

Fabrication of Nanoelectrospray Emitters for LC-MS

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Method Article

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Abstract

Ionization efficiency and consequently the sensitivity of mass spectrometric analyses strongly depend on the geometry of nanoelectrospray emitter used, particularly on the terminal inner diameter (ID) of the tip. This protocol has been optimized to produce emitters with a tip opening of 2-3 μm in diameter, from 50 μm ID fused silica capillaries. These emitters feature a porous silicate frit to alleviate the risk of microparticulate clogging of the narrow bore tip. As a result, they maintain stable spray performance while achieving ionization efficiencies as high as 20% without the need for specialized metal or polymer coatings. The overall time required is less than one hour.

Introduction

Ionization efficiency and consequently the sensitivity of mass spectrometric analyses strongly depend on the geometry of nanoelectrospray emitter used, particularly on the terminal inner diameter (ID) of the tip. This protocol has been optimized to produce emitters with a tip opening of 2-3 μm in diameter, from 50 μm ID fused silica capillaries. These emitters feature a porous silicate frit to alleviate the risk of microparticulate clogging of the narrow bore tip. As a result, they maintain stable spray performance while achieving ionization efficiencies as high as 20% without the need for specialized metal or polymer coatings. The overall time required is less than one hour.

Reagents

360/50 μm polyimide-coated fused silica capillary from Polymicro by Molex (# TSP050375) KASIL 1 (K silicate) from PQ Corporation Formamide from Sigma (# F9037-100) 1.5 ml tubes from Axygen (#MCT-175-L-C) Hydrofluoric acid from Sigma (#339261-100ML) Methanol MS grade water, formic acid and acetonitrile from Fisher (Optima)

Equipment

Laser capillary puller from Sutton (P-2000) Fused silica diamond cutter tool Gas burner Vortex Optical microscope Heating block Nanoscale liquid chromatograph

Procedure

1. Pre-warm the heating block to 95 $^{\circ}\text{C}$. Equilibrate the capillary puller laser for 10 minutes.
2. Cut the silica tubing in 10 cm pieces.
3. In a clean Axygen tube mix and immediately vortex 170 μL K silicate and 30 μL formamide.
4. Keeping the capillary almost horizontal, dip one end into the silicate solution until the liquid fills 2-3 cm of the tubing by capillary action (approximately 20 seconds). Gently blot the excess solution on the outer part of the tubing, if any.
5. Position the liquid-filled end of the capillary under the heating block and incubate for 1 hour at 95 $^{\circ}\text{C}$ to polymerize silicate.
6. Mount the capillary on the liquid chromatograph so that the solvent flows from the fritted end towards the open end of the capillary.

Flush the capillary with 50 μ L of 50% acetonitrile and 0.1% formic acid in water. 7. Visually check for polymerization (frit) at the edge. NOTE: Flushing the capillary with the same flow direction as in the final emitter will expel particulate deposited on the inner side of the frit before the tip is pulled, together with any unpolymerized silicate. 8. Using the gas burner, burn approximately 1 cm of the polyimide coating, 5 cm from the fritted end of the capillary. Let the capillary cool to room temperature and carefully wipe-off burnt polyimide debris using a Kimwipe soaked with methanol. CAUTION: stripped silica is extremely fragile. Hot silica may fracture if placed in contact with liquids before cooling down. 9. Mount the fritted capillary on the laser puller, with the inner edge of the fritted section 2 cm from the laser target (center of the mirror cover). 10. Pull the silica capillary using the following program: HEAT=250, FIL= , VEL=25, DEL=180, PULL=25 11. Using a microscope, check the tip geometry at 4X magnification and diameter at 20X magnification (Figure 1). 12. Optional: To reduce the surface of the tip, etch the tip by dipping the terminus in hydrofluoric acid for 10-15 seconds. Rinse in clean water. Etching may reduce the irregularities in the cross section of the tip, but results in slightly larger and less reproducible tip ID. Prolonged etching may damage the tip. 13. Cut the blunt end of the emitter to obtain a 1 mm frit.

Timing

1h

Anticipated Results

The procedure results in emitter tips with reproducible geometry.

References

Ficarro, S. et al. Improved electrospray ionization efficiency compensates for diminished chromatographic resolution and enables proteomics analysis of tyrosine signaling in embryonic stem cells. *Anal Chem.* 2009 May 1; 81(9):3440-7. Gibson, G.T. et al. Nanoelectrospray emitters: trends and perspective. *Mass Spectrom Rev.* 2009 Nov-Dec;28(6):918-36. Valaskovic, G.A. et al. Attomole protein characterization by capillary electrophoresis-mass spectrometry. *Science.* 1996 Aug 30;273(5279):1199-202.

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Figures

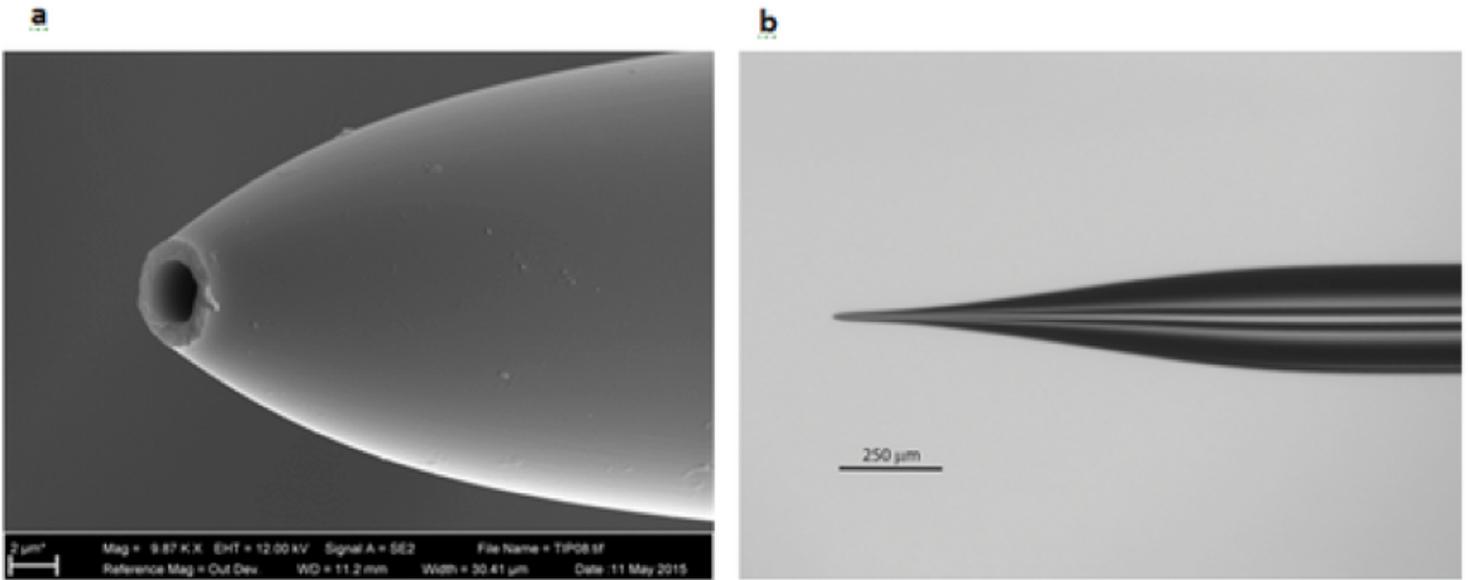


Figure 1

Figure1 Resulting tip geometry Typical emitter tip at a) 9870X (SEM scan) and b) 4X (optical transmission microscope)