

In vitro human Th17 development

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Method Article

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Abstract

Introduction

This protocol describes the isolation of naive human T cells and culture conditions that result in the development of IL-17 producing helper Th17 cells. The addition of IL-23 or IL-1 is essential for this process as is T cell receptor activation.

Reagents

-Recombinant hIL-1 β (50 ng/ml), rhIL-4 (10 ng/ml), rhIL-12 (5 ng/ml), rhIL-6 (30 ng/ml), rhTGF β (10 ng/ml), and rhIL-2 (100 U/ml) were purchased from R&D Systems. Recombinant hIL-23 was generated in house. -Medium : Yssel's medium with 1% human serum AB was from Gemini Bio-Products -Human CD4+ T cell isolation kit II (#130-091-155), human CD45RO microbeads (#130-046-001), and human T cell activation/expansion kit (#130-091-441) were from Miltenyi Biotec.

Equipment

AutoMACS, Milteniy Biotec

Procedure

1. Human PBMC were isolated from buffy coats of healthy donors by Ficoll hypaque density centrifugation.
2. Total CD4+ T cells were isolated using the CD4+ T cell Isolation Kit II from Miltenyi Biotec: - Total PBMC were resuspended in 40 μ l of MACS buffer (PBS 0.5% BSA 2mM EDTA) per 10 cells, and incubated with 10 μ l of Biotin-Antibody cocktail per 10 cells for 10 min at 4°C. - Subsequently, 30 μ l of MACS buffer per 10 cells was added, and cells were incubated with 20 μ l of Anti-Biotin Microbeads per 10 cells for 15 min at 4°C. - Cells were washed with MACS buffer and resuspended up to 10 cells in 500 μ l of MACS buffer. - CD4+ T cells were sorted by two rounds of magnetic depletion on the AutoMACS (program Deplete S).
3. CD4+ T cells (negative fraction) were washed with MACS buffer and resuspended in 80 μ l of MACS buffer per 10 cells. Then, 20 μ l of CD45RO microbeads per 10 cells was added for 15 min at 4°C. Naïve CD45RO- T cells were isolated by two rounds of depletion on the AutoMACS (as in step 2.).
4. Naïve CD4+ CD45RO- T cells were cultured at 1 \times 10 cells/well in 96-well flat-bottom plates or 5 \times 10 cells/well in 24-well plates (Falcon) for 5-6 days in Yssel's medium containing 1% human AB serum (Gemini Bio Products) with the appropriate cytokines and beads coated with anti-CD3, anti-CD28, and anti-CD2 antibodies (T Cell Activation/Expansion Kit, Miltenyi Biotec) (1 bead/10 cells).
5. After 5-6 days, cells were collected, counted, washed and cultured at 1 \times 10 cells/well in 96-well flat-bottom plates or 5 \times 10 cells/well in 24-well plates for an additional period of 5-6 days in the presence of the indicated cytokines and IL-2.
6. After 10-12 days of culture, cells were counted and stimulated at 5 \times 10 cells/ml with anti-CD3, anti-CD28, and anti-CD2 antibodies in the presence of IL-2 for 24 h to analyze gene expression or 48 h to assess cytokine level in cell-free supernatants.

Timing

14 days