effect of the biomarker regression coefficient estimate ($\hat{\beta}_{BM}$) for the full cohort and different sampling designs. Accuracy of surrogate: sensitivity (i.e. probability of having a positive surrogate if the biomarker is positive)=0.7 and specificity (i.e. probability of having a negative surrogate if the biomarker is negative)=0.7, biomarker common (a) and rare (b)

| a) SAMPLING DESIGN | Stratification Variable | n | BIAS | | | SE EMPIRICAL | | | MSE | | | POWER (%) | | | DESIGN EFFECT | | |
|--------------------------|----------------------------|------|----------------|--------|--------|----------------|-------|-------|----------------|-------|-------|----------------|-----|-----|----------------|-------|-------|
| | | | Censoring Rate | | | Censoring Rate | | | Censoring Rate | | | Censoring Rate | | | Censoring Rate | | |
| | | | 0 | 0.1 | 0.4 | 0 | 0.1 | 0.4 | 0 | 0.1 | 0.4 | 0 | 0.1 | 0.4 | 0 | 0.1 | 0.4 |
| Full cohort | - | 2000 | 0.008 | -0.015 | 0.009 | 0.093 | 0.095 | 0.112 | 0.009 | 0.009 | 0.013 | 99 | 97 | 95 | - | - | - |
| 1. SRS | - | 600 | 0.004 | -0.013 | 0.006 | 0.182 | 0.187 | 0.206 | 0.033 | 0.035 | 0.042 | 64 | 58 | 53 | - | - | - |
| 2. PPS | Event | 599 | 0.007 | -0.015 | 0.007 | 0.173 | 0.180 | 0.199 | 0.029 | 0.033 | 0.039 | 65 | 58 | 54 | 1.003 | 1.003 | 1.005 |
| 2a. PPS | Event; Risk factor | 598 | 0.008 | -0.016 | 0.004 | 0.172 | 0.175 | 0.205 | 0.029 | 0.031 | 0.042 | 65 | 58 | 52 | 1.002 | 1.003 | 1.002 |
| 2b. PPS | Event; Confounder | 598 | 0.003 | -0.015 | 0.002 | 0.174 | 0.179 | 0.203 | 0.030 | 0.032 | 0.041 | 65 | 57 | 51 | 0.999 | 1.002 | 1.000 |
| 2c. PPS | Event; Surrogate | 598 | 0.007 | -0.013 | 0.013 | 0.161 | 0.171 | 0.190 | 0.026 | 0.029 | 0.036 | 69 | 64 | 57 | 1.106 | 1.129 | 1.104 |
| 3. CC | Event | 600 | 0.011 | -0.008 | 0.019 | 0.159 | 0.158 | 0.179 | 0.025 | 0.025 | 0.032 | 74 | 68 | 67 | 1.179 | 1.219 | 1.352 |
| 3a. CC | Event; Risk factor | 600 | 0.010 | -0.009 | 0.008 | 0.162 | 0.166 | 0.182 | 0.026 | 0.028 | 0.033 | 72 | 65 | 62 | 1.139 | 1.176 | 1.307 |
| 3b. CC | Event; Confounder | 600 | 0.012 | -0.015 | 0.010 | 0.162 | 0.161 | 0.175 | 0.026 | 0.026 | 0.031 | 73 | 65 | 66 | 1.182 | 1.187 | 1.354 |
| 3c. CC | Event; Surrogate | 600 | 0.008 | -0.016 | 0.012 | 0.148 | 0.153 | 0.170 | 0.022 | 0.024 | 0.029 | 76 | 71 | 69 | 1.334 | 1.363 | 1.495 |
| 4. NCC | Event | 550 | 0.008 | -0.018 | 0.014 | 0.169 | 0.165 | 0.175 | 0.029 | 0.028 | 0.031 | 68 | 63 | 67 | 1.066 | 1.144 | 1.378 |
| 5. CM | Event; Surrogate | 546 | -0.044 | -0.058 | -0.009 | 0.151 | 0.153 | 0.165 | 0.025 | 0.027 | 0.027 | 67 | 61 | 67 | 1.379 | 1.395 | 1.536 |
| b) SAMPLING DESIGN | Stratification Variable | n | BIAS | | | SE EMPIRICAL | | | MSE | | | POWER (%) | | | DESIGN EFFECT | | |
| | | | Censoring Rate | | | Censoring Rate | | | Censoring Rate | | | Censoring Rate | | | Censoring Rate | | |
| | | | 0 | 0.1 | 0.4 | 0 | 0.1 | 0.4 | 0 | 0.1 | 0.4 | 0 | 0.1 | 0.4 | 0 | 0.1 | 0.4 |
| Full cohort | - | 2000 | -0.028 | -0.004 | -0.023 | 0.193 | 0.199 | 0.225 | 0.038 | 0.039 | 0.052 | 52 | 53 | 41 | - | - | - |
| 1. SRS | - | 600 | -0.045 | -0.041 | -0.093 | 0.376 | 0.392 | 0.802 | 0.144 | 0.156 | 0.652 | 23 | 23 | 19 | - | - | - |
| 2 PPS | Event | 599 | -0.051 | -0.023 | -0.060 | 0.380 | 0.394 | 0.453 | 0.147 | 0.155 | 0.209 | 25 | 24 | 19 | 1.079 | 1.131 | 1.139 |
| 2a. PPS | Event; Risk factor | 598 | -0.048 | -0.039 | -0.081 | 0.372 | 0.393 | 0.721 | 0.141 | 0.156 | 0.526 | 22 | 24 | 21 | 1.085 | 1.109 | 1.145 |
| 2b. PPS | Event; Confounder | 598 | -0.057 | -0.033 | -0.059 | 0.381 | 0.393 | 0.453 | 0.148 | 0.156 | 0.209 | 23 | 23 | 20 | 1.077 | 1.104 | 1.140 |
| 2c. PPS | Event; Surrogate | 598 | -0.055 | -0.022 | -0.073 | 0.388 | 0.391 | 0.597 | 0.153 | 0.153 | 0.362 | 24 | 25 | 19 | 1.090 | 1.147 | 1.158 |
| 3. CC | Event | 600 | -0.003 | 0.021 | 0.003 | 0.332 | 0.339 | 0.368 | 0.110 | 0.116 | 0.135 | 24 | 26 | 22 | 1.257 | 1.317 | 1.509 |
| 3a. CC | Event; Risk factor | 600 | -0.011 | 0.015 | 0.006 | 0.345 | 0.357 | 0.384 | 0.119 | 0.127 | 0.148 | 23 | 24 | 21 | 1.193 | 1.283 | 1.418 |
| 3b. CC | Event; Confounder | 600 | -0.023 | 0.024 | 0.011 | 0.328 | 0.344 | 0.363 | 0.108 | 0.119 | 0.132 | 23 | 26 | 22 | 1.267 | 1.329 | 1.513 |
| 3c. CC | Event; Surrogate | 565 | -0.018 | 0.011 | -0.010 | 0.311 | 0.313 | 0.345 | 0.097 | 0.098 | 0.119 | 26 | 29 | 24 | 1.419 | 1.607 | 1.757 |
| 4. NCC | Event | 545 | -0.020 | 0.024 | -0.000 | 0.354 | 0.355 | 0.363 | 0.126 | 0.127 | 0.132 | 21 | 24 | 21 | 1.126 | 1.222 | 1.513 |
| 5. CM | Event; Surrogate | 529 | -0.057 | -0.041 | -0.028 | 0.311 | 0.315 | 0.341 | 0.099 | 0.101 | 0.117 | 26 | 27 | 25 | 1.541 | 1.679 | 1.823 |

Legend: SRS–Simple Random Sample; PPS- Probability Proportional to size; CC- Case-Control; NCC- Nested Case-Control; CM- Counter-matching

Table 2. Efficiency (refers to the full cohort), design effect (refers to Simple Random Sampling) and power for Case-Control (CC) designs with hypothetical hazard ratio of the biomarker of interest (HR_{BM}) of 1.3 and 1.5, biomarker common (25%), censoring rate $\rho=0.1$, type I error 0.05

| EFFICIENCY | | | |
|----------------------|---------------------|-----------------------------------|-------------------------------------|
| | Case-control | CC stratified by surrogate | CC stratified by risk factor |
| $HR_{BM} = 1.3$ | 38.91% | 43.06% | 34.47% |
| $HR_{BM} = 1.5$ | 36.26% | 38.51% | 32.73% |
| DESIGN EFFECT | | | |
| | Case-control | CC stratified by surrogate | CC stratified by risk factor |
| $HR_{BM} = 1.3$ | 1.23 | 1.37 | 1.20 |
| $HR_{BM} = 1.5$ | 1.22 | 1.36 | 1.18 |
| POWER | | | |
| | Case-control | CC stratified by surrogate | CC stratified by risk factor |
| $HR_{BM} = 1.3$ | 54.8% | 60.1% | 54.3% |
| $HR_{BM} = 1.5$ | 68.1% | 70.65% | 65.4% |

LIST OF FIGURES

Figure 1. Causal diagram where the variables in the boxes are connected each other through the black arrows, denoting association. The dashed line box indicates a variable measured only in the sub-cohort

Figure 2. Probability Proportional to size (PPS) and Case-Control (CC) sampling from phase I cohort are shown in the upper part (panel a), left and right, respectively. Dots represents individuals in the strata (case or control and strata=1 or strata=2). Arrows correspond to the sampling from phase I to phase II. The number of sampled individuals in each stratum (phase II) depends on the sampling design. Nested Case-Control (NCC) and Counter-matching (CM) sampling are shown below in the figure (panel b). The lines represent the follow-up over which individuals are observed and the solid lines represented the sampled subjects. Black dot symbol represents the occurrence of an event and the arrow indicated the corresponding sampled control. For NCC, sampling is conducted in the same stratum and for CM, cases are matched with controls from the opposite stratum

Figure 3. Power and design effect for different sensitivity levels (i.e. probability of having a positive surrogate if the biomarker is positive) of the surrogate variable. Scenario: specificity (i.e. probability of having a negative surrogate if the biomarker is negative) =0.7, censoring rate $\rho=0.1$, hazard ratio of biomarker =1.5 and sample size of second phase (n) =600. Legend: CC stra surr (Case-Control stratified by surrogate), CC post surr (Case-Control post stratified by surrogate), CC event (Case-Control), CM (Counter-Matching) and SRS (simple random sampling)

Figure 4. Power for different sample sizes of second phase (n). Scenario: censoring rate $\rho=0.4$, common biomarker (25%), hazard ratio of biomarker=1.5, sensitivity (i.e. probability of having a positive surrogate if the biomarker is positive) =0.7 and specificity (i.e. probability of having a negative surrogate if the biomarker is negative)=0.7. Legend: CC stra surr (Case-Control stratified by surrogate), CC post surr (Case-Control post stratified by surrogate), CC event (Case-Control), CM (Counter-Matching) and SRS (simple random sampling)

Figures

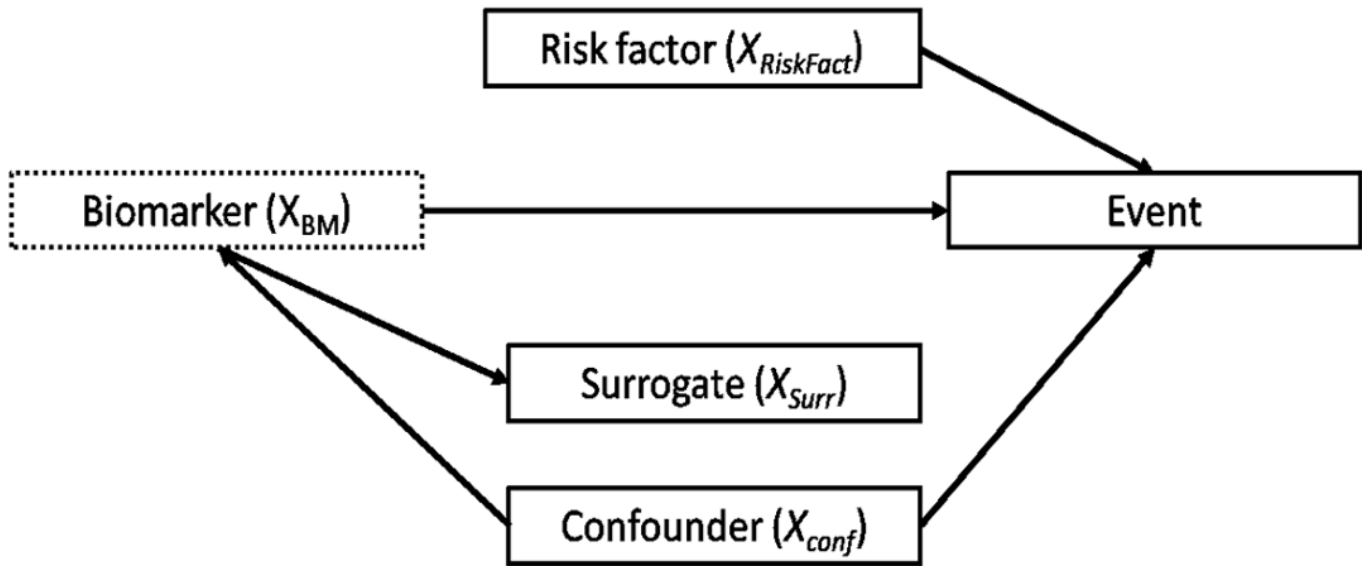


Figure 1

Causal diagram where the variables in the boxes are connected each other through the black arrows, denoting association. The dashed line box indicates a variable measured only in the sub-cohort

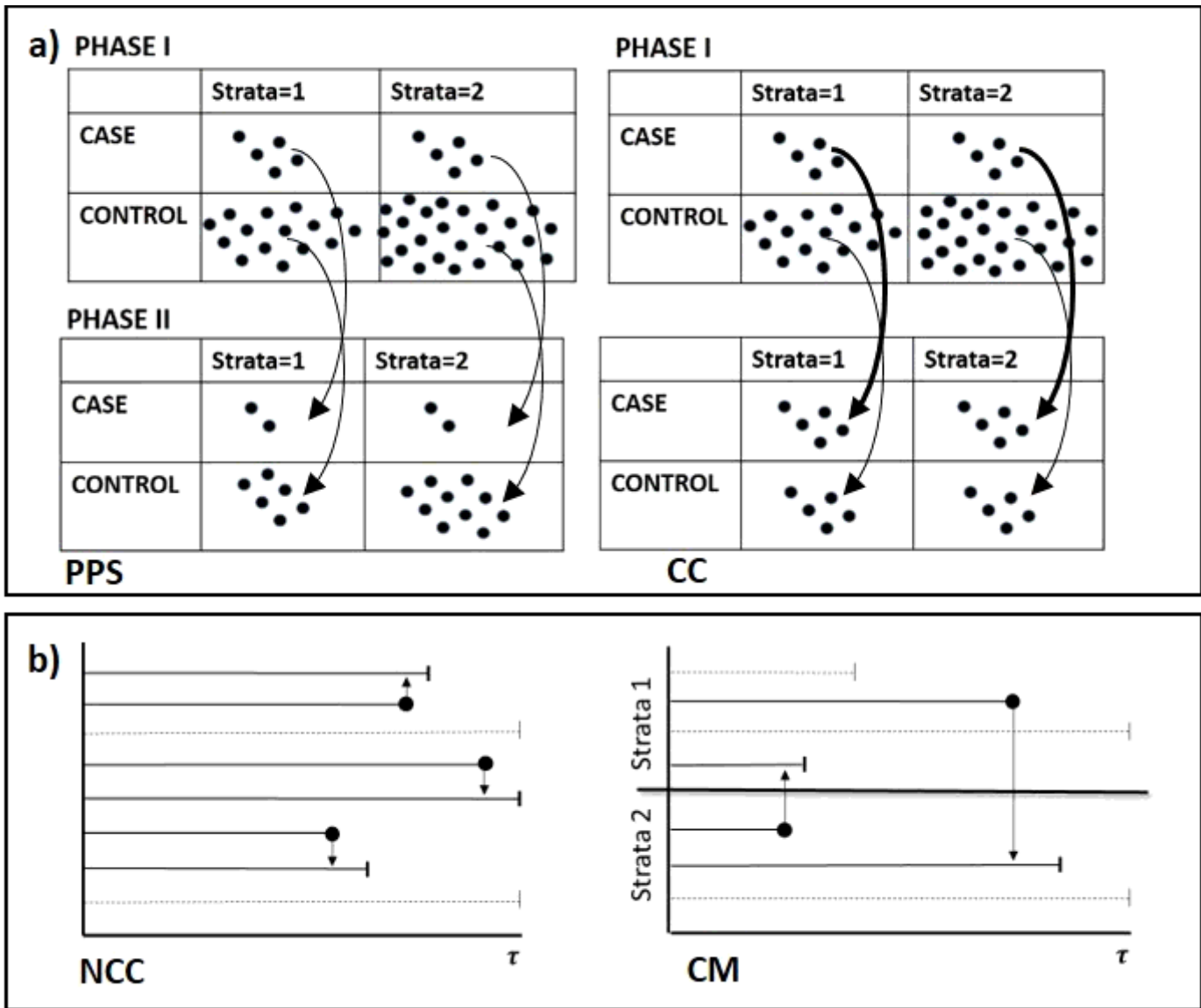


Figure 2

Probability Proportional to size (PPS) and Case-Control (CC) sampling from phase I cohort are shown in the upper part (panel a), left and right, respectively. Dots represent individuals in the strata (case or control and strata=1 or strata=2). Arrows correspond to the sampling from phase I to phase II. The number of sampled individuals in each stratum (phase II) depends on the sampling design. Nested Case-Control (NCC) and Counter-matching (CM) sampling are shown below in the figure (panel b). The lines represent the follow-up over which individuals are observed and the solid lines represent the sampled subjects. Black dot symbol represents the occurrence of an event and the arrow indicated the corresponding sampled control. For NCC, sampling is conducted in the same stratum and for CM, cases are matched with controls from the opposite stratum

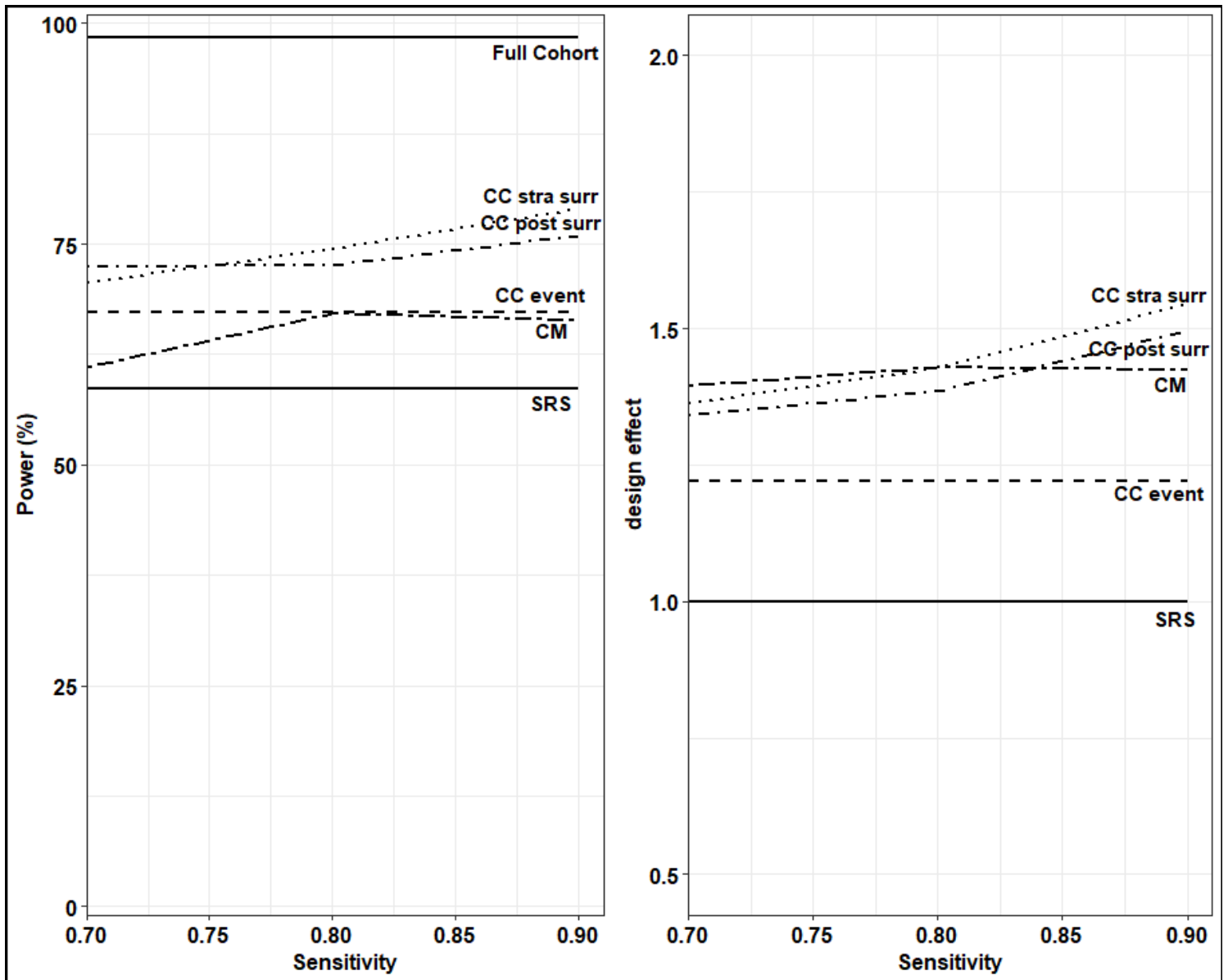


Figure 3

Power and design effect for different sensitivity levels (i.e. probability of having a positive surrogate if the biomarker is positive) of the surrogate variable. Scenario: specificity (i.e. probability of having a negative surrogate if the biomarker is negative) =0.7, censoring rate $\rho=0.1$, hazard ratio of biomarker =1.5 and sample size of second phase (n) =600. Legend: CC stra surr (Case-Control stratified by surrogate), CC post surr (Case-Control post stratified by surrogate), CC event (Case-Control), CM (Counter-Matching) and SRS (simple random sampling)

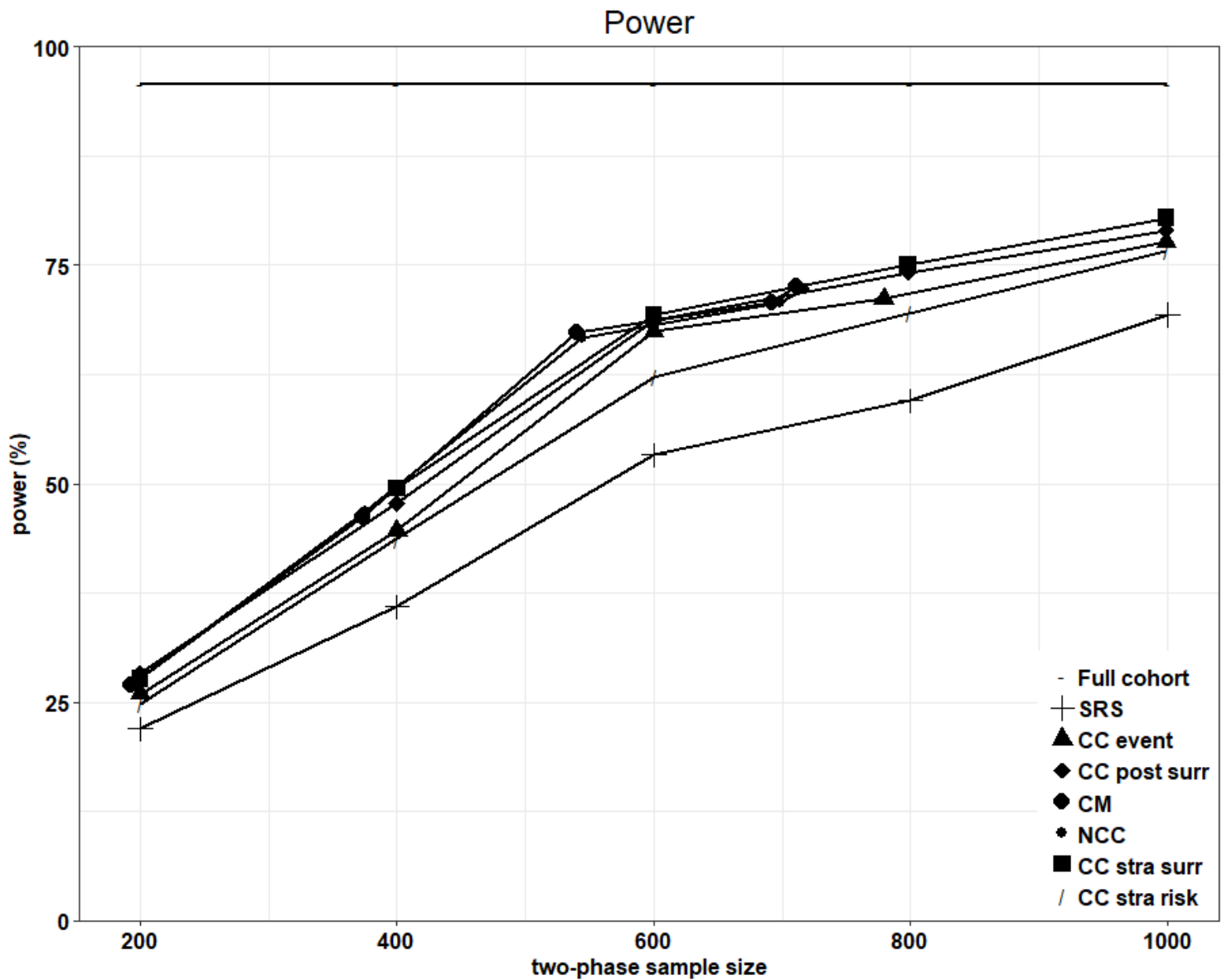


Figure 4

Power for different sample sizes of second phase (n). Scenario: censoring rate $\rho=0.4$, common biomarker (25%), hazard ratio of biomarker=1.5, sensitivity (i.e. probability of having a positive surrogate if the biomarker is positive) =0.7 and specificity (i.e. probability of having a negative surrogate if the biomarker is negative)=0.7. Legend: CC stra surr (Case-Control stratified by surrogate), CC post surr (Case-Control post stratified by surrogate), CC event (Case-Control), CM (Counter-Matching) and SRS (simple random sampling)

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [AdditionalFile.pdf](#)